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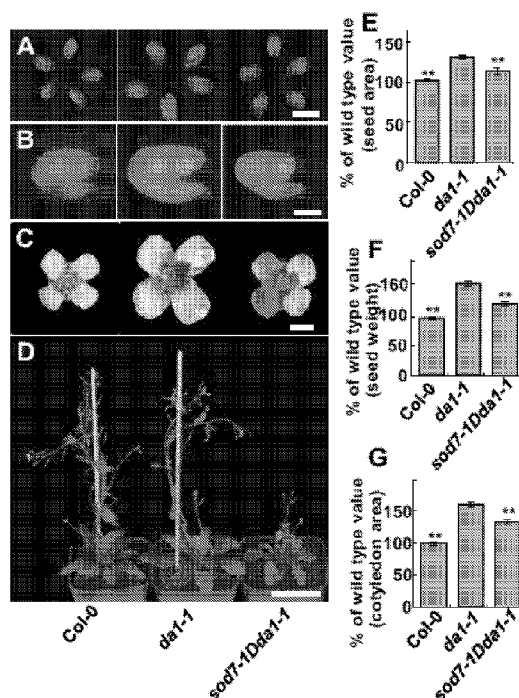
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(54) Title: PLANTS WITH INCREASED SEED SIZE

Figure 1



(57) Abstract: The invention relates to genetically modified plants with an altered seed phenotype, in particular increased seed size. The invention relates to a plant that does not produce a functional NGAL2 polypeptide or functional NGAL2 and NGAL3 polypeptides. NGAL2 and NGAL3 are members of the RAV family and comprise a B3 DNA-binding domain and a transcriptional repression motif.

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PLANTS WITH INCREASED SEED SIZE

Field of the invention

5 The invention relates to transgenic plants with improved growth and yield-related traits, in particular increased seed size. Also within the scope of the invention are related methods, uses, isolated nucleic acids and vector constructs.

Introduction

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The ever-increasing world population and the dwindling supply of arable land available for agriculture fuels research towards increasing the efficiency of agriculture and providing food security. Conventional means for crop and horticultural improvements utilise selective breeding techniques to identify plants having desirable characteristics.

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However, such selective breeding techniques have several drawbacks, namely that these techniques are typically labour intensive and result in plants that often contain heterogeneous genetic components that may not always result in the desirable trait being passed on from parent plants. Advances in molecular biology have allowed mankind to modify the germplasm of animals and plants. Genetic engineering of plants entails the isolation and manipulation of genetic material (typically in the form of DNA or RNA) and the subsequent introduction of that genetic material into a plant. Such technology has the capacity to deliver crops or plants having various improved economic, agronomic or horticultural traits, including increased yield. There are a number of methods that can be used, for example genome editing (using CRISPR or

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A trait of particular economic interest is increased seed size. Seed size is an important agronomic trait which increased crop yield, and is also a key ecological trait that influences many aspects of a species' regeneration strategy, such as seedling survival rates and seed dispersal syndrome (Harper et al., 1970; Westoby et al., 2002; Moles et al., 2005; Fan et al., 2006; Orsi and Tanksley, 2009; Gegas et al., 2010). Although the size of seeds is one of the most important agronomic traits in plants, the genetic and molecular mechanisms that set the final size of seeds are almost unknown. In higher plants, seed development starts with a double fertilization process, in which one of the

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while the other sperm nucleus fuses with the diploid central cell to form the triploid endosperm (Lopes and Larkins, 1993). The integuments surrounding the ovule are maternal tissues and form the seed coat after fertilization. Therefore, the size of the seed is the result of the growth of the embryo, the endosperm and the maternal tissues. However, the genetic and molecular mechanisms setting the limits of seed growth are almost unknown in plants.

Several factors that function maternally to regulate seed size have been identified in Arabidopsis. For example, TRANSPARENT TESTA GLABRA 2 (TTG2) influences seed growth by increasing cell elongation in the maternal integuments (Garcia et al., 2005; Ohto et al., 2009), while APETALA2 (AP2) may control seed growth by limiting cell elongation in the maternal integuments (Jofuku et al., 2005; Ohto et al., 2005; Ohto et al., 2009). By contrast, AUXIN RESPONSE FACTOR 2 (ARF2) acts maternally to control seed growth by restricting cell proliferation (Schruff et al., 2006). Similarly, the ubiquitin receptor DA1 acts synergistically with the E3 ubiquitin ligases DA2 and EOD1/BB to control seed size by limiting cell proliferation in the maternal integuments (Li et al., 2008; Xia et al., 2013). Mutations in the suppressor of *da1-1* (SOD2), which encodes the ubiquitin-specific protease (UBP15), suppress the large seed phenotype of *da1-1* (Du et al., 2014). DA1 physically associates with UBP15/SOD2 and modulates the stability of UBP15. These studies show that the ubiquitin pathway plays an important part in the maternal control of seed size. KLU/CYTOCHROME P450 78A5 (CYP78A5) regulates seed size by increasing cell proliferation in the maternal integuments of ovules (Adamski et al., 2009). KLU has also been suggested to generate mobile plant-growth substances that promote cell proliferation (Anastasiou et al., 2007; Adamski et al., 2009). By contrast, overexpression of *CYP78A6/EOD3* increases both cell proliferation and cell elongation in the integuments, resulting in large seeds (Fang et al., 2012). Seed size is also determined by zygotic tissues. Several factors have been described to influence seed size via the zygotic tissues in Arabidopsis, including HAIKU1(IKU1), IKU2, MINISEED3 (MINI3) and SHORT HYPOCOTYL UNDER BLUE1 (SHB1) (Garcia et al., 2003; Luo et al., 2005; Zhou et al., 2009; Wang et al., 2010; Kang et al., 2013). *iku* and *mini3* mutants form small seeds due to precocious cellularization of the endosperm (Garcia et al., 2003; Luo et al., 2005; Wang et al., 2010). SHB1 associates with MINI3 and IKU2 promoters and regulates expression of *MINI3* and *IKU2* (Zhou et al., 2009; Kang et al., 2013). ABA INSENSITIVE5 (ABI5) has been recently described to repress the expression of *SHB1* (Cheng et al., 2014), and MINI3 has been reported to activate expression of the

cytokinin oxidase (CKX2) (Li et al., 2013), suggesting the roles of phytohormones in regulating endosperm growth. In addition, the endosperm growth is influenced by parent of-origin effects (Scott et al., 1998; Xiao et al., 2006).

- 5 The invention is aimed at providing plants with improved yield traits that are beneficial to agriculture.

Summary of the invention

- 10 In a first aspect, the invention relates to a plant generated that does not produce a functional NGAL2 polypeptide or does not produce functional NGAL2 and NGAL3 polypeptides.

- In another aspect, the invention relates to a method for altering a plant phenotype comprising reducing or abolishing the expression of a nucleic acid sequence encoding a NGAL2 polypeptide or reducing or abolishing the activity of a NGAL2 or reducing or
15 abolishing the expression of a nucleic acid sequences encoding NGAL2 and NGAL3 polypeptides or reducing or abolishing the activity of a NGAL2 and NGAL3 polypeptide relative to a control plant.

- In another aspect, the invention relates to a method for making a plant with an altered
20 phenotype comprising reducing or abolishing the expression of a nucleic acid sequence encoding a NGAL2 polypeptide or reducing or abolishing the activity of a NGAL2 or reducing or abolishing the expression of a nucleic acid sequences encoding NGAL2 and NGAL3 polypeptides or reducing or abolishing the activity of a NGAL2 and NGAL3 polypeptide relative to a control plant..

- 25 In another aspect, the invention relates to a plant obtained or obtainable any method described above.

In another aspect, the invention relates to an isolated nucleic acid comprising a sequence comprising or consisting of SEQ ID NO: 1 or 2 or a functional variant or homologue thereof.

- 30 In another aspect, the invention relates to a vector comprising an isolated nucleic acid described above.

In another aspect, the invention relates to a silencing nucleic acid construct targeting sequence comprising or consisting of SEQ ID NO: 1, 2 or 3 or a functional variant, part or homologue thereof.

35 Figures

The invention is further described in the following non-limiting figures.

Figure 1. Isolation of a suppressor of *da1-1* (*sod7-1D*).

(A) Seeds from wild-type, *da 1-1* and *sod7-1D da1-1* plants (from left to right). (B) Mature embryos of the wild type, *da 1-1* and *sod7-1D da1-1* (from left to right). (C) Flowers from wild-type, *da 1-1* and *sod7-1D da1-1* plants (from left to right). (D) 30-day-old plants of the wild type, *da 1-1* and *sod7-1D da1-1* (from left to right). (E) Projective area of wild-type, *da1-1* and *sod7-1D da1-1* seeds. (F) Weight of wild-type, *da1-1* and *sod7-1D da 1-1* seeds. (G) Cotyledon area of 10-d-old wild-type, *da 1-1* and *sod7-1D da1-1* seedlings. Values (E-G) are given as mean \pm SD relative to the respective wild-type values, set at 100%. **, $P < 0.01$ compared with *da 1-1* (Student's t-test). Bars = 0.5 mm in (A), 0.2 mm in (B), 1 mm in (C) and 5 cm in (D).

Figure 2. Seed and organ size in the *sod7-1D* mutant.

(A and B) Seeds of Col-0 (A) and *sod7-1D* (B). (C and D) Mature embryos of Col-0 (C) and *sod7-1D* (D). (E and F) 10-day-old seedlings of Col-0 (E) and *sod7-1D* (F). (G) Projective area of Col-0 and *sod7-1D* seeds. (H) Weight of Col-0 and *sod7-1D* seeds. (I) Cotyledon area of 10-day-old Col-0 and *sod7-1D* seedlings. Values (G-I) are given as mean \pm SD relative to the respective wild-type values, set at 100%. **, $P < 0.01$ compared with the wild type (Student's t-test). Bars = 0.5 mm in (A) and (B), 0.2 mm in (C) and (D), and 1 mm in (E) and (F).

Figure 3. Cloning of the *SOD7* gene.

(A) Structure of the T-DNA insertion in the *sod7-1D* mutant. (B) Expression levels of At3g1 1580 (*SOD7*) and At3g1 1590 in *da 1-1* and *sod7-1D da1* seedlings. (C) The *SOD7* protein contains a B3 DNA binding domain (second domain in lighter shading) and a transcriptional repression motif (small light box in darker shading, marked with an arrow). (D) Projective area of Col-0, 35S:*GFP-SOD7#3* and 35S:*GFP-SOD7#5* seeds. (E) Cotyledon area of 10-day-old Col-0, 35S:*GFP-SOD7#3* and 35S:*GFP-SOD7#5* seedlings. (F) Expression levels of *SOD7* in Col-0, 35S:*GFP-SOD7#3* and 35S:*GFP-SOD7#5* seedlings. Values (D-F) are given as mean \pm SD relative to the respective wild-type values, set at 100%. **, $P < 0.01$ compared with the wild type (Student's t-test).

Figure 4. Expression pattern and subcellular localization of *SOD7*.

(A-K) *SOD7* expression activity was monitored by p*SOD7*:GUS transgene expression. Histochemical analysis of GUS activity in the developing leaves (A, B and C), the developing sepals (D, E), the developing petals (F, G), the developing stamens (H, I), and the developing carpels (J, K). (L) GFP fluorescence of *SOD7*-GFP in a young ovule of p*SOD7*:*SOD7*-GFP transgenic plants. (M-O) GFP fluorescence of *SOD7*-GFP (M),

DAPI staining (N), and merged (O) images are shown. Epidermal cells in *pSOD7:SOD7-GFP* leaves were used to observe GFP signal. (P-R) GFP fluorescence of GFP-SOD7 (P), DAPI staining (Q), and merged (R) images are shown. Epidermal cells in *35S:GFP-SOD7* leaves were used to observe GFP signal. Bars = 100 μ m in (A-K), 10 μ m in (L), and 2 μ m in (M-R).

Figure 5. SOD7 acts redundantly with NGAL3 to control seed size.

(A) The *SOD7* gene structure. The start codon (ATG) and the stop codon (TGA) are shown. Closed boxes indicate the coding sequence, and the line between boxes indicates intron. The T-DNA insertion site (*sod7-ko1*) in the *SOD7* gene was indicated. (B) The *NGAL3* gene structure. The start codon (ATG) and the stop codon (TGA) are shown. Closed boxes indicate the coding sequence, and the line between boxes indicates intron. The T-DNA insertion site (*ngal3-ko1*) in the *NGAL3* gene was indicated. (C) Seeds from Col-0, *sod7-ko1*, *ngal3-ko1* and *sod7-ko1 ngal3-ko1* plants (from left to right). (D) Mature embryos of Col-0, *sod7-ko1*, *ngal3-ko1* and *sod7-ko1 ngal3-ko1* (from left to right). (E) 25-day-old plants of Col-0, *sod7-ko1*, *ngal3-ko1* and *sod7-ko1 ngal3-ko1* (from left to right). (F) Flowers of Col-0, *sod7-ko1*, *ngal3-ko1* and *sod7-ko1 ngal3-ko1* (from left to right). (G) Projective area of Col-0, *sod7-ko1*, *ngal3-ko1* and *sod7-ko1 ngal3-ko1* seeds. (H) Weight of Col-0, *sod7-ko1*, *ngal3-ko1* and *sod7-ko1 ngal3-ko1* seeds. (I) Cotyledon area of Col-0, *sod7-ko1*, *ngal3-ko1* and *sod7-ko1 ngal3-ko1* seedlings. Values (G-I) are given as mean \pm SD relative to the respective wild-type values, set at 100%. **, $P < 0.01$ compared with the wild type (Col-0) (Student's t-test). Bars = 0.5 mm in (C), 0.2 mm in (D), 5 cm in (E), and 1 mm in (F).

Figure 6. SOD7 acts maternally to determine seed size.

(A) Projective area of Col-OxCol-0 (C/C) F₁, Col-0 \times *sod7-ko1 ngal3-ko1* (C/d) F₁, *sod7-ko1 ngal3-ko1* \times Col-0 (d/C) F₁ and *sod7-ko1 ngal3-ko1* (d/d) F₁ seeds. Values are given as mean \pm SD relative to the respective wild-type values, set at 100%. (B) Projective area of Col-OxCol-0 (C/C) F₂, Col-0 \times *sod7-ko1 ngal3-ko1* (C/d) F₂, *sod7-ko1 ngal3-ko1* \times Col-0 (d/C) F₂ and *sod7-ko1 ngal3-ko1* (d/d) F₂ seeds. Values are given as mean \pm SD relative to the respective wild-type values, set at 100%. (C and D) Mature ovules of Col-0 (C) and *sod7-ko1 ngal3-ko1* (D). (E) Outer integument length of mature Col-0 (lighter bar to the left) and *sod7-ko1 ngal3-ko1* (darker bar to the right) ovules. Values are given as mean \pm SD. (F) The number of cells in the outer integuments of Col-0 and *sod7-ko1 ngal3-ko1* at 0, 6 and 8 DAP. Values are given as mean \pm SD. (F) The length of cells in the outer integuments of Col-0 and *sod7-ko1 ngal3-ko1* at 0, 6 and 8 DAP. Values are given as

mean \pm SD. **, $P < 0.01$ compared with the wild type (Col-0) (Student's t-test). Bars = 50 μ m in (C) and (D).

Figure 7. *klu-4* is epistatic to *sod7-ko1 ngal3-ko1* with respect to seed size.

(A) Seed area of Col-0, *klu-4*, *sod7-ko1 ngal3-ko1* and *klu-4 sod7-ko1 ngal3-ko1* (from left to right). Values are given as mean \pm SD relative to the respective wild-type values, set at 100%. (B) Seed weight of Col-0, *klu-4*, *sod7-ko1 ngal3-ko1* and *klu-4 sod7-ko1 ngal3-ko1* (from left to right). Values are given as mean \pm SD relative to the respective wild-type values, set at 100%. (C) The outer integument length of Col-0, *klu-4*, *sod7-ko1 ngal3-ko1* and *klu-4 sod7-ko1 ngal3-ko1* (from left to right). *ngal3-ko1* at 0 and 8 DAP. Values are given as mean \pm SD. (D) The number of cells in the outer integuments of Col-0, *klu-4*, *sod7-ko1 ngal3-ko1* and *klu-4 sod7-ko1 ngal3-ko1* (from left to right) at 0 and 8 DAP. Values are given as mean \pm SD. **, $P < 0.01$ compared with their respective controls (Student's t-test).

Figure 8. SOD7 directly binds to the promoter of KLU and represses the expression of KLU.

(A) Expression dynamics of *SOD7* and *KLU* in *pER8-SOD7* transgenic plants treated with β -estradiol for 0, 4 and 8 hours. Means were calculated from three biological samples. Values are given as mean \pm SD. **, $P < 0.01$, compared with the expression level of *KLU* and *SOD7* at 0 hour, respectively (Student's t-test). (B) A 2-kb promoter region of *KLU* upstream of its ATG codon contains a CACTTG sequence. PF1 and PF2 represent PCR fragments used for ChIP-quantitative PCR analysis. A and A-m indicate the wild-type probe and the mutated probe used in the EMSA assay, respectively. (C) ChIP-qPCR analysis shows that *SOD7* binds to the promoter fragment PF1 of *KLU*. Chromatin from *35S:GFP* and *35S:GFP-SOD7* transgenic plants was immunoprecipitated by anti-GFP, and the enrichment of the fragments was determined by quantitative real-time PCR. The ACTIN7 promoter was used as a negative control. The fold enrichment was normalized to the ACTIN7 amplicon, set at 1. Means were calculated from three biological samples. Values are given as mean \pm SD. **, $P < 0.01$, compared with *35S:GFP* transgenic plants (Student's t-test). (D) Direct interaction between *SOD7* and the *KLU* promoter determined by EMSA. The biotin-labeled probe A and MBP-SOD7 formed the DNA-protein complex, but the mutated probe A-m and MBP-SOD7 did not form the DNA-protein complex. The retarded DNA-protein complex was reduced by competition using the unlabeled probe A.

Figure 9. The organ size phenotype of 35S:GFP-SOD7 transgenic plants.

Overexpression of *SOD7* results in small plants compared with the wild type. Bar = 5 cm.

Figure 10. Phylogenetic tree of the RAV family members in Arabidopsis.

Figure 11. SOD7 acts redundantly with NGAL3 to influence organ size.

Petal area of *Col-0*, *sod7-ko1*, *ngal3-ko1* and *sod7-ko1 ngal3-ko1*. (B) The seventh leaf area of *Col-0*, *sod7-ko1*, *ngal3-ko1* and *sod7-ko1 ngal3-ko1*. Values (A and B) are given as mean \pm SD relative to the respective wild-type values, set at 100%. **, $P < 0.01$ and *, $P < 0.05$ compared with the wild type (*Col-0*).

Figure 12: Conserved domains in NGAL2, NGAL3 and homologs. a) B box motif.
b) Repressor motif

Figure 13: Alignment of sequences. The following sequences are shown (from top to bottom): RMZM2G053008, HvMLOC_57250, Os12g0157000, GmLoc100778733, Bra004501, Bra000434, Bra040478, Bra014415, Bra003482, Bra007646, GmLoc100781489, GRMZM2G024948_T01, os02g0683500, HvMLOC_66387, os04g0581400, GRMZM2G102059_T01, Os10g0537100, GRMZM2G142999_T01, GRMZM2G125095_T01, os03g0120900, GRMZM2G098443_T01, GRMZM2G082227_T01, Os1 1g01 56000, GRMZM2G328742_T01, GmLoc100802734, GmLod 00795470, GmLod 0081 8164, Bra017262, At2g36080/NGAL1, Bra005301, At3g1 1580/SOD7, BraLOC1 03849927, Bra034828, At5g06250/NGAL3, Bra005886, GmLod 02660503, HvMLOC_38822, os01g0693400, HvMLOC44012, HvMLOC_7940, HvMLOC_75135, TRAECDM81004, HvMLOC_56567, TRAES3BF098300010CFD_t1, HvMLOC_63261, TRAES3BF062700040CFD_t1, TRAES3BF062600010CFD_t1, Bra038346, GmLoc732601, GmLod 00789009, GmLod 00776987, GmLod 00801 107. Conserved B3 domain and repressor motif are boxed.

Figure 14: Genome editing experiments to knock out rice genes Os1 1g01560000 and Os12g0157000 in rice. gRNA stands for guide RNA, target site linked with gRNA scaffold will recruit CAS9 enzyme to target site in the genome and cause gene-editing.

Detailed description

The present invention will now be further described. In the following passages, different aspects of the invention are defined in more detail. Each aspect so defined may be combined with any other aspect or aspects unless clearly indicated to the contrary. In particular, any feature indicated as being preferred or advantageous may be combined with any other feature or features indicated as being preferred or advantageous.

The practice of the present invention will employ, unless otherwise indicated, conventional techniques of botany, microbiology, tissue culture, molecular biology,

chemistry, biochemistry and recombinant DNA technology, bioinformatics which are within the skill of the art. Such techniques are explained fully in the literature.

As used herein, the words "nucleic acid", "nucleic acid sequence", "nucleotide",
5 "nucleic acid molecule" or "polynucleotide" are intended to include DNA molecules (e.g., cDNA or genomic DNA), RNA molecules (e.g., mRNA), naturally occurring, mutated, synthetic DNA or RNA molecules, and analogues of the DNA or RNA generated using nucleotide analogues. It can be single-stranded or double-stranded. Such nucleic acids or polynucleotides include, but are not limited to, coding sequences
10 of structural genes, anti-sense sequences, and non-coding regulatory sequences that do not encode mRNAs or protein products. These terms also encompass a gene. The term "gene" or "gene sequence" is used broadly to refer to a DNA nucleic acid associated with a biological function. Thus, genes may include introns and exons as in the genomic sequence, or may comprise only a coding sequence as in cDNAs, and/or
15 may include cDNAs in combination with regulatory sequences.

The terms "peptide", "polypeptide" and "protein" are used interchangeably herein and refer to amino acids in a polymeric form of any length, linked together by peptide bonds.

For the purposes of the invention, "transgenic", "transgene" or "recombinant" means with regard to, for example, a nucleic acid sequence, an expression cassette, gene construct or a vector comprising the nucleic acid sequence or an organism transformed with the nucleic acid sequences, expression cassettes or vectors according to the
20 invention, all those constructions brought about by recombinant methods in which either

(a) the nucleic acid sequences encoding proteins useful in the methods of the invention, or

(b) genetic control sequence(s) which is operably linked with the nucleic acid
25 sequence according to the invention, for example a promoter, or

(c) both (a) and (b)

are not located in their natural genetic environment or have been modified by genetic intervention techniques, it being possible for the modification to take the form of, for example, a substitution, addition, deletion, inversion or insertion of one or more
30 nucleotide residues. The natural genetic environment is understood as meaning the natural genomic or chromosomal locus in the original plant or the presence in a

genomic library. In the case of a genomic library, the natural genetic environment of the nucleic acid sequence is preferably retained, at least in part. The environment flanks the nucleic acid sequence at least on one side and has a sequence length of at least 50 bp, preferably at least 500 bp, especially preferably at least 1000 bp, most preferably at least 5000 bp. A naturally occurring expression cassette - for example the naturally occurring combination of the natural promoter of the nucleic acid sequences with the corresponding nucleic acid sequence encoding a polypeptide useful in the methods of the present invention, as defined above - becomes a transgenic expression cassette when this expression cassette is modified by non-natural, synthetic ("artificial") methods such as, for example, mutagenic treatment. Suitable methods are described, for example, in US 5,565,350 or WO 00/15815 both incorporated by reference.

In certain embodiments, a transgenic plant for the purposes of the invention is thus understood as meaning, as above, that the nucleic acids used in the method of the invention are not at their natural locus in the genome of said plant, it being possible for the nucleic acids to be expressed homologously or heterologously. Thus, the plant can express a silencing construct transgene. However, as mentioned, in certain embodiments, transgenic also means that, while the nucleic acids according to the different embodiments of the invention are at their natural position in the genome of a plant, the sequence has been modified with regard to the natural sequence, and/or that the regulatory sequences of the natural sequences have been modified, for example by mutagenesis.

Transgenic is preferably understood as meaning the expression of the nucleic acids according to the invention at an unnatural locus in the genome, i.e. homologous or, preferably, heterologous expression of the nucleic acids takes place. According to the invention, the transgene is stably integrated into the plant and the plant is preferably homozygous for the transgene.

The various aspects of the invention use genetic engineering methods. Thus, the plants have been generated using genetic engineering methods, for example transgene expression, mutagenesis, gene targeting, gene silencing or genome editing as detailed below. Thus, the various aspects of the invention can involve recombinant DNA technology. The plants of the invention are thus mutant plants which have been genetically engineered, that is manipulated by human intervention. The plants of the

various aspects of the invention do not relate to natural variants which have not been manipulated by genetic engineering methods. The plant may be a transgenic plant in some embodiments, for example a plant which comprises a nucleic acid construct expressing a silencing construct.

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In preferred embodiments exclude embodiments that are solely based on generating plants by traditional breeding methods.

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The inventor has identified a B3 domain transcriptional repressor termed AtNGAL2, encoded by the suppressor of *Atda1-1* (*AtSOD7*), which acts maternally to control seed size by restricting cell proliferation in the integuments of ovules and developing seeds.

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The inventor previously identified the ubiquitin receptor DA1 as a negative regulator of seed size in Arabidopsis (Li et al., 2008). The *da1-1* mutant formed large seeds due to increased cell proliferation in the maternal integuments (Li et al., 2008; Xia et al., 2013). To identify novel components in the DA1 pathway or other seed size regulators, the inventor initiated a T-DNA activation tagging screen for modifiers of *da1-1* (Fang et al., 2012). A dominant suppressor of *da1-1* (*sod7-1D*) was isolated from seeds produced from approximate 16,000 T1 plants (Fig.1A). Seeds of the *sod7-1D da1-1* double mutant were significantly smaller and lighter than *da1-1* seeds (Figures 1A, E and F). The results show that the *sod7-1D* mutation suppressed the seed and organ size phenotypes of *da1-1*. The *SOD7* gene was isolated and found to encode a NGATHA like protein (NGAL2) containing a B3 DNA-binding domain and a transcriptional repression motif (Figure 3C) (Alvarez et al., 2009; Ikeda and Ohme-Takagi, 2009; Trigueros et al., 2009). *SOD7* belongs to the RAV gene family that consists of 13 members in Arabidopsis (Figure 10) (Swaminathan et al., 2008). Several members of the RAV family contain the putative transcriptional repression motifs, including NGA1, NGA2, NGA3, NGA4, NGAL1, NGAL2/SOD7 and NGAL3 (Figure 10) (Ikeda and Ohme-Takagi, 2009). The transcriptional repression motifs in NGA1, NGAL1 and NGAL2/SOD7 have been known to possess the repressive activity (Ikeda and Ohme-Takagi, 2009), indicating that they are transcriptional repressors. *SOD7* exhibits the highest similarity to Arabidopsis NGAL3/DEVELOPMENT-RELATED PcG TARGET IN THE APEX 4 (DPA4) (Figure 10), which has known roles in the regulation of leaf serrations (Engelhorn et al., 2012), but no previously identified function in seed size control.

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The inventor has shown that overexpression of *AtSOD7* significantly decreases seed size of wild-type plants, while the disruption of *AtSOD7* increases seed size. The inventors have shown that disruption of *AtNGAL3*, a close homolog of *AtSOD7* also increases seed size. Moreover, the simultaneous disruption of *AtSOD7* and *AtNGAL3* further increases seed size in a synergistic manner. Genetic analyses carried out by the inventor indicate that *AtSOD7* acts in a common pathway with the seed size regulator *AtKLU* to control seed growth, but does so independently of *AtDA1*. Further results show that *AtSOD7* directly binds to the promoter of *AtKLU* *in vitro* and *in vivo* and represses expression of *AtKLU*. Therefore, the inventor's findings show that *AtSOD7* (aka *AtNGAL2*) is a target for seed size improvement in crops. The plants of the invention are characterised by increased organ size, for example increased seed size, and also increased petal size, increased embryo size, for example. Increased seed size leads to an increase in seed yield and the plants of the invention are thus characterised by increased seed yield.

Thus, the invention relates to a plant wherein said plant does not produce a functional NGAL2 and/or NGAL3 polypeptide. For example, the plant does not produce a full length transcript of a nucleic acid sequence encoding a NGAL2 and/or NGAL3 protein. In another embodiment, the plant produces a full length transcript of a nucleic acid sequence encoding a NGAL2 and/or NGAL3, but the resulting protein is not functional. In a preferred embodiment, said plant does not produce a functional NGAL2 polypeptide and also does not produce a functional NGAL3 polypeptide. Such plants are double knock-out or knock-down mutants (loss of function mutants) and methods according to the invention as described below relate to making such double mutants.

The plants of the invention are mutant plants which have been genetically modified and are not naturally occurring varieties. Thus, the plants have been generated using genetic engineering methods, for example mutagenesis, gene targeting, gene silencing or genome editing as detailed below. Thus, the various aspects of the invention can involve recombinant DNA technology. The plant may be a transgenic plant in some embodiments, for example a plant which comprises a transgene to silence gene expression of *SOD7* and/or *NGAL3*. In other embodiments, the plant does not carry a transgene, but is a mutant plant wherein the endogenous nucleic acid sequence encoding a NGAL2 and/or NGAL3 polypeptide or the endogenous *SOD7* and/or *NGAL3* promoter sequence has been manipulated to either reduce or abolish expression of a nucleic acid sequence encoding a NGAL2 and/or NGAL3 polypeptide

or reduce or abolish the activity of a NGAL2 and/or NGAL3 polypeptide. The plants of the various aspects of the invention do not relate to natural variants which have not been manipulated by genetic engineering methods.

5 In one aspect, the invention relates to a plant generated by genetic engineering methods wherein the expression of a nucleic acid sequence encoding a NGAL2 and/or NGAL3 polypeptide and/or the activity of a NGAL2 and/or NGAL3 polypeptide is reduced or abolished relative to a control plant. In one embodiment, expression of a nucleic acid sequence encoding a NGAL2 polypeptide or the activity of a NGAL2
10 polypeptide is reduced or abolished. In another embodiment, expression of a nucleic acid sequence encoding a NGAL3 polypeptide or the activity of a NGAL3 polypeptide is reduced or abolished. In a preferred embodiment the presence of function of both proteins is affected, in other words, the plant is characterised in that expression of a nucleic acid sequence encoding a NGAL2 polypeptide or the activity of a NGAL2
15 polypeptide is reduced or abolished and also expression of a nucleic acid sequence encoding a NGAL3 polypeptide or the activity of a NGAL3 polypeptide is reduced or abolished in said plant.

For example, said plant can have reduced or abolished expression of a nucleic acid
20 sequence encoding a NGAL2 polypeptide and reduced or abolished expression of a nucleic acid sequence encoding a NGAL3 polypeptide. In another embodiment, said plant can have reduced or abolished activity of a NGAL2 polypeptide and reduced or abolished activity of a NGAL3 polypeptide. In another embodiment, said plant can have reduced or abolished expression of a nucleic acid sequence encoding a NGAL2
25 polypeptide and reduced or abolished activity of a NGAL3 polypeptide. In another embodiment, said plant can have reduced or abolished expression of a nucleic acid sequence encoding a NGAL3 polypeptide and reduced or abolished activity of a NGAL2 polypeptide.

30 A NGAL2 or NGAL3 polypeptide as described in the various aspects of the invention has a characteristic domain structure as explained below.

A NGAL2 OR NGLA3 polypeptide as described in the various aspects of the invention comprises a B3 DNA binding domain which has the structure shown in figure 12 .

35

In one embodiment, the domain is: SNNNNNNGGSGDDVACHFQRFDLHRLFIGWRGE

(SEQ ID NO:6) or a domain with at least 80%, at least 95% or at least 95% sequence identity thereto.

5 A NGAL2 OR NGAL3 polypeptide as described in the various aspects of the invention also comprises a transcriptional repression motif shown in figure 12.

In one embodiment, the domain is: VRLFGVNLE (SEQ ID NO:7) or a domain with at least 95% sequence identity thereto.

10 In one embodiment, the NGAL2 protein is AtNGAL2, a functional variant, part or homologue thereof. AtNGAL2 is encoded by AtSOD7. The term AtSOD7 refers to the wild type *AtSOD7* nucleic acid sequence comprising or consisting of SEQ ID NO. 1 (CDNA) or SEQ ID NO 2 (genomic DNA). The protein encoded by *AtSOD7* is termed *AtNGAL2* SEQ ID NO.3. In one embodiment, said functional homologue is not
15 AtNGAL3.

In one embodiment, the NGAL3 protein is AtNGAL3, a functional variant, part or homologue thereof. The term *AtNGAL3* refers to the wild type *AtNGAL3* nucleic acid sequence comprising or consisting of SEQ ID NO. 4. The protein encoded by *AtNGAL3*
20 is termed *AtNGAL3* SEQ ID NO.5.

The term "functional" refers to the biological function of the NGAL2 or NGAL3, that is their function in controlling organ size, in particular seed size. The terms "functional variant" or "functional part" as used herein, for example with reference to SEQ ID NOs:
25 1, 2 or 3, or SEQ ID NOs: 4 or 5 refers to a variant gene or polypeptide sequence or part of the gene or polypeptide sequence which retains the biological function of the full non-variant SOD7/NGAL2 or NGAL2/NGAL3 sequence, that is regulation of seed size. Such sequences complement the *Atsod7-1D* mutant or *Atngal3* mutant respectively.

30 Thus, it is understood, as those skilled in the art will appreciate, that the aspects of the invention, encompass not only targeting a *AtSOD7* and/or *AtNGAL3* nucleic acid, for example a nucleic acid sequence comprising or consisting of SEQ ID NO: 1 or SEQ ID NO: 2, or SEQ ID NO: 4 respectively or a polypeptide comprising or consisting of SEQ ID NO: 3, or SEQ ID NO: 5, or a promoter of a *AtSOD7* and/or *AtNGAL3* nucleic acid.
35 The aspects of the invention encompass also functional variants of AtNGAL2 or AtNGAL3 that do not affect the biological activity and function of the resulting protein.

Alterations in a nucleic acid sequence which result in the production of a different amino acid at a given site that do however not affect the functional properties of the encoded polypeptide, are well known in the art. For example, a codon for the amino acid alanine, a hydrophobic amino acid, may be substituted by a codon encoding another less hydrophobic residue, such as glycine, or a more hydrophobic residue, such as valine, leucine, or isoleucine. Similarly, changes which result in substitution of one negatively charged residue for another, such as aspartic acid for glutamic acid, or one positively charged residue for another, such as lysine for arginine, can also produce a functionally equivalent product. Each of the proposed modifications is well within the routine skill in the art, as is determination of retention of biological activity of the encoded products. Also encompassed is a variant that is substantially identical, i.e. has only some sequence variations, for example in non-conserved residues, to the wild type sequences as shown herein and is biologically active.

Generally, variants of a particular *SOD7/NGAL3* nucleotide sequence or *NGAL2/NGAL3* polypeptide as described herein will have at least about 60%, preferably at least about 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 92%, 94%, 95%, 96%, 97%, 98% or 99% or more sequence identity to that particular non-variant nucleotide sequence, as determined by sequence alignment programs described elsewhere herein.

Furthermore, the various the aspects of the invention encompass not only a *AtSOD7* and/or *AtNGAL3* nucleic acid, for example a nucleic acid sequence comprising or consisting of SEQ ID NO: 1 or SEQ ID NO: 2, or SEQ ID NO: 4 respectively or a polypeptide comprising or consisting of SEQ ID NO: 3, or SEQ ID NO: 5, or their functional variants but also homologues of *AtSOD7* and/or *AtNGAL3* in *Arabidopsis* or other plants. Also within the scope of the invention are functional variants of such homologues as defined above.

The term homologue as used herein also designates an *AtSOD7* and/or *AtNGAL3* orthologue from other plant species. A homologue of *AtNGAL2* or *AtNGAL3* polypeptide respectively has, in increasing order of preference, at least 25%, 26%, 27%, 28%, 29%, 30%, 31%, 32%, 33%, 34%, 35%, 36%, 37%, 38%, 39%, 40%, 41%, 42%, 43%, 44%, 45%, 46%, 47%, 48%, 49%, 50%, 51%, 52%, 53%, 54%, 55%, 56%, 57%, 58%, 59%, 60%, 61%, 62%, 63%, 64%, 65%, 66%, 67%, 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%,

87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or at least 99% overall sequence identity to the amino acid represented by SEQ ID NO: 3 or 5 respectively. Preferably, overall sequence identity is at least 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99%, most preferably 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or at least 99%.

In another embodiment, the homologue of a *AtSOD7* or *AtNGAL3* nucleic acid sequence respectively has, in increasing order of preference, at least 25%, 26%, 27%, 28%, 29%, 30%, 31%, 32%, 33%, 34%, 35%, 36%, 37%, 38%, 39%, 40%, 41%, 42%, 43%, 44%, 45%, 46%, 47%, 48%, 49%, 50%, 51%, 52%, 53%, 54%, 55%, 56%, 57%, 58%, 59%, 60%, 61%, 62%, 63%, 64%, 65%, 66%, 67%, 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or at least 99% overall sequence identity to the nucleic acid represented by SEQ ID NO: 1 or 2 or 4 respectively. Preferably, overall sequence identity is at least 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99%, most preferably 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or at least 99%. The overall sequence identity is determined using a global alignment algorithm known in the art, such as the Needleman Wunsch algorithm in the program GAP (GCG Wisconsin Package, Accelrys).

In a preferred embodiment, the NGAL2 or NGAL3 homologue is from a plant that is not *Arabidopsis*.

In one embodiment, an *AtNGAL2* or a homologue thereof or *AtNGAL3* or a homologue thereof comprises a B3 domain having the sequence as defined above

In one embodiment, an *AtNGAL2* or a homologue thereof or *AtNGAL3* or a homologue thereof comprises a transcriptional repression motif having the sequence as defined above

Examples of homologues are shown in figure 13 and in SEQ ID NO: 49-145. In certain embodiments, if a plant has more than one *AtNGAL2* and/or *AtNGAL3* homologue, then all homologues are knocked out or knocked down. Suitable homologues can be

identified by sequence comparisons and identifications of conserved domains. There are predictors in the art that can be used to identify such sequences. The function of the homologue can be identified as described herein and a skilled person would thus be able to confirm the function, for example when overexpressed in a plant or knocked
5 out in a plant or when expressed in a plant or by expressing the homologous nucleic acid sequence in an Arabidopsis gain of function mutant.

Thus, the nucleotide sequences of the invention and described herein can also be used to isolate corresponding sequences from other organisms, particularly other plants, for
10 example crop plants. In this manner, methods such as PCR, hybridization, and the like can be used to identify such sequences based on their sequence homology to the sequences described herein. Topology of the sequences and the characteristic domains structure can also be considered when identifying and isolating homologues. Sequences may be isolated based on their sequence identity to the entire sequence or
15 to fragments thereof. In hybridization techniques, all or part of a known nucleotide sequence is used as a probe that selectively hybridizes to other corresponding nucleotide sequences present in a population of cloned genomic DNA fragments or cDNA fragments (i.e., genomic or cDNA libraries) from a chosen plant. The hybridization probes may be genomic DNA fragments, cDNA fragments, RNA
20 fragments, or other oligonucleotides, and may be labelled with a detectable group, or any other detectable marker. Thus, for example, probes for hybridization can be made by labelling synthetic oligonucleotides based on the ABA-associated sequences of the invention. Methods for preparation of probes for hybridization and for construction of cDNA and genomic libraries are generally known in the art and are disclosed in
25 Sambrook, et al., (1989) Molecular Cloning: A Library Manual (2d ed., Cold Spring Harbor Laboratory Press, Plainview, New York).

Hybridization of such sequences may be carried out under stringent conditions. By "stringent conditions" or "stringent hybridization conditions" is intended conditions
30 under which a probe will hybridize to its target sequence to a detectably greater degree than to other sequences (e.g., at least 2-fold over background). Stringent conditions are sequence dependent and will be different in different circumstances. By controlling the stringency of the hybridization and/or washing conditions, target sequences that are 100% complementary to the probe can be identified (homologous probing).
35 Alternatively, stringency conditions can be adjusted to allow some mismatching in sequences so that lower degrees of similarity are detected (heterologous probing).

Generally, a probe is less than about 1000 nucleotides in length, preferably less than 500 nucleotides in length.

Typically, stringent conditions will be those in which the salt concentration is less than about 1.5 M Na ion, typically about 0.01 to 1.0 M Na ion concentration (or other salts) at pH 7.0 to 8.3 and the temperature is at least about 30°C for short probes (e.g., 10 to 50 nucleotides) and at least about 60°C for long probes (e.g., greater than 50 nucleotides). Duration of hybridization is generally less than about 24 hours, usually about 4 to 12. Stringent conditions may also be achieved with the addition of destabilizing agents such as formamide.

According to the invention, preferred homologues of *AtSOD7* and *AtNGAL3* peptides are selected from crop plants, for example cereal crops. Preferred homologues of *AtNGAL2* and *AtNGAL3* and their polypeptide sequences are also shown in Fig. 13.

A plant according to the various aspects of the invention, including the transgenic plants, methods and uses described herein may be a monocot or a dicot plant.

A dicot plant may be selected from the families including, but not limited to *Asteraceae*, *Brassicaceae* (e.g. *Brassica napus*), *Chenopodiaceae*, *Cucurbitaceae*, *Leguminosae* (*Caesalpiniaceae*, *Aesalpiniaceae*, *Mimosaceae*, *Papilionaceae* or *Fabaceae*), *Malvaceae*, *Rosaceae* or *Solanaceae*. For example, the plant may be selected from lettuce, sunflower, *Arabidopsis*, broccoli, spinach, water melon, squash, cabbage, tomato, potato, yam, capsicum, tobacco, cotton, okra, apple, rose, strawberry, alfalfa, bean, soybean, field (fava) bean, pea, lentil, peanut, chickpea, apricots, pears, peach, grape vine, bell pepper, chilli or citrus species.

A monocot plant may, for example, be selected from the families *Arecaceae*, *Amaryllidaceae* or *Poaceae*. For example, the plant may be a cereal crop, such as maize, wheat, rice, barley, oat, sorghum, rye, millet, buckwheat, or a grass crop such as *Lolium* species or *Festuca* species, or a crop such as sugar cane, onion, leek, yam or banana.

Also included are biofuel and bioenergy crops such as rape/canola, sugar cane, sweet sorghum, *Panicum virgatum* (switchgrass), linseed, lupin and willow, poplar, poplar hybrids, *Miscanthus* or gymnosperms, such as loblolly pine. Also included are crops for

silage (maize), grazing or fodder (grasses, clover, sanfoin, alfalfa), fibres (e.g. cotton, flax), building materials (e.g. pine, oak), pulping (e.g. poplar), feeder stocks for the chemical industry (e.g. high erucic acid oil seed rape, linseed) and for amenity purposes (e.g. turf grasses for golf courses), ornamentals for public and private
5 gardens (e.g. snapdragon, petunia, roses, geranium, *Nicotiana* sp.) and plants and cut flowers for the home (African violets, Begonias, chrysanthemums, geraniums, *Coleus* spider plants, *Dracaena*, rubber plant).

10 Preferably, the plant is a crop plant. By crop plant is meant any plant which is grown on a commercial scale for human or animal consumption or use. In a preferred embodiment, the plant is a cereal.

Most preferred plants are maize, rice, wheat, oilseed rape/canola, sorghum, soybean,
15 sunflower, alfalfa, potato, tomato, tobacco, grape, barley, pea, bean, field bean, lettuce, cotton, sugar cane, sugar beet, broccoli or other vegetable brassicas or poplar.

The term "plant" as used herein encompasses whole plants, ancestors and progeny of the plants and plant parts, including seeds, fruit, shoots, stems, leaves, roots (including
20 tubers), flowers, and tissues and organs, wherein each of the aforementioned comprise the gene/nucleic acid of interest. The term "plant" also encompasses plant cells, suspension cultures, callus tissue, embryos, meristematic regions, gametophytes, sporophytes, pollen and microspores, again wherein each of the aforementioned comprises the gene/nucleic acid of interest.

25 According to the various aspects of the invention, including the plants and methods of the invention, abolishing, inactivating, repressing, reducing or down-regulating the activity of a *NGAL2* and/or *NGAL3* polypeptide can be achieved through different means. Such means that are within the scope of the various aspects of the invention
30 are methods for abolishing or reducing translation or transcription of the *SOD7* and/or *NGAL3* gene, destabilizing *SOD7* and/or *NGAL3* transcript stability, destabilizing *NGAL2* and/or *NGAL3* polypeptide stability or abolishing or reducing the activation or activity of the *NGAL2* and/or *NGAL3* or polypeptide. Thus, in one embodiment, endogenous *SOD7* and/or *NGAL3* gene or its promoter carry a functional mutation so
35 that no full length transcript is made. In another embodiment, the *SOD7* and/or *NGAL3* gene is silenced in said plant using gene silencing techniques. In another embodiment,

the *SOD7* and/or *NGAL3* nucleic acid sequence has been altered to introduce a mutation which results in a *NGAL2/NGAL3* protein with reduced or abolished activity. These embodiments and the techniques used are described in more detail below.

5 In another aspect, the invention relates to a method for altering a plant phenotype comprising reducing or abolishing the expression of a nucleic acid sequence encoding a *NGAL2* and/or *NGAL3* polypeptide and/or reducing or abolishing the activity of a *NGAL2* and/or *NGAL3* polypeptide relative to a control plant.

10 In another aspect, the invention relates to a method for making a plant with an altered phenotype comprising reducing or abolishing the expression of a nucleic acid sequence encoding a *NGAL2* and/or *NGAL3* polypeptide and/or reducing or abolishing the activity of a *NGAL2* and/or *NGAL3* polypeptide relative to a control plant.

15 As previously described, such methods above use genetic engineering methods.

In this aspect, a wild type plant may be targeted to simultaneously knock out or down both *SOD7* and *NGAL3* function. Alternatively, the method may comprise the following steps

20 a) Knocking out or down *SOD7* function in a first plant;
b) knocking out or down *NGAL3* function in a second plant and
c) crossing plants regenerated from said first plant with plants regenerated from said second plant.

25 In one embodiment of these methods, expression of a nucleic acid sequence encoding a *NGAL2* polypeptide or the activity of a *NGAL2* polypeptide is reduced or abolished. In another embodiment, expression of a nucleic acid sequence encoding a *NGAL3* polypeptide or the activity of a *NGAL3* polypeptide is reduced or abolished. In a preferred embodiment, the method comprises reducing or abolishing expression of a
30 nucleic acid sequence encoding a *NGAL2* polypeptide or the activity of a *NGAL2* polypeptide and reducing or abolishing expression of a nucleic acid sequence encoding a *NGAL3* polypeptide or the activity of a *NGAL3* polypeptide to create a double loss of function mutant.

35 For example, the method comprises reducing or abolishing expression of a nucleic acid sequence encoding a *NGAL2* polypeptide and reducing or abolishing expression of a

nucleic acid sequence encoding a NGAL3 polypeptide. In another embodiment, the method comprises reducing or abolishing activity of a NGAL2 polypeptide and reducing or abolishing activity of a NGAL3 polypeptide. In another embodiment, the method comprises reducing or abolishing expression of a nucleic acid sequence encoding a NGAL2 polypeptide and reducing or abolishing activity of a NGAL3 polypeptide. In another embodiment the method comprises reducing or abolishing expression of a nucleic acid sequence encoding a NGAL3 polypeptide or reducing or abolishing activity of a NGAL2 polypeptide.

According to these methods, the phenotype is preferably selected from increased organ size, for example increased seed size or increased seed weight. Increased seed size leads to an increase in yield and the methods of the invention also increased yield.

The term "yield" in general means a measurable produce of economic value, typically related to a specified crop, to an area, and to a period of time. Individual plant parts directly contribute to yield based on their number, size and/or weight, or the actual yield is the yield per square meter for a crop and year, which is determined by dividing total production (includes both harvested and appraised production) by planted square meters. The term "yield" as described herein relates to yield-related traits and may relate to vegetative biomass (root and/or shoot biomass), to reproductive organs, and/or to propagules (such as seeds) of that plant. Thus, according to the invention, the term yield refers to organ size, in particular seed size and can be measured by assessing seed size or seed weight or cotyledon size.

The terms "increase", "improve" or "enhance" are interchangeable. Yield or seed size for example is increased by at least a 2%, 3%, 4%, 5%, 6%, 7%, 8%, 9% or 10%, preferably at least 15% or 20%, more preferably 25%, 30%, 35%, 40% or 50% or more in comparison to a control plant.

A control plant as used herein according to all of the aspects of the invention is a plant which has not been modified according to the methods of the invention. Accordingly, the control plant has not been genetically modified to alter either expression of a nucleic acid encoding a NGAL2 or NGAL3 polypeptide or to alter the activity of a NGAL2 or NGAL3 polypeptide as described herein. In one embodiment, the control plant is a wild type plant that has not been genetically altered. In another embodiment, the control plant is a transgenic plant that does not have altered expression of a nucleic

acid encoding a NGAL2 or NGAL3 polypeptide or altered activity of a NGAL2 or NGAL3 polypeptide, but has been genetically altered in other ways, for example by expressing a desirable transgene to confer certain traits.

5 The reduction, decrease, down-regulation or repression of the activity of the NGAL2 and/or NGAL3 polypeptide or corresponding *SOD7* and/or *NGAL3* nucleic acid sequences according to the aspects of the invention is at least 10%, 20%, 30%, 40% or 50% in comparison to the control plant.

10 For example, the plant is a reduction (knock down) or loss of function (knock out) mutant wherein the function of the *SOD7* and/or *NGAL3* nucleic acid sequence is reduced or lost compared to a wild type control plant. To this end, a mutation is introduced into the *SOD7* and/or *NGAL3* nucleic acid sequence or the corresponding promoter sequence which disrupts the transcription of the gene leading to a gene
15 product which is not functional or has a reduced function. The mutation may be a deletion, insertion or substitution. The expression of active protein may thus be abolished by mutating the nucleic acid sequences in the plant cell which encode the NGAL2 or NGAL3 polypeptide and regenerating a plant from the mutated cell. The nucleic acids may be mutated by insertion or deletion of one or more nucleotides.

20 Techniques for the inactivation or knockout of target genes are well-known in the art. These techniques include gene target using vectors that target the gene of interest and which allow integration allows for integration of transgene at a specific site. The targeting construct is engineered to recombine with the target gene, which is accomplished by incorporating sequences from the gene itself into the construct.
25 Recombination then occurs in the region of that sequence within the gene, resulting in the insertion of a foreign sequence to disrupt the gene. With its sequence interrupted, the altered gene will be translated into a nonfunctional protein, if it is translated at all. Other techniques include genome editing (targeted genome engineering) as described below. Using either of these techniques, in preferred embodiment, conserved domains
30 which confer function of NGAL2 or NGAL3 respectively are modified.

A skilled person will know further approaches can be used to generate such mutants. In one embodiment, insertional mutagenesis is used, for example using T-DNA mutagenesis (which inserts pieces of the T-DNA from the *Agrobacterium tumefaciens*
35 T-Plasmid into DNA causing either loss of gene function or gain of gene function mutations), site-directed nucleases (SDNs) or transposons as mutagens. Insertional

mutagenesis is an alternative means of disrupting gene function and is based on the insertion of foreign DNA into the gene of interest (see Krysan et al, The Plant Cell, Vol. 11, 2283-2290, December 1999).

5 In one embodiment, as discussed in the examples, T-DNA may be used as an insertional mutagen which disrupts *SOD7* and/or *NGAL3* gene expression. T-DNA not only disrupts the expression of the gene into which it is inserted, but also acts as a marker for subsequent identification of the mutation. Since the sequence of the inserted element is known, the gene in which the insertion has occurred can be
10 recovered, using various cloning or PCR-based strategies. The insertion of a piece of T-DNA on the order of 5 to 25 kb in length generally produces a disruption of gene function. If a large enough population of T-DNA transformed lines is generated, there are reasonably good chances of finding a transgenic plant carrying a T-DNA insert within any gene of interest. Transformation of spores with T-DNA is achieved by an
15 *Agrobacterium*-mediated method which involves exposing plant cells and tissues to a suspension of *Agrobacterium* cells.

The details of this method are well known to a skilled person. In short, plant transformation by *Agrobacterium* results in the integration into the nuclear genome of a
20 sequence called T-DNA, which is carried on a bacterial plasmid. The use of T-DNA transformation leads to stable single insertions. Further mutant analysis of the resultant transformed lines is straightforward and each individual insertion line can be rapidly characterized by direct sequencing and analysis of DNA flanking the insertion. Gene expression in the mutant is compared to expression of the *SOD7* and/or *NGAL3*
25 nucleic acid sequence in a wild type plant and phenotypic analysis is also carried out. Other techniques for insertional mutagenesis include the use of transposons.

In another embodiment, mutagenesis is physical mutagenesis, such as application of ultraviolet radiation, X-rays, gamma rays, fast or thermal neutrons or protons. The
30 targeted population can then be screened to identify a *SOD7* or *NGAL3* loss of function mutant.

In another embodiment of the various aspects of the invention, the plant is a mutant plant derived from a plant population mutagenised with a mutagen. The mutagen may
35 be fast neutron irradiation or a chemical mutagen, for example selected from the following non-limiting list: ethyl methanesulfonate (EMS), methylmethane sulfonate

(MMS), N-ethyl-N-nitrosourea (ENU), triethylmelamine (1E M), N-methyl-N-nitrosourea (MNU), procarbazine, chlorambucil, cyclophosphamide, diethyl sulfate, acrylamide monomer, melphalan, nitrogen mustard, vincristine, dimethylnitrosamine, N-methyl-N'-nitro-Nitrosoguanidine (MNNG), nitrosoguanidine, 2-aminopurine, 7,12 dimethyl-benz(a)anthracene (DMBA), ethylene oxide, hexamethylphosphoramide, bisulfan, diepoxyalkanes (diepoxyoctane (DEO), diepoxybutane (BEB), and the like), 2-methoxy-6-chloro-9 [3-(ethyl-2-chloroethyl)aminopropylamino]acridine dihydrochloride (ICR-170) or formaldehyde.

In one embodiment, the method used to create and analyse mutations is targeting induced local lesions in genomes (TLLING), reviewed in Henikoff et al, 2004. In this method, seeds are mutagenised with a chemical mutagen, for example EMS. The resulting M1 plants are self-fertilised and the M2 generation of individuals is used to prepare DNA samples for mutational screening. DNA samples are pooled and arrayed on microtiter plates and subjected to gene specific PCR. The PCR amplification products may be screened for mutations in the *SOD7* and/or *NGAL3* target gene using any method that identifies heteroduplexes between wild type and mutant genes. For example, but not limited to, denaturing high pressure liquid chromatography (dHPLC), constant denaturant capillary electrophoresis (CDCE), temperature gradient capillary electrophoresis (TGCE), or by fragmentation using chemical cleavage. Preferably the PCR amplification products are incubated with an endonuclease that preferentially cleaves mismatches in heteroduplexes between wild type and mutant sequences. Cleavage products are electrophoresed using an automated sequencing gel apparatus, and gel images are analyzed with the aid of a standard commercial image-processing program. Any primer specific to the *SOD7* or *NGAL3* nucleic acid sequence may be utilized to amplify the *SOD7* or *NGAL3* nucleic acid sequence within the pooled DNA sample. Preferably, the primer is designed to amplify the regions of the *SOD7* and/or *NGAL3* gene where useful mutations are most likely to arise, specifically in the areas of the *SOD7* and/or *NGAL3* gene that are highly conserved and/or confer activity as explained elsewhere. To facilitate detection of PCR products on a gel, the PCR primer may be labelled using any conventional labelling method.

Rapid high-throughput screening procedures thus allow the analysis of amplification products for identifying a mutation conferring the reduction or inactivation of the expression of the *SOD7* and/or *NGAL3* gene as compared to a corresponding non-mutagenised wild type plant. Once a mutation is identified in a gene of interest, the

seeds of the M2 plant carrying that mutation are grown into adult M3 plants and screened for the phenotypic characteristics associated with the target gene *SOD7* or *NGAL3*. Loss of function or reduced function mutants with increased seed size compared to a control can thus be identified.

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Plants obtained or obtainable by such method which carry a functional mutation in the endogenous *SOD7* and/or *NGAL3* locus are also within the scope of the invention

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In another embodiment, RNA-mediated gene suppression or RNA silencing may be used to achieve silencing of the *SOD7* and/or *NGAL3* nucleic acid sequence. "Gene silencing" is a term generally used to refer to suppression of expression of a gene via sequence-specific interactions that are mediated by RNA molecules. The degree of reduction may be so as to totally abolish production of the encoded gene product, but more usually the abolition of expression is partial, with some degree of expression remaining. The term should not therefore be taken to require complete "silencing" of expression.

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Transgenes may be used to suppress endogenous plant genes. This was discovered originally when chalcone synthase transgenes in petunia caused suppression of the endogenous chalcone synthase genes and indicated by easily visible pigmentation changes. Subsequently it has been described how many, if not all plant genes can be "silenced" by transgenes. Gene silencing requires sequence similarity between the transgene and the gene that becomes silenced. This sequence homology may involve promoter regions or coding regions of the silenced target gene. When coding regions are involved, the transgene able to cause gene silencing may have been constructed with a promoter that would transcribe either the sense or the antisense orientation of the coding sequence RNA. It is likely that the various examples of gene silencing involve different mechanisms that are not well understood. In different examples there may be transcriptional or post-transcriptional gene silencing and both may be used according to the methods of the invention.

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The mechanisms of gene silencing and their application in genetic engineering, which were first discovered in plants in the early 1990s and then shown in *Caenorhabditis elegans* are extensively described in the literature.

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RNA-mediated gene suppression or RNA silencing according to the methods of the invention includes co-suppression wherein over-expression of the target sense RNA or mRNA, that is the *SOD7* and/or *NGAL3* sense RNA or mRNA, leads to a reduction in the level of expression of the genes concerned. RNAs of the transgene and homologous endogenous gene are co-ordinately suppressed. Other techniques used in the methods of the invention include antisense RNA to reduce transcript levels of the endogenous target gene in a plant. In this method, RNA silencing does not affect the transcription of a gene locus, but only causes sequence-specific degradation of target mRNAs. An "antisense" nucleic acid sequence comprises a nucleotide sequence that is complementary to a "sense" nucleic acid sequence encoding a *NGAL2* and/or *NGAL3* protein, or a part of the protein, i.e. complementary to the coding strand of a double-stranded cDNA molecule or complementary to an mRNA transcript sequence. The antisense nucleic acid sequence is preferably complementary to the endogenous *SOD7* and/or *NGAL3* gene to be silenced. The complementarity may be located in the "coding region" and/or in the "non-coding region" of a gene. The term "coding region" refers to a region of the nucleotide sequence comprising codons that are translated into amino acid residues. The term "non-coding region" refers to 5' and 3' sequences that flank the coding region that are transcribed but not translated into amino acids (also referred to as 5' and 3' untranslated regions).

Antisense nucleic acid sequences can be designed according to the rules of Watson and Crick base pairing. The antisense nucleic acid sequence may be complementary to the entire *SOD7* and/or *NGAL3* nucleic acid sequence, but may also be an oligonucleotide that is antisense to only a part of the nucleic acid sequence (including the mRNA 5' and 3' UTR). For example, the antisense oligonucleotide sequence may be complementary to the region surrounding the translation start site of an mRNA transcript encoding a polypeptide. The length of a suitable antisense oligonucleotide sequence is known in the art and may start from about 50, 45, 40, 35, 30, 25, 20, 15 or 10 nucleotides in length or less. An antisense nucleic acid sequence according to the invention may be constructed using chemical synthesis and enzymatic ligation reactions using methods known in the art. For example, an antisense nucleic acid sequence (e.g., an antisense oligonucleotide sequence) may be chemically synthesized using naturally occurring nucleotides or variously modified nucleotides designed to increase the biological stability of the molecules or to increase the physical stability of the duplex formed between the antisense and sense nucleic acid sequences, e.g., phosphorothioate derivatives and acridine-substituted nucleotides

may be used. Examples of modified nucleotides that may be used to generate the antisense nucleic acid sequences are well known in the art. The antisense nucleic acid sequence can be produced biologically using an expression vector into which a nucleic acid sequence has been subcloned in an antisense orientation (i.e., RNA transcribed from the inserted nucleic acid will be of an antisense orientation to a target nucleic acid of interest). Preferably, production of antisense nucleic acid sequences in plants occurs by means of a stably integrated nucleic acid construct comprising a promoter, an operably linked antisense oligonucleotide, and a terminator.

The nucleic acid molecules used for silencing in the methods of the invention hybridize with or bind to mRNA transcripts and/or insert into genomic DNA encoding a polypeptide to thereby inhibit expression of the protein, e.g., by inhibiting transcription and/or translation. The hybridization can be by conventional nucleotide complementarity to form a stable duplex, or, for example, in the case of an antisense nucleic acid sequence which binds to DNA duplexes, through specific interactions in the major groove of the double helix. Antisense nucleic acid sequences may be introduced into a plant by transformation or direct injection at a specific tissue site. Alternatively, antisense nucleic acid sequences can be modified to target selected cells and then administered systemically. For example, for systemic administration, antisense nucleic acid sequences can be modified such that they specifically bind to receptors or antigens expressed on a selected cell surface, e.g., by linking the antisense nucleic acid sequence to peptides or antibodies which bind to cell surface receptors or antigens. The antisense nucleic acid sequences can also be delivered to cells using vectors.

RNA interference (RNAi) is another post-transcriptional gene-silencing phenomenon which may be used according to the methods of the invention. This is induced by double-stranded RNA in which mRNA that is homologous to the dsRNA is specifically degraded. It refers to the process of sequence-specific post-transcriptional gene silencing mediated by short interfering RNAs (siRNA). The process of RNAi begins when the enzyme, DICER, encounters dsRNA and chops it into pieces called small-interfering RNAs (siRNA). This enzyme belongs to the RNase III nuclease family. A complex of proteins gathers up these RNA remains and uses their code as a guide to search out and destroy any RNAs in the cell with a matching sequence, such as target mRNA.

Artificial and/or natural microRNAs (miRNAs) may be used to knock out gene expression and/or mRNA translation. MicroRNAs (miRNAs) miRNAs are typically single stranded small RNAs typically 19-24 nucleotides long. Most plant miRNAs have perfect or near-perfect complementarity with their target sequences. However, there are natural targets with up to five mismatches. They are processed from longer non-coding RNAs with characteristic fold-back structures by double-strand specific RNases of the Dicer family. Upon processing, they are incorporated in the RNA-induced silencing complex (RISC) by binding to its main component, an Argonaute protein. miRNAs serve as the specificity components of RISC, since they base-pair to target nucleic acids, mostly mRNAs, in the cytoplasm. Subsequent regulatory events include target mRNA cleavage and destruction and/or translational inhibition. Effects of miRNA overexpression are thus often reflected in decreased mRNA levels of target genes. Artificial microRNA (amiRNA) technology has been applied in *Arabidopsis thaliana* and other plants to efficiently silence target genes of interest. The design principles for amiRNAs have been generalized and integrated into a Web-based tool (<http://wmd.weigelworld.org>).

Thus, according to the various aspects of the invention a plant may be transformed to introduce a RNAi, shRNA, snRNA, dsRNA, siRNA, miRNA, ta-siRNA, amiRNA or cosuppression molecule that has been designed to target the expression of an *SOD7* and/or *NGAL3* nucleic acid sequence and selectively decreases or inhibits the expression of the gene or stability of its transcript. Preferably, the RNAi, snRNA, dsRNA, shRNA siRNA, miRNA, amiRNA, ta-siRNA or cosuppression molecule used according to the various aspects of the invention comprises a fragment of at least 17 nt, preferably 22 to 26 nt and can be designed on the basis of the information shown in SEQ ID NO: 1. Guidelines for designing effective siRNAs are known to the skilled person. Briefly, a short fragment of the target gene sequence (e.g., 19-40 nucleotides in length) is chosen as the target sequence of the siRNA of the invention. The short fragment of target gene sequence is a fragment of the target gene mRNA. In preferred embodiments, the criteria for choosing a sequence fragment from the target gene mRNA to be a candidate siRNA molecule include 1) a sequence from the target gene mRNA that is at least 50-100 nucleotides from the 5' or 3' end of the native mRNA molecule, 2) a sequence from the target gene mRNA that has a G/C content of between 30% and 70%, most preferably around 50%, 3) a sequence from the target gene mRNA that does not contain repetitive sequences (e.g., AAA, CCC, GGG, TTT, AAAA, CCCC, GGGG, TTTT), 4) a sequence from the target gene mRNA that is

accessible in the mRNA, 5) a sequence from the target gene mRNA that is unique to the target gene, 6) avoids regions within 75 bases of a start codon. The sequence fragment from the target gene mRNA may meet one or more of the criteria identified above. The selected gene is introduced as a nucleotide sequence in a prediction
5 program that takes into account all the variables described above for the design of optimal oligonucleotides. This program scans any mRNA nucleotide sequence for regions susceptible to be targeted by siRNAs. The output of this analysis is a score of possible siRNA oligonucleotides. The highest scores are used to design double stranded RNA oligonucleotides that are typically made by chemical synthesis. In
10 addition to siRNA which is complementary to the mRNA target region, degenerate siRNA sequences may be used to target homologous regions. siRNAs according to the invention can be synthesized by any method known in the art. RNAs are preferably chemically synthesized using appropriately protected ribonucleoside phosphoramidites and a conventional DNA/RNA synthesizer. Additionally, siRNAs can be obtained from
15 commercial RNA oligonucleotide synthesis suppliers.

siRNA molecules according to the aspects of the invention may be double stranded. In one embodiment, double stranded siRNA molecules comprise blunt ends. In another embodiment, double stranded siRNA molecules comprise overhanging nucleotides
20 (e.g., 1-5 nucleotide overhangs, preferably 2 nucleotide overhangs). In some embodiments, the siRNA is a short hairpin RNA (shRNA); and the two strands of the siRNA molecule may be connected by a linker region (e.g., a nucleotide linker or a non-nucleotide linker). The siRNAs of the invention may contain one or more modified nucleotides and/or non-phosphodiester linkages. Chemical modifications well known in
25 the art are capable of increasing stability, availability, and/or cell uptake of the siRNA. The skilled person will be aware of other types of chemical modification which may be incorporated into RNA molecules.

In one embodiment, recombinant DNA constructs as described in US 6635805,
30 incorporated herein by reference, may be used.

The silencing RNA molecule is introduced into the plant using conventional methods, for example a vector and *Agrobacterium*-mediated transformation. Stably transformed plants are generated and expression of the *SOD7* and/or *NGAL3* gene compared to a
35 wild type control plant is analysed.

Silencing of the *SOD7* and/or *NGAL3* nucleic acid sequence may also be achieved using virus-induced gene silencing.

Thus, in one embodiment of the invention, the plant expresses a nucleic acid construct comprising a RNAi, shRNA snRNA, dsRNA, siRNA, miRNA, ta-siRNA, amiRNA or co-suppression molecule that targets the *SOD7* or *NGAL3* nucleic acid sequence as described herein and reduces expression of the endogenous *SOD7* or *NGAL3* nucleic acid sequence. A gene is targeted when, for example, the RNAi, snRNA, dsRNA, siRNA, shRNA miRNA, ta-siRNA, amiRNA or cosuppression molecule selectively decreases or inhibits the expression of the gene compared to a control plant. Alternatively, a RNAi, snRNA, dsRNA, siRNA, miRNA, ta-siRNA, amiRNA or cosuppression molecule targets A *SOD7* or *NGAL3* nucleic acid sequence when the RNAi, shRNA snRNA, dsRNA, siRNA, miRNA, ta-siRNA, amiRNA or cosuppression molecule hybridises under stringent conditions to the gene transcript.

Gene silencing may also occur if there is a mutation on an endogenous gene and/or a mutation on an isolated gene/nucleic acid subsequently introduced into a plant. The reduction or substantial elimination may be caused by a non-functional polypeptide. For example, the polypeptide may bind to various interacting proteins; one or more mutation(s) and/or truncation(s) may therefore provide for a polypeptide that is still able to bind interacting proteins (such as receptor proteins) but that cannot exhibit its normal function (such as signalling ligand).

A further approach to gene silencing is by targeting nucleic acid sequences complementary to the regulatory region of the gene (e.g., the promoter and/or enhancers) to form triple helical structures that prevent transcription of the gene in target cells. Other methods, such as the use of antibodies directed to an endogenous polypeptide for inhibiting its function *in planta*, or interference in the signalling pathway in which a polypeptide is involved, will be well known to the skilled man. In particular, it can be envisaged that manmade molecules may be useful for inhibiting the biological function of a target polypeptide, or for interfering with the signalling pathway in which the target polypeptide is involved.

In one embodiment, the suppressor nucleic acids may be anti-sense suppressors of expression of the *NGAL2* or *NGAL3* polypeptides. In using anti-sense sequences to down-regulate gene expression, a nucleotide sequence is placed under the control of a

promoter in a "reverse orientation" such that transcription yields RNA which is complementary to normal mRNA transcribed from the "sense" strand of the target gene.

5 An anti-sense suppressor nucleic acid may comprise an anti-sense sequence of at least 10 nucleotides from the target nucleotide sequence. It may be preferable that there is complete sequence identity in the sequence used for down-regulation of expression of a target sequence, and the target sequence, although total complementarity or similarity of sequence is not essential. One or more nucleotides
10 may differ in the sequence used from the target gene. Thus, a sequence employed in a down-regulation of gene expression in accordance with the present invention may be a wild-type sequence (e.g. gene) selected from those available, or a variant of such a sequence.

15 The sequence need not include an open reading frame or specify an RNA that would be translatable. It may be preferred for there to be sufficient homology for the respective anti-sense and sense RNA molecules to hybridise. There may be down regulation of gene expression even where there is about 5%, 10%, 15% or 20% or more mismatch between the sequence used and the target gene. Effectively, the
20 homology should be sufficient for the down-regulation of gene expression to take place.

Suppressor nucleic acids may be operably linked to tissue-specific or inducible promoters. For example, integument and seed specific promoters can be used to specifically down-regulate a *SOD7* or *NGAL3* nucleic acids in developing ovules and
25 seeds to increase final seed size.

Nucleic acid which suppresses expression of a *NGAL2* or *NGAL3* polypeptide as described herein may be operably linked to a heterologous regulatory sequence, such as a promoter, for example a constitutive, inducible, tissue-specific or developmental
30 specific promoter. The construct or vector may be transformed into plant cells and expressed as described herein. Plant cells comprising such vectors are also within the scope of the invention.

In another aspect, the invention relates to a silencing construct to silence expression of
35 *NGAL2* or *NGAL3* obtainable or obtained by a method as described herein and to a plant cell comprising such construct. Accordingly, the invention also relates to the use

of a nucleic acid sequence comprising or consisting of SEQ ID NO: 1, 2 or 3 or a part thereof or a homologue of SEQ ID NO: 1, 2 or 3 or a part thereof in silencing expression of *NGAL2* or *NGAL3*. Host cells transformed with such construct are also within the scope of the invention.

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Recently, genome editing techniques have emerged as alternative methods to conventional mutagenesis methods (such as physical and chemical mutagenesis) or methods using the expression of transgenes in plants to produce mutant plants with improved phenotypes that are important in agriculture. These techniques employ sequence-specific nucleases (SSNs) including zinc finger nucleases (ZFNs), transcription activator-like effector nucleases (TALENs), and the RNA-guided nuclease Cas9 (CRISPR/Cas9), which generate targeted DNA double-strand breaks (DSBs), which are then repaired mainly by either error-prone non-homologous end joining (NHEJ) or high-fidelity homologous recombination (HR). The SSNs have been used to create targeted knockout plants in various species ranging from the model plants, *Arabidopsis* and tobacco, to important crops, such as barley, soybean, rice and maize. Heritable gene modification has been demonstrated in *Arabidopsis* and rice using the CRISPR/Cas9 system and TALENs.

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Targeted genome modification or targeted genome editing is a genome engineering technique that uses targeted DNA double-strand breaks (DSBs) to stimulate genome editing through homologous recombination (HR)-mediated recombination events. To achieve effective genome editing via introduction of site-specific DNA DSBs, four major classes of customizable DNA binding proteins can be used: meganucleases derived from microbial mobile genetic elements, ZF nucleases based on eukaryotic transcription factors, transcription activator-like effectors (TALEs) from *Xanthomonas* bacteria, and the RNA-guided DNA endonuclease Cas9 from the type II bacterial adaptive immune system CRISPR (clustered regularly interspaced short palindromic repeats). Meganuclease, ZF, and TALE proteins all recognize specific DNA sequences through protein-DNA interactions. Although meganucleases integrate its nuclease and DNA-binding domains, ZF and TALE proteins consist of individual modules targeting 3 or 1 nucleotides (nt) of DNA, respectively. ZFs and TALEs can be assembled in desired combinations and attached to the nuclease domain of FokI to direct nucleolytic activity toward specific genomic loci.

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Upon delivery into host cells via the bacterial type III secretion system, TAL effectors enter the nucleus, bind to effector-specific sequences in host gene promoters and activate transcription. Their targeting specificity is determined by a central domain of tandem, 33-35 amino acid repeats. This is followed by a single truncated repeat of 20 amino acids. The majority of naturally occurring TAL effectors examined have between 12 and 27 full repeats.

These repeats only differ from each other by two adjacent amino acids, their repeat-variable di-residue (RVD). The RVD that determines which single nucleotide the TAL effector will recognize: one RVD corresponds to one nucleotide, with the four most common RVDs each preferentially associating with one of the four bases. Naturally occurring recognition sites are uniformly preceded by a T that is required for TAL effector activity. TAL effectors can be fused to the catalytic domain of the FokI nuclease to create a TAL effector nuclease (TALEN) which makes targeted DNA double-strand breaks (DSBs) *in vivo* for genome editing. The use of this technology in genome editing is well described in the art, for example in US 8,440,431, US 8,440,432 and US 8,450,471. Reference 30 describes a set of customized plasmids that can be used with the Golden Gate cloning method to assemble multiple DNA fragments. As described therein, the Golden Gate method uses Type IIS restriction endonucleases, which cleave outside their recognition sites to create unique 4 bp overhangs. Cloning is expedited by digesting and ligating in the same reaction mixture because correct assembly eliminates the enzyme recognition site. Assembly of a custom TALEN or TAL effector construct and involves two steps: (i) assembly of repeat modules into intermediary arrays of 1-10 repeats and (ii) joining of the intermediary arrays into a backbone to make the final construct.

Another genome editing method that can be used according to the various aspects of the invention is CRISPR. The use of this technology in genome editing is well described in the art, for example in US 8,697,359 and references cited herein. In short, CRISPR is a microbial nuclease system involved in defense against invading phages and plasmids. CRISPR loci in microbial hosts contain a combination of CRISPR-associated (Cas) genes as well as non-coding RNA elements capable of programming the specificity of the CRISPR-mediated nucleic acid cleavage (sgRNA). Three types (I-III) of CRISPR systems have been identified across a wide range of bacterial hosts. One key feature of each CRISPR locus is the presence of an array of repetitive sequences (direct repeats) interspaced by short stretches of non-repetitive sequences

(spacers). The non-coding CRISPR array is transcribed and cleaved within direct repeats into short crRNAs containing individual spacer sequences, which direct Cas nucleases to the target site (protospacer). The Type II CRISPR is one of the most well characterized systems and carries out targeted DNA double-strand break in four sequential steps. First, two non-coding RNA, the pre-crRNA array and tracrRNA, are transcribed from the CRISPR locus. Second, tracrRNA hybridizes to the repeat regions of the pre-crRNA and mediates the processing of pre-crRNA into mature crRNAs containing individual spacer sequences. Third, the mature crRNA:tracrRNA complex directs Cas9 to the target DNA via Watson-Crick base-pairing between the spacer on the crRNA and the protospacer on the target DNA next to the protospacer adjacent motif (PAM), an additional requirement for target recognition. Finally, Cas9 mediates cleavage of target DNA to create a double-stranded break within the protospacer.

Cas9 is thus the hallmark protein of the type II CRISPR-Cas system, and a large monomeric DNA nuclease guided to a DNA target sequence adjacent to the PAM (protospacer adjacent motif) sequence motif by a complex of two noncoding RNAs: CRISPR RNA (crRNA) and trans-activating crRNA (tracrRNA). The Cas9 protein contains two nuclease domains homologous to RuvC and HNH nucleases. The HNH nuclease domain cleaves the complementary DNA strand whereas the RuvC-like domain cleaves the non-complementary strand and, as a result, a blunt cut is introduced in the target DNA. Heterologous expression of Cas9 together with an sgRNA can introduce site-specific double strand breaks (DSBs) into genomic DNA of live cells from various organisms. For applications in eukaryotic organisms, codon optimized versions of Cas9, which is originally from the bacterium *Streptococcus pyogenes*, have been used.

The single guide RNA (sgRNA) is the second component of the CRISPR/Cas system that forms a complex with the Cas9 nuclease. sgRNA is a synthetic RNA chimera created by fusing crRNA with tracrRNA. The sgRNA guide sequence located at its 5' end confers DNA target specificity. Therefore, by modifying the guide sequence, it is possible to create sgRNAs with different target specificities. The canonical length of the guide sequence is 20 bp. In plants, sgRNAs have been expressed using plant RNA polymerase III promoters, such as U6 and U3.

Using these techniques, it is possible to specifically target conserved domains to abolish the function of the NGAL2 and/or NGAL3 polypeptide.

For example, the conserved B3 domain or repression motif may be targeted.

5 Thus, in another embodiment of the invention directed to a mutant plant, plant cell, plant or a part thereof characterised in that the activity of a NGAL2 polypeptide is altered and said plant expresses a nucleic acid comprising a mutant SEQ ID NO. 1 or 2 and encoding a mutant NGAL2 polypeptide, a functional homologue or variant thereof, for example one which carries a mutation in the B3 or repressor domain.

10 Thus, in another embodiment of the invention directed to a mutant plant, plant cell, plant or a part thereof characterised in that the activity of a NGAL3 polypeptide is altered and said plant expresses a nucleic acid comprising a mutant SEQ ID NO. 4 and encoding a mutant NGAL3 polypeptide, a functional homologue or variant thereof which carries a mutation in the B3 or repressor domain.

15 In a preferred embodiment, the invention directed to a mutant plant, plant cell, plant or a part thereof characterised in that the activity of a NGAL2 and a NGAL3 polypeptide is altered and said plant expresses a nucleic acid comprising a mutant SEQ ID NO. 1 or 2 and encoding a mutant NGAL2 polypeptide, a functional homologue or variant thereof, 20 for example one which carries a mutation in the B3 or repressor domain and said plant expresses a nucleic acid comprising a mutant SEQ ID NO. 4 and encoding a mutant NGAL3 polypeptide which carries a mutation in the B3 or repressor domain.

25 Mutations in the promoter region of *SOD7* and/or *NGAL3* resulting in a loss of function are also within the scope of the invention.

Constructs designed using the genome editing technologies to knock out or knock down *NGAL2* or *NGAL3*, for example as shown herein, are also within the scope of the invention as well as host cells comprising these constructs. In one embodiment, the 30 constructs comprise or consist of a sequence selected from SEQ ID NO: 155, 156, 157 or 158. Accordingly, in a further aspect of the invention, there is provided a nucleic acid construct comprising a sequence selected from SEQ ID NO: 155, 156, 157 or 158. In a further aspect of the invention, there is provided a nucleic acid construct comprising at least one CRISPR target sequence, wherein the target sequence is selected from SEQ 35 ID Nos 150, 160, 161, 162 and 163. Preferably, the target sequence comprises at least

two CRISPR target sequences, preferably SEQ ID No 159 and 160 or SEQ ID No 161 and 162, or SEQ ID No 161 and 163 or SEQ ID No 159 and 163.

5 In another embodiment of the methods of the invention, inactivating, repressing or down-regulating the activity of NGAL2 and/or NGAL3 can be achieved by manipulating the expression of *SOD7* and/or *NGAL3* inhibitors in a plant, for example transgenic plant. For example, a gene expressing a protein that inhibits the expression of the *SOD7* and/or *NGAL3* gene or activity of the *SOD7* and/or *NGAL3* protein can be introduced into a plant and over-expressed. The inhibitor may interact with the regulatory sequences that direct *SOD7* and/or *NGAL3* gene expression to down-regulate or repress *SOD7* and/or *NGAL3* gene expression. For example, the inhibitor may be a transcriptional repressor. Alternatively, it may interact and repress transcriptional regulators, for example transcription factors, that positively regulate expression of the *SOD7* and/or *NGAL3* gene. Alternatively, the inhibitor it may directly interact with the NGAL2 and/or NGAL3 protein to inhibit its activity or interact with modulators of the NGAL2 and/or NGAL3 protein. For example, the activity of the NGAL2 and/or NGAL3 protein may be inactivated, repressed or down-regulated by manipulating post-transcriptional modifications, of the NGAL2 and/or NGAL3 protein resulting in a reduced or lost activity.

20 In one embodiment, the methods of the invention comprise comparing the activity of the NGAL2 and/or NGAL3 polypeptide and/or expression of the *SOD7* and/or *NGAL3* gene with the activity of the NGAL2 and/or NGAL3 polypeptide and/or expression of the *SOD7* and/or *NGAL3* gene in a control plant.

25 In another aspect, the invention relates to a plant obtainable or obtained by a method as described herein.

30 In another aspect, the invention relates to an expression cassette comprising an isolated nucleic acid sequence comprising or consisting of a sequence as shown in SEQ ID NO: 1 or 2 a functional part, variant, homologue or orthologue thereof operably linked to a regulatory element. In another aspect, the invention relates to an expression cassette comprising an isolated nucleic acid sequence comprising or consisting of a sequence as shown in SEQ ID NO: 4 or a functional part, variant, homologue or orthologue thereof operably linked to a regulatory element. The regulatory element may be a promoter. The invention also relates to a vector comprising such expression

cassette. The invention also relates to a composition comprising the two expression cassettes above.

5 In the methods described here, plants can be regenerated from plants transformed or genetically altered as described above and the phenotype, specifically the seed phenotype is analysed by known methods.

10 Transformation methods are known in the art. The nucleic acid sequence is introduced into said plant through a process called transformation. The term "introduction" or "transformation" as referred to herein encompasses the transfer of an exogenous polynucleotide into a host cell, irrespective of the method used for transfer. Plant tissue capable of subsequent clonal propagation, whether by organogenesis or embryogenesis, may be transformed with a genetic construct of the present invention and a whole plant regenerated there from. The particular tissue chosen will vary
15 depending on the clonal propagation systems available for, and best suited to, the particular species being transformed. Exemplary tissue targets include leaf disks, pollen, embryos, cotyledons, hypocotyls, megagametophytes, callus tissue, existing meristematic tissue (e.g., apical meristem, axillary buds, and root meristems), and induced meristem tissue (e.g., cotyledon meristem and hypocotyl meristem). The
20 polynucleotide may be transiently or stably introduced into a host cell and may be maintained non-integrated, for example, as a plasmid. Alternatively, it may be integrated into the host genome. The resulting transformed plant cell may then be used to regenerate a transformed plant in a manner known to persons skilled in the art.

25 The transfer of foreign genes into the genome of a plant is called transformation. Transformation of plants is now a routine technique in many species. Advantageously, any of several transformation methods may be used to introduce the gene of interest into a suitable ancestor cell. The methods described for the transformation and regeneration of plants from plant tissues or plant cells may be utilized for transient or
30 for stable transformation. Transformation methods include the use of liposomes, electroporation, chemicals that increase free DNA uptake, injection of the DNA directly into the plant, particle gun bombardment, transformation using viruses or pollen and microprojection. Methods may be selected from the calcium/polyethylene glycol method for protoplasts, electroporation of protoplasts, microinjection into plant material,
35 DNA or RNA-coated particle bombardment, infection with (non-integrative) viruses and

the like. Transgenic plants, including transgenic crop plants, are preferably produced via *Agrobacterium tumefaciens* mediated transformation.

To select transformed plants, the plant material obtained in the transformation is, as a rule, subjected to selective conditions so that transformed plants can be distinguished from untransformed plants. For example, the seeds obtained in the above-described manner can be planted and, after an initial growing period, subjected to a suitable selection by spraying. A further possibility is growing the seeds, if appropriate after sterilization, on agar plates using a suitable selection agent so that only the transformed seeds can grow into plants. Alternatively, the transformed plants are screened for the presence of a selectable marker such as the ones described above. Following DNA transfer and regeneration, putatively transformed plants may also be evaluated, for instance using Southern analysis, for the presence of the gene of interest, copy number and/or genomic organisation. Alternatively or additionally, expression levels of the newly introduced DNA may be monitored using Northern and/or Western analysis, both techniques being well known to persons having ordinary skill in the art.

The generated transformed plants may be propagated by a variety of means, such as by clonal propagation or classical breeding techniques. For example, a first generation (or T1) transformed plant may be selfed and homozygous second-generation (or T2) transformants selected, and the T2 plants may then further be propagated through classical breeding techniques. The generated transformed organisms may take a variety of forms. For example, they may be chimeras of transformed cells and non-transformed cells; clonal transformants (e.g., all cells transformed to contain the expression cassette); grafts of transformed and untransformed tissues (e.g., in plants, a transformed rootstock grafted to an untransformed scion).

The various aspects of the invention described herein clearly extend to any plant cell or any plant produced, obtained or obtainable by any of the methods described herein, and to all plant parts and propagules thereof unless otherwise specified. The present invention extends further to encompass the progeny of a primary transformed or transfected cell, tissue, organ or whole plant that has been produced by any of the aforementioned methods, the only requirement being that progeny exhibit the same genotypic and/or phenotypic characteristic(s) as those produced by the parent in the methods according to the invention.

The invention also extends to harvestable parts of a plant of the invention as described above such as, but not limited to seeds, leaves, fruits, flowers, stems, roots, rhizomes, tubers and bulbs. The invention furthermore relates to products derived, preferably
5 directly derived, from a harvestable part of such a plant, such as dry pellets or powders, oil, fat and fatty acids, starch or proteins. The invention also relates to food products and food supplements comprising the plant of the invention or parts thereof.

While the foregoing disclosure provides a general description of the subject matter
10 encompassed within the scope of the present invention, including methods, as well as the best mode thereof, of making and using this invention, the following examples are provided to further enable those skilled in the art to practice this invention and to provide a complete written description thereof. However, those skilled in the art will appreciate that the specifics of these examples should not be read as limiting on the
15 invention, the scope of which should be apprehended from the claims and equivalents thereof appended to this disclosure. Various further aspects and embodiments of the present invention will be apparent to those skilled in the art in view of the present disclosure.

20 All documents mentioned in this specification are incorporated herein by reference in their entirety, including references to gene and protein accession numbers.

"and/or" where used herein is to be taken as specific disclosure of each of the multiple specified features or components with or without the other at each combination unless
25 otherwise dictated. For example "A, B and/or C" is to be taken as specific disclosure of each of (i) A, (ii) B, (iii) C, (iv) A and B, (v) B and C or (vi) A and B and C, just as if each is set out individually herein.

Unless context dictates otherwise, the descriptions and definitions of the features set
30 out above are not limited to any particular aspect or embodiment of the invention and apply equally to all aspects and embodiments which are described.

The invention is further described in the following non-limiting examples.

35 **Examples**

METHODS

Plant materials and growth conditions

Arabidopsis thaliana Columbia (Col-0) was used as wild-type line. The *da 1-1*, *sod7-1 D*, *sod7-ko1* and *ngal3-ko1* were in the Col-0 background. *sod7-1D* was identified as a suppressor of *da 1-1* by using T-DNA activation tagging method. The *sod7-ko1* (SM_3_34191) and *ngal3-ko1* (SM_3_36641) were identified in AtIDB (www.atidb.org) and obtained from Arabidopsis Stock Centre NASC collection. T-DNA insertions were confirmed by PCR and sequencing by using the primers described in Table 1. Arabidopsis plants were grown under long-day conditions (16 h light/8 h dark) at 22°C.

Activation tagging screening The activation tagging plasmid pJFAT260 was introduced into the *da1-1* mutant plants using Agrobacterium tumefaciens strain GV3101 (Fan et al., 2009; Fang et al., 2012), and T1 plants were selected by using the herbicide Basta. Seeds produced from T1 plants were used to isolate modifiers of *da 1-1*.

Morphological and cellular analysis

To measure seed size, we photographed dry seeds of the wild type and mutants under a Leica microscope (LEICA S8APO) using Leica CCD (DFC420). The projective area of wild-type and mutant seeds was measured by using Image J software. Average seed weight was determined by weighing mature dry seeds in batches of 100 using an electronic analytical balance (METTLER TOLEDO AL104, China). The weights of five sample batches were measured for each seed lot. Fully expanded cotyledons, petals (stage 14) and leaves were scanned to produce digital images for area measurement. To measure cell number and cell size, petals, leaves, ovules and seeds were placed in a drop of clearing solution [30ml H₂O, 80g Chloral hydrate (Sigma, C8383), 10 ml 100% Glycerol (Sigma, G6279)]. Cleared Samples were imaged under a Leica microscope (LEICA DM2500) with differential interference contrast (DIC) optics and photographed with a SPOT FLEX Cooled CCD Digital Imaging System. Area measurement was made by using Image J software.

Cloning of the SOD7 gene

The flanking sequences of the T-DNA insertion of the *sod7-1D* mutant were identified by the thermal asymmetric interlaced PCR (TAIL-PCR) according to a previously reported method (Liu et al., 1995). Briefly, TAIL-PCR utilizes three nested specific primers (OJF22, OJF23 and OJF24) within the T-DNA region of the pJFAT260 vector and a shorter arbitrary degenerate primer (AD1). Thus, the relative amplification efficiencies of specific and non-specific products can be thermally controlled. TAIL-

PCR products were sequenced using the primer OJF24. The specific primers OJF22, OJF23 and OJF24 and an arbitrary degenerate (AD1) primer are described in Table 1.

Constructs and plant transformation

5 The *35S:GFP-SOD7*, *pSOD7:SOD7-GFP* and *pSOD7:GUS* constructs were made using a PCR-based Gateway system. The coding sequence (CDS) of SOD7 was amplified using the primers SOD7CDS-F and SOD7CDS-R (Table 1). PCR products were cloned into pCR8/TOPO TA cloning vector. The SOD7 CDS was then subcloned into the binary vector pMDC43 with the GFP gene to generate the transformation
10 plasmid *35S:GFP-SOD7*. The SOD7 genomic sequence containing 2040-bp promoter sequence and 2104-bp SOD7 gene was amplified using the primers SOD7G-F and SOD7G-R (Table 1). PCR products were cloned into pCR8/TOPO TA cloning vector. The SOD7 genomic sequence was then subcloned into the binary vectors pMDC107 with the GFP gene to generate the transformation plasmid *pSOD7:SOD7-GFP*. The
15 2262-bp SOD7 promoter sequence was amplified using the primers SOD7P-F and SOD7P-R (Table 1). PCR products were cloned into pCR8/TOPO TA cloning vector. The SOD7 promoter was then subcloned into the binary vectors pGWB3 with the GUS gene to generate the transformation plasmid *pSOD7:GUS*. The plasmids *35S:GFP-SOD7*, *pSOD7:SOD7-GFP* and *pSOD7:GUS* were introduced into Col-0 or *sod7-ko1*
20 *ngal3ko1* plants using *Agrobacterium tumefaciens* GV3101, respectively, and transformants were selected on hygromycin (30µg/ml)-containing medium. The SOD7 cDNA was cloned into the *Apal* and *SpeI* sites of the binary vector pER8 to generate a chemically inducible construct pER8-SOD7. The specific primers for the pER8-SOD7 construct were SOP7ER-F and SOD7ER-R. The plasmid pER8-SOD7 was introduced
25 into Col-0 plants using *Agrobacterium tumefaciens* GV3101, and transformants were selected on hygromycin (30µg/ml)-containing medium. GUS staining Samples (*pSOD7:GUS*) were stained in a GUS staining solution (1 mM X-gluc, 50 mM NaPO₄ buffer, 0.4 mM each K₃Fe(CN)₆/K₄Fe(CN)₆, and 0.1% (v/v) Triton X-100) and incubated at 37°C for 3 hours. After GUS staining, chlorophyll was removed by 70%
30 ethanol. RT-PCR and quantitative real-time RT-PCR. Total RNA was extracted from *Arabidopsis* seedlings using an RNAPrep pure Plant kit (TIANGEN). mRNA was reverse transcribed into cDNA using SuperScriptIII reverse transcriptase (Invitrogen). cDNA samples were standardized on ACTIN2 transcript amount using the primers ACTIN2-F and ACTIN2-R (Table 1). Quantitative real-time RT-PCR analysis was
35 performed with a Lightcycler 480 machine (Roche) using the Lightcycler 480 SYBR Green I Master (Roche). ACTIN2 mRNA was used as an internal control, and relative

amounts of mRNA were calculated using the comparative threshold cycle method. The primers used for RT-PCR and quantitative real-time RT-PCR are described in Table 1.

The chromatin immunoprecipitation (ChIP) assay

5 The chromatin immunoprecipitation (ChIP) assay was performed as described previously with minor modifications (Gendrel et al., 2005). Briefly, 35S:GFP and 35S:GFP-SOD7 transgenic seeds were grown on 1/2 MS plates for 10 days. The seedlings were cross-linked by 1% formaldehyde for 15 min in vacuum and stopped by 0.125 M Glycine. Samples were ground in liquid nitrogen, and nuclei were isolated.
10 Chromatin was immunoprecipitated by anti-GFP (Roche, 11814460001) and protein A+G beads (Millipore Magna ChIP Protein A+G Magnetic Beads, 16-663). DNA was precipitated by glycogen, NaOAc and ethanol, washed by 70% ethanol, and dissolved in 60 µl of water. Gene-specific primers (PF1-F, PF1-R, PF2-F, PF2-R, ACTIN7-ChIP-F, and ACTIN7-ChIP-R) were used to quantify the enrichment of each fragment (Table
15 1).

The DNA electrophoretic mobility shift assay (EMSA)

The coding sequence of SOD7 was cloned into the NdeI and BamHI sites of the pMAL-C2 vector to generate the construct MBP-SOD7. MBP-SOD7 fusion proteins were
20 expressed in Escherichia coli BL21 (DE3) (Biomed) and purified by Amylose resins (New England Biolabs). The biotin-labeled and unlabeled probes were synthesized as forward and reverse strands. The forward and reverse strands were then incubated in a solution (50mM Tris-HCl, 5mM EDTA and 250mM NaCl) at 95 °C for 10min and renatured to double stranded probes at room temperature. The gel-shift
25 assay was performed according to the method described previously (Smaczniak et al., 2012).

Results

sod7-1D suppresses the seed size phenotype of da1-1

We previously identified the ubiquitin receptor DA1 as a negative regulator of seed size in Arabidopsis (Li et al., 2008). The *da1-1* mutant formed large seeds due to increased cell proliferation in the maternal integuments (Li et al., 2008; Xia et al., 2013). To identify novel components in the DA1 pathway or other seed size regulators, we
35 initiated a T-DNA activation tagging screen for modifiers of *da1-1* (Fang et al., 2012). A dominant suppressor of *da1-1* (*sod7-1D*) was isolated from seeds produced from

approximate 16,000 T1 plants (Fig.1A). Seeds of the *sod7-1D da1-1* double mutant were significantly smaller and lighter than *da1-1* seeds (Figures 1A, E and F). The embryo constitutes the major volume of a mature seed in Arabidopsis. *sod7-1D da1-1* embryos were smaller than *da1-1* embryos (Figure 1B). The size of *sod7-1D da1-1* cotyledons was significantly reduced, compared with that of *da1-1* cotyledons (Figure 1G). In addition, *sod7-1D da1-1* double mutant formed smaller leaves and flowers than *da1-1* (Figures 1C and 1D). Thus, these results show that the *sod7-1D* mutation suppressed the seed and organ size phenotypes of *da1-1*.

sod7-1D produces small seeds

We isolated the single *sod7-1D* mutant among F2 progeny derived from a cross between the wild type (Col-0) and *sod7-1D da1-1*. The *sod7-1D* seeds were significantly smaller and lighter than wild-type seeds (Figures 2A, B, G and H). We further isolated and visualized embryos from mature wild-type and *sod7-1D* seeds. The *sod7-1D* embryos were obviously smaller than wild-type embryos (Figures 2C and D). The changes in seed size were also reflected in the size of seedlings (Figures 2E and F). The 10-d old *sod7-1D* cotyledons were significantly smaller than wild-type cotyledons (Figure. 2E, F and I). In addition, the *sod7-1D* mutants exhibited small leaves and flowers compared with the wild type. The decreased size of *sod7-1D* leaves and petals was not caused by smaller cells, indicating that the *sod7-1D* mutation results in a decrease in cell number. In fact, the average area of epidermal cells in *sod7-1D* petals was larger than that in wild-type petals, suggesting a possible compensation mechanism between cell number and cell size.

SOD7 encodes a B3 domain transcriptional repressor NGAL2

To determine whether the seed and organ size phenotypes of *sod7-1D* was caused by the T-DNA insertion, we firstly analyzed the genetic linkage of the mutant phenotypes with Basta resistance, which is conferred by the selectable marker of the activation tagging vector (Fan et al., 2009). In a T2 population, 181 plants with *sod7-1D da1-1* phenotypes were resistant, whereas 55 plants with *da1-1* phenotypes were sensitive, indicating that the insertion is cosegregated with the *sod7-1D* phenotypes. To clone the SOD7 gene, we isolated the T-DNA flanking sequences using thermal asymmetric interlaced PCR (Liu et al., 1995). DNA sequencing revealed that the T-DNA had inserted approximately 5.6 kb upstream of the At3g1 1580 and about 3.7 kb upstream of the At3g1 1590 gene (Figure 3A). To determine which gene is responsible for the *sod7-1D* phenotypes, we examined the mRNA levels of these two genes. The mRNA

of the At3g1 1590 gene accumulated at a similar level in *sod7-1D da1-1* and *da1-1*, suggesting that At3g1 1590 is not the *SOD7* gene (Figure 3B). By contrast, expression level of the At3g1 1580 gene in *sod7-1D da1-1* plants was dramatically higher than that in *da1-1* plants, suggesting that At3g1 1580 is the *SOD7* gene (Figure 3B). To further confirm whether the *sod7-1D* phenotypes were caused by ectopic At3g1 1580 expression, we overexpressed the At3g1 1580 gene (35S:GFP-*SOD7*) in wild-type plants (Col-0) and isolated 37 transgenic plants. Most transgenic lines showed small seeds and organs (Figures 3D-F), similar to those observed in the *sod7-1D* single mutant, indicating that At3g1 1580 is the *SOD7* gene. The *SOD7* gene encodes a NGATHA like protein (NGAL2) containing a B3 DNA-binding domain and a transcriptional repression motif (Figure 3C) (Alvarez et al., 2009; Ikeda and Ohme-Takagi, 2009; Trigueros et al., 2009). *SOD7* belongs to the RAV gene family that consists of 13 members in Arabidopsis (Figure 10) (Swaminathan et al., 2008). Several members of the RAV family contain the putative transcriptional repression motifs, including NGA1, NGA2, NGA3, NGA4, NGAL1, NGAL2/*SOD7* and NGAL3 (Figure 10) (Ikeda and Ohme-Takagi, 2009). The transcriptional repression motifs in NGA1, NGAL1 and NGAL2/*SOD7* have been known to possess the repressive activity (Ikeda and Ohme-Takagi, 2009), indicating that they are transcriptional repressors. *SOD7* exhibits the highest similarity to Arabidopsis NGAL3/DEVELOPMENT-RELATED PcG TARGET IN THE APEX 4 (DPA4) (Figure 10), which has known roles in the regulation of leaf serrations (Engelhorn et al., 2012), but no previously identified function in seed size control.

Expression pattern and subcellular localization of *SOD7*

To monitor *SOD7* expression pattern during development, the *pSOD7:GUS* and *pSOD7:SOD7-GFP* vectors were constructed and transformed to wild-type plants, respectively. The tissue-specific expression patterns of *SOD7* were examined using a histochemical assay for GUS activity. In seedlings, relatively higher GUS activity was detected in younger leaves than in older leaves (Figures 4A -C). In flowers, GUS activity was observed in sepals, petals, stamens and carpels (Figures 4D-K). GUS activity was stronger in younger floral organs than in older ones (Figures 4D-K). Expression of *SOD7* was also detected in ovules (Figure.4L). Thus, these analyses indicate that *SOD7* is a temporally and spatially expressed gene. As *SOD7* encodes a B3 domain transcriptional repressor, we speculated that *SOD7* is localized in the nucleus. To determine subcellular localization of *SOD7*, we observed GFP inflorescence in *pSOD7:SOD7-GFP* transgenic plants. As shown in Figures 4M-0,

GFP signal was only detected in nuclei. We also expressed a GFP-SOD7 fusion protein under the control of the 35S promoter in wild-type plants. Transgenic lines overexpressing *GFP-SOD7* formed smaller seeds than the wild type (Figure 3D), indicating that the *GFP-SOD7* fusion protein is functional. As shown in Figures 4P-R, GFP fluorescence in *35S:GFP-SOD7* transgenic plants was exclusively observed in nuclei. Thus, these results show that SOD7 is a nuclear-localized protein.

SOD7/NGAL2 acts redundantly with NGAL3 to control seed size

In order to further investigate the function of SOD7 in seed size control, we isolated T-DNA inserted loss-of-function mutants for *SOD7* and *NGAL3*, the most closely related family member. *sod7-ko1* (SM_3_34191) was identified with T-DNA insertion in the first exon of the *SOD7* gene (Figure 5A). *ngal3-ko1* (SM_3_36641) had T-DNA insertion in the first exon of the *NGAL3* gene (Figure 5B). The T-DNA insertion sites were confirmed by PCR using T-DNA specific and flanking primers and sequencing PCR products. *sod7-ko1* and *ngal3-ko1* mutants had no detectable full-length transcripts of *SOD7* and *NGAL3*, respectively. Seeds from *sod7-ko1* and *ngal3-ko1* mutants were slightly larger and heavier than seeds from wild-type plants (Figures 5C, G and H). The cotyledon area of *sod7-ko1* and *ngal3-ko1* mutants was increased, compared with that of the wild type (Figure 5I). Considering that SOD7 shares the highest similarity with *NGAL3*, we speculated that SOD7 may act redundantly with *NGAL3* to influence seed size. To test this, we generated the *sod7-ko1 ngal3-ko1* double mutant. As shown in Figures 5C, D, G and H, the seed size and weight phenotypes of *sod7-ko1* mutant were synergistically enhanced by the disruption of *NGAL3*, indicating that SOD7 functions redundantly with *NGAL3* to control seed size. We further measured the cotyledon area of 10-d-old seedlings. A synergistic enhancement of cotyledon size of *sod7-ko1* by the *ngal3-ko1* mutation was also observed (Figure 5I). In addition, the *sod7-ko1 ngal3-ko1* double mutant formed larger leaves and flowers than their parental lines (Figures 5E and F; 11). Thus, these results indicate that SOD7 and *NGAL3* act redundantly to control seed and organ growth.

SOD7 acts maternally to control seed size

As the size of a seed is determined by the zygotic and/or maternal tissues (Garcia et al., 2005; Xia et al., 2013; Du et al., 2014), we asked whether SOD7 functions maternally or zygotically. We therefore performed reciprocal cross experiments between the wild type and *sod7-ko1 ngal3-ko1*. The effect of *sod7-ko1 ngal3-ko1* on seed size was observed only when *sod7-ko1 ngal3-ko1* was used as maternal plants

(Figure 6A). The size of seeds from *sod7-ko1 ngal3-ko1* plants pollinated with wild-type pollen was similar to that from the self-pollinated *sod7-ko1 ngal3-ko1* plants (Figure 6A). By contrast, the size of seeds from wild-type plants pollinated with *sod7-ko1 ngal3-ko1* mutant pollen was similar to that from the self-pollinated wild-type plants (Figure 6A). These results indicate that *sod7-ko1 ngal3-ko1* acts maternally to influence seed size. We further investigated the size of Col-O/Col-O F2, Col-O/*sod7-ko1 ngal3-ko1* F2, *sod7-ko1 ngal3-ko1*/Col-O F2 and *sod7-ko1 ngal3-ko1* F2 seeds. As shown in Figure 6B, *sod7-ko1 ngal3-ko1* F2 seeds were larger than wild-type seeds, while the size of Col-O/*sod7-ko1 ngal3-ko1* F2 and *sod7-ko1 ngal3-ko1*/Col-O F2 seeds was similar to that of wild-type seeds. Thus, these results indicate that the embryo and endosperm genotypes for SOD7 do not determine seed size, and SOD7 is required in the sporophytic tissue of the mother plant to control seed growth.

SOD7 regulates cell proliferation in the maternal integuments

The reciprocal crosses showed that SOD7 functions maternally to influence seed size. The integuments surrounding the ovule are maternal tissues, which could set the growth potential of the seed coat after fertilization. Consistent with this idea, several studies showed that the integument size influences the final size of seeds in *Arabidopsis* (Garcia et al., 2005; Schruff et al., 2006; Adamski et al., 2009; Xia et al., 2013; Du et al., 2014). We therefore asked whether SOD7 acts through the maternal integuments to determine seed size. To test this, we characterized mature ovules of the wild type and *sod7-ko1 ngal3-ko1*. As shown in Figures 6C and D, the *sod7-ko1 ngal3-ko1* ovules were obviously larger than wild-type ovules. The outer integument length of *sod7-ko1 ngal3-ko1* ovules was significantly increased, compared with that of wild-type ovules (Figure 6E). As the size of the integument is determined by cell proliferation and cell expansion, we examined the number and size of outer integument cells in wild-type and *sod7-ko1 ngal3-ko1* ovules. As shown in Figure 6F, the number of outer integument cells in *sod7-ko1 ngal3-ko1* ovules was increased, compared with that in wild-type ovules. By contrast, the length of outer integument cells in *sod7-ko1 ngal3-ko1* ovules was similar to that in wild-type ovules (Figure 6G). These results showed that SOD7 is required for cell proliferation in the maternal integuments of ovules. After fertilization, cells in the integument mainly undergo expansion but still have division. We further examined the number and size of outer integument cells in wild-type and *sod7-ko1 ngal3-ko1* seeds at 6 and 8 day after pollination (DAP). In wild-type seeds, the number of outer integument cells at 6 DAP was comparable with that at

8 DAP (Figure 6F), indicating that cells in the outer integuments of wild-type seeds completely stop dividing by 6 DAP. Similarly, cells in the outer integuments of *sod7-ko1 ngal3-ko1* seeds also cease division by 6 DAP. The number of outer integument cells in *sod7-ko1 ngal3-ko1* seeds was significantly increased, compared with that in wild-type seeds (Figure 6F). By contrast, the length of outer integument cells in *sod7-ko1 ngal3-ko1* seeds was not increased in comparison to that in wild-type seeds (Figure 6G). Thus, these analyses indicate that *SOD7* is required for cell proliferation in the maternal integuments of ovules and developing seeds.

SOD7 acts in a common pathway with *KLU* to control seed size, but does so independently of *DM*

The Arabidopsis *klu* mutants formed small seeds due to the decreased cell proliferation in the integuments, while plants overexpressing *KLU/CYP78A5* produced large seeds as a result of the increased cell proliferation in the integuments (Adamski et al., 2009), suggesting that *SOD7* and *KLU* could function antagonistically in a common pathway to control seed growth. To test for genetic interactions between *SOD7* and *KLU*, we generated the *klu-4 sod7-ko1 ngal3-ko1* triple mutant and measured the size of seeds from wild-type, *klu-4*, *sod7-ko1 ngal3-ko1* and *klu-4 sod7-ko1 ngal3-ko1* plants. As shown in Figures 7A and B, the average size and weight of *klu-4 sod7-ko1 ngal3-ko1* seeds were similar to those of the *klu-4* single mutant, indicating that *klu-4* is epistatic to *sod7-ko1 ngal3-ko1* with respect to seed size and weight. We further investigated the mature ovules from wild-type, *klu-4*, *sod7-ko1 ngal3-ko1* and *klu-4 sod7-ko1 ngal3-ko1* plants. The outer integument length of *klu-4 sod7-ko1 ngal3-ko1* ovules was comparable with that of *klu-4* ovules (Figure 7C). Similarly, the outer integument length of *klu-4 sod7-ko1 ngal3-ko1* seeds was indistinguishable from that of *klu-4* seeds at 8 DAP (Figure 7C). In addition, the size of *klu-4 sod7-ko1 ngal3-ko1* petals was similar to that of *klu-4* petals).

Thus, these genetic analyses show that *klu-4* is epistatic to *sod7-ko1 ngal3-ko1* with respect to seed and organ size, indicating that *SOD7* and *KLU* act antagonistically in a common pathway to control seed and organ growth. To further understand the cellular basis of epistatic interactions between *SOD7* and *KLU*, we investigated the outer integument cell number of ovules and developing seeds from wild-type, *klu-4*, *sod7-ko1 ngal3-ko1* and *klu-4 sod7-ko1 ngal3-ko1* plants. The number of outer integument cells in *klu-4 sod7-ko1 ngal3-ko1* ovules was similar to that in *klu-4* ovules (Figure 7D). Similarly, the number of outer integument cells in *klu-4 sod7-ko1 ngal3-ko1* seeds was

comparable with that in *klu-4* seeds (Figure 7D). These results indicate that *klu-4* is epistatic to *sod7-ko1 ngal3-ko1* with respect to the number of outer integument cells. We also observed that cells in the outer integuments of *klu-4* and *klu-4 sod7-ko1 ngal3-ko1* seeds were slightly longer than those in wild-type seeds, suggesting a possible compensation mechanism between cell proliferation and cell expansion. Together, these findings show that SOD7 functions antagonistically in a common pathway with KLU to control cell proliferation in the maternal integuments.

Considering that *sod7-1D* was identified as a suppressor of *da 1-1* in seed size, we further asked whether SOD7 and DA1 could act in the same genetic pathway. To test this, we measured the size of wild-type, *da 1-1*, *sod7-1D* and *sod7-1D da 1-1* seeds. The genetic interaction between *sod7-1D* and *da 1-1* was essentially additive for seed size, compared with that of *sod7-1D* and *da 1-1* single mutants, indicating that SOD7 might function independently of *DM* to control seed size. We further crossed *sod7-ko1 ngal3-ko1* with *da 1-1* and generated the *sod7-ko1 ngal3-ko1 da 1-1* triple mutant and measured its seed size. The genetic interaction between *sod7-ko1 ngal3-ko1* and *da 1-1* was also additive for seed size, compared with their parental lines, further supporting that SOD7 functions to control seed growth separately from *DM*.

SOD7 directly binds to the promoter of *KLU* and represses the expression of *KLU*

Considering that SOD7 acts antagonistically in a common pathway with *KLU* to control seed size, we asked whether the transcription repressor SOD7 could repress the expression of *KLU*. We therefore investigated the expression of *KLU* in the chemically-inducible SOD7 (*pER8-SOD7*) transgenic plants. After the *pER8-SOD7* transgenic plants were treated with the inducer (β -estradiol), the expression of SOD7 was strongly induced at 4 and 8 hours (Figure 8A). As expected, the expression of *KLU* was dramatically repressed at 4 and 8 hours (Figure 8A). Thus, these results indicate that SOD7 represses the expression of *KLU* and also suggest that *KLU* might be a direct target of SOD7.

To determine whether SOD7 can directly bind to the promoter of the *KLU* gene, we performed a chromatin immunoprecipitation (ChIP) assay with 35S:*GFP* and 35:*GFP-SOD7* transgenic plants. It has been reported that the CACCTG sequence is recognized by the B3 domain of RAV1, one member of the RAV family (Kagaya et al., 1999; Yamasaki et al., 2004). We therefore analyzed the promoter sequence of *KLU* and did not find an intact CACCTG sequence within 2 kb promoter region of *KLU*.

However, we found a similar sequence (CACTTG) in the promoter region of *KLU* (Figure 8B), which could be the potential SOD7-binding site. To test this, we examined the enrichment of a *KLU* promoter fragment (PF1) containing the CACTTG sequence by ChIP analyses and found that the fragment PF1 was strongly enriched in the chromatin-immunoprecipitated DNA with anti-GFP antibody (Figures 8B and C). By contrast, we did not detect significant enrichment of an ACTIN7 promoter sequence and the *KLU* promoter fragment PF2, which do not contain the CACTTG sequence (Figures 8B and C). This result shows that SOD7 associates with the promoter of *KLU* in vivo. We further expressed SOD7 as a MBP fusion protein (MBP-SOD7) and performed the DNA electrophoretic mobility shift assays (EMSA). As shown in Figures 8B and D, MBP-SOD7 was able to bind to the biotin-labeled probe A containing the CACTTG sequence, and the binding was reduced by the addition of an unlabeled probe A. By contrast, MBP-SOD7 failed to bind to a probe A-m with mutations in the CACTTG sequence (Figures 8B and D). Taken together, these results show that SOD7 directly binds to the promoter of *KLU* and represses *KLU* expression.

Discussion

Seed size is crucial for plant fitness and agricultural purposes, but little is known about the genetic and molecular mechanisms that set the final size of seeds in plants. In this study, we show that SOD7 acts maternally to control seed size by restricting cell proliferation in the integuments of ovules and developing seeds. SOD7 encodes a B3 domain transcriptional repressor NGAL2 and acts redundantly with its closest homolog NGAL3 to control seed size. Genetic analyses indicate that SOD7 functions in a common pathway with the maternal factor *KLU* to control seed growth, but does so independently of DA1. Further results reveal that SOD7 directly binds to the promoter region of *KLU* and represses *KLU* expression. Thus, our findings identify SOD7 as a negative factor for seed size and define the genetic and molecular mechanisms of SOD7 and *KLU* in seed size control.

SOD7 acts maternally to regulate seed size

The *sod7-1D* gain-of-function mutant was identified as a suppressor of the large seed phenotype of *da 1-1*. However, genetic analyses showed that SOD7 functions independently of DA1 to control seed growth. The *sod7-1D* single mutant produced small seeds and organs (Figure 2), while the simultaneous disruption of SOD7 and the closely related family member NGAL3 resulted in large seeds and organs (Figure 5), indicating that SOD7 is a negative regulator of seed and organ size. Several previous

studies suggest that there is a possible link between seed size and organ growth. For instance, *arf2*, *da1-1*, *da2A* and *eod3-1D* mutants produced large seeds and organs (Schruff et al., 2006; Li et al., 2008; Fang et al., 2012; Xia et al., 2013), whereas *klu* and *sod2/ubp15* mutants formed small seeds and organs (Anastasiou et al., 2007; Adamski et al., 2009; Du et al., 2014). However, seed size is not invariably associated with organ size. For example, *eod8/med25* mutants with large organs formed normal-sized seeds (Xu and Li, 2011), while *ap2* mutants with normal-sized organs produced large seeds (Jofuku et al., 2005; Ohto et al., 2005). Thus, these findings suggest that seeds and organs not only share common mechanisms but also possess distinct pathways to control their respective size.

Reciprocal cross experiments showed that *SOD7* acts maternally to restrict seed growth, and the endosperm and embryo genotypes for *SOD7* do not determine seed size (Figure 6). The integuments surrounding the ovule are maternal tissues and form the seed coat after fertilization. Arabidopsis *arf2*, *ap2*, *da1-1*, *da2-1* and *eod3-1D* mutants with large integuments formed large seeds (Jofuku et al., 2005; Ohto et al., 2005; Schruff et al., 2006; Li et al., 2008; Fang et al., 2012; Xia et al., 2013), while *klu-4* and *ubp15/sod2* mutants with small integuments produced small seeds (Adamski et al., 2009; Du et al., 2014), indicating that the maternal integuments are crucial for determining seed size in Arabidopsis. Consistent with this notion, mature *eod7-ko1 ngal3-ko1* ovules were larger than wild-type ovules (Figures 6C and D). The outer integument length of *eod7-ko1 ngal3-ko1* ovules and developing seeds was significantly increased, compared with that of wild-type ovules and seeds (Figures 6E and 7C). Considering that the maternal integument or seed coat not only acts as a protective structure but also restricts seed growth, the regulation of maternal integument size is one of important mechanisms for seed size control. The size of the integument is determined by cell proliferation and cell expansion; these two processes are assumed to be coordinated. The number of outer integument cells in *sod7-ko1 ngal3-ko1* ovules and seeds was significantly increased, compared with that in wild-type ovules and seeds (Figure 6F), indicating that *SOD7* controls seed growth by limiting cell proliferation in the maternal integuments. Similarly, several mutants with the increased number of cells in the maternal integuments produced large seeds in Arabidopsis (Schruff et al., 2006; Li et al., 2008; Xia et al., 2013). By contrast, several other mutants with the decreased number of cells in the maternal integuments formed small seeds in Arabidopsis (Adamski et al., 2009; Du et al., 2014). Considering that cells in the integuments mainly undergo expansion after fertilization (Garcia et al.,

2005), it is possible that the number of cells in the integuments determines the growth potential of the seed coat after fertilization.

The genetic and molecular mechanisms of *SOD7* and *KLU* in seed size control

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The *sod7-1D* mutant had small seeds and organs (Figure 2), as had been seen in *klu* mutants (Anastasiou et al., 2007; Adamski et al., 2009). *KLU* encodes a cytochrome P450 CYP78A5 that has been proposed to generate mobile plant-growth substances (Anastasiou et al., 2007; Adamski et al., 2009). *KLU* regulates seed size by promoting cell proliferation in the maternal integuments of ovules (Anastasiou et al., 2007; Adamski et al., 2009). By contrast, *SOD7* acts maternally to control seed size by limiting cell proliferation in the integuments of ovules and developing seeds (Figure 6). These results suggest that *SOD7* could function antagonistically in a common pathway with *KLU* to control seed size. In our growth conditions, *klu-4* formed slightly smaller seeds than the wild type due to the decreased cell number and the slightly increased cell length in the integuments of developing seeds (Figures 7A and D), suggesting a possible compensation mechanism between cell proliferation and cell expansion in *klu-4* integuments. Importantly, our genetic analyses showed that *klu-4* is epistatic to *sod7-ko1 ngal3-ko1* with respect to seed and organ size (Figures 7A and B). *klu-4* is also epistatic to *sod7-ko1 ngal3-ko1* for the outer integument length (Figure 7C). Further results revealed that the number of cells in the outer integuments of *klu-4 sod7-ko1 ngal3-ko1* ovules and developing seeds was similar to that of *klu-4* ovules and developing seeds (Figure 7D). Thus, these genetic results demonstrate that *SOD7* act in a common pathway with *KLU* to control seed size by regulating cell proliferation in the maternal integuments.

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SOD7 encodes a B3 domain transcriptional repressor NGAL2 that is localized in nuclei of Arabidopsis cells (Figures 4M-R). Thus, it is possible that *SOD7* could directly bind to the promoter of *KLU* and repress *KLU* expression. Supporting this idea, the inducible expression of *SOD7* resulted in a strong reduction of *KLU* expression (Figure 8A). Our ChIP-qPCR data showed that *SOD7* associates with the promoter region of *KLU* in vivo (Figures 8B and C). EMSA experiments revealed that *SOD7* directly binds to the CACTTG sequence in the promoter of the *KLU* gene (Figures 8B and D). Thus, these results illustrate that *SOD7* directly targets the promoter region of *KLU* and represses the expression of *KLU*, thereby determining seed size. Taken together, these findings

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reveal the genetic and molecular mechanisms of *SOD7* and *KLU* in regulating Arabidopsis seed size.

For many plants, the seeds are the main product to be harvested, and an increase in seed size would be beneficial for growers. In this study, we identify *SOD7* as a negative regulator of seed size, and demonstrate that *SOD7* acts in a common genetic pathway with *KLU* to control seed size. Our current knowledge of *SOD7* functions suggests that the *SOD7* gene (and its homologs in other plant species) could be used to engineer large seed size in crops. Considering that crop plants have undergone selection for large seed size during domestication (Fan et al., 2006; Song et al., 2007; Gegas et al., 2010), it will be a worthwhile challenge to know whether beneficial alleles of the *SOD7* gene have already been utilized by plant breeders.

Knockout experiments in rice using genome editing

Genome editing experiments to knock out os1 1g01560000 and/or Os12g0157000 in rice are being carried out using the crispr-cas9 system. Four vectors, each with two recognition (CRISPR target) sites, have been constructed, to achieve these knock outs, as described in Fig. 14. In summary, the vectors were obtained as follows:

1. The target sites were identified. The target site should be (or approximately so) 20 nucleotides before a NGG sequence, N being for any nucleotide. The target sequence was then evaluated using the website: <http://cbi.hzau.edu.cn/crispr/help.php> (incorporated herein by reference). Of note, the target site should be unique in the genome.

2. Using overlap PCR, the target sequence is linked with the U6 sequence, as shown in Figure 14. U6 is for transcriptional activity.

3. Using infusion technology we connected the U6-guide-gRNA scaffold fragment to the vector pMDC99-cas9 to obtain the pMDC99-cas9- U6-guide-gRNA scaffold constructs. These constructs were named zyy1, zyy2, zyy3, zyy4. The full sequences of these constructs are represented in SEQ ID NO: 155, 156, 157 and 158 respectively. Each construct contains two recognition sites, which are highlighted in the sequence information, and are represented separately as SEQ ID Nos 159, 160, 161, 162 and 163.

4. We then transformed these constructs into Agrobacteria and used an Agrobacteria mediated method to transform rice and obtain gene-edited rice.

Transformation of plants is a routine technique that is well known to the skilled person. Nonetheless, a brief outline of transformation techniques is provided above.

Knock out lines are being analysed to assess the phenotype.

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Table 1. Primers used in this study

Primer Name Primer Sequences

Primers for T-DNA identification

SM_3_34191-LP ACCATGACATTCGAGGTTTAC (SEQ ID NO. 8)

10 SM_3_34191-RP ATCACCACCAAAACGACGTAG (SEQ ID NO. 9)

SM_3_36641-RP TACGTCATGCTTCAAATCGTG (SEQ ID NO. 10)

SM_3_36641-RP AGGACACGAACAATTCATTCG (SEQ ID NO. 11)

Spm32 TACGAATAAGAGCGTCCATTTTAGAGTGA (SEQ ID NO. 12)

SM_3_39145- LP ACCCAAAGAACAGCAATCATG (SEQ ID NO. 13)

15 SM_3_39145- RP AAAACACTCCGCCATTAAACC (SEQ ID NO. 14)

Primers for TAIL-PCR

OJF22 CGAGTATCAATGGAACTTAACCG (SEQ ID NO. 15)

OJF23 AACGGAGAGTGGCTTGAGAT (SEQ ID NO. 16)

OJF24 TGGCCCTTATGGTTTCTGCA (SEQ ID NO. 17)

20 AD1 NTCGA(G/C)T(A/T)T(G/C)G(A/T)GTT (SEQ ID NO. 18)

Primers for Constructs

SOD7CDS-F ATGTCAGTCAACCATTACCAC (SEQ ID NO. 19)

SOD7CDS-R CAGGTAGGAGATGGACGAGGTTGA (SEQ ID NO. 20)

SOD7G-F TGAGAGGAACCATTCTTAGAGG (SEQ ID NO. 21)

25 SOD7G-R ACCTCGTCCATCTCCTACCTGC (SEQ ID NO. 22)

SOD7P-F AAACACGTCAAATATAACGAAT (SEQ ID NO. 23)

SOD7P-R CTTTTTTTTGGTTTCTTGGAGTGAGAGAGAG (SEQ ID NO. 24)

SOD7-ER-F AGTCTGGGCCCATGTCAAGTCAACCATTAC (SEQ ID NO. 25)

SOD7-ER-R GCGACTAGTTTATAAAAGAGTTAAATTA (SEQ ID NO. 25)

30 MBP-SOD7-FP CGGGATCCTCAGTCAACCATTACC (SEQ ID NO. 27)

MBP-SOD7-RP ACTAGTCGACTCAACCTCGTCCATCTCC (SEQ ID NO. 28)

Primers for RT-PCR and qRT-PCR

ACTIN2-F GAAATCACAGCACTTGCACC (SEQ ID NO. 29)

ACTIN2-R AAGCCTTTGATCTTGAGAGC (SEQ ID NO. 30)

35 SOD7-EX-F GCGACGACGGAGAAAGGG (SEQ ID NO. 31)

SOD7-EX-R ACGACGGCGCCATAGTGT (SEQ ID NO. 32)

NGAL3-EX-F TTTGAAGACGAGTCAGGCAAGT (SEQ ID NO. 33)

NGAL3-EX-R TACGGCGGCTCCATAGTGGG (SEQ ID NO. 34)

SOD7-q-FP GTATTGGAGCGGCTTGACTACACC (SEQ ID NO. 35)

SOD7-q-RP GACGGCATCACCATGACATTCG (SEQ ID NO. 36)

5 KLU-q-FP TGATTCTGACATGATTGCTGTTCT (SEQ ID NO. 37)

KLU-q-RP TCGCAACTGTATCTGTCCCTCTA (SEQ ID NO. 38)

Primers for ChIP assay

ACTIN7-ChIP-FP CGTTTCGCTTTCCTTAGTGTTAGCT (SEQ ID NO. 29)

ACTIN7-ChIP-RP AGCGAACGGATCTAGAGACTCACCTTG (SEQ ID NO. 40)

10 PF1-F CAGGCCTAAGCCTAACAGTAGAC (SEQ ID NO. 41)

PF1-R TGTACTAGGATTTATTTACGTAG (SEQ ID NO. 42)

PF2-F TATTGTTTCATAGAAACCCTGCAAA (SEQ ID NO. 43)

PF2-R AGTCAATGGTTTAATGGCGGAGTG (SEQ ID NO. 44)

Probes for EMSA

15 A-Biotin-FP TTCTACTACACTTGCTCTCTGTA (SEQ ID NO. 45)

A-Biotin-RP TACAGAGAGCAAGTGTAGTAGAA (SEQ ID NO. 46)

A-Biotin-m-FP TTCTACTAACACCTCTCTCTGTA (SEQ ID NO. 47)

A-Biotin-m-RP TACAGAGAGAGGTGTTAGTAGAA (SEQ ID NO. 48)

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Identity of homologs to NGAL2 is indicated

AtSOD7 nucleic acid SEQ ID NO. 1 (cDNA) At3g1 1580

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 ATGTCAGTCAACCATTACCACAACACTCTCTCGTTGCATCATCACCACCAAAACGA
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 15 GAGCGGCTTGACTACACCTTATCGTCAAGTACACGCGTCAACTACTTACCCTAATA
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 TGTCATGGTGATGCCGTCGAGCCACCACCGCGTCCTGATGTCTATAATGACCAAC
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 20 TGGAGCAGGTAGGAGATGGACGAGGTTGA

AtSOD7 nucleic acid SEQ ID NO. 2 (genomic DNA).

ttgttcggctatttgttatactattgttataacagtcacaagacttgacctcaacgaaaactttacaaaacgtgaattggaaa
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25 tatactcctctctgttctcttttaagattgtcttttataaaaaatagatgattcgtaattgtattgcataatttacatgttctcttaaaa
aaagtaatagagattaatattttatgcatggatttttagattatctgcctactttatatggtagtaacaagaacattcatctttatt
tggttttataaacaataatgagaatttttaagggttagggcaagcacttgaaagctcaaccatttttagttagctgggtggaa
tatctttctataaaaagcaaatgagttatctaaaactatatgacaatttttagttgctgtgtaattgtatataaaaataacaac
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30 atccggtacatgcgctgaggagaaccgtccaatccacttagactaacgtgccctttatttctccttttaattctatgttaaaaa
aacaatttaactaaaagatgcgacgtgtcttgacggtggaaaaaaattgtagGCGCCGTCGTTGATCATG
CTCAGTCGATACCACCGGTGGTCGCAGGTAGCTCGAGGACGGTGAGGCTTTTTG
GCGTGAACCTCGAATGTCATGGTGATGCCGTCGAGCCACCACCGCGTCCTGATG
TCTATAATGACCAACACATTTACTATTACTCAACTCCTCATCCCATGgtaaaatattttttttt
35 acattttgtcagattcaaattttgtcttacgtatgatataattattaaacagatgtcgtggctgtttctcgagacgagacagatg
aaaattagtaattttaaaatagacctgaaagagattttatgtttaataaattatataaaggaggaatcagagagaataata

ctatacacttgactgtaaaaccacatggccaatttggttttatttgattactttgattgtttgtttactctttgtctctgtagcctcct
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cgtcagttttttgtgaaaccaaattattgactaataagctggaaagcaaaactgactaaaagcattacaaacttatcaatg
acataagttttgaattattaccatgtttgtaatgttcagatataattgaaatgcttagaattatatattgtataacttaaattaatg
5 aaataaagtgaataactaaagatagttttattttcatattattctatacaattcgggtgtacaattgttttgatgataataaaaaata
ataaaaattgcgtgttggaattgtgaaacagAATATATCATTTGCTGGGGAAGCATTGGAGCAGGT
AGGAGATGGACGAGGT

AtNGAL2 SEQ ID N0.3 (protein encoded by *AtSOD7*)..

10 MSVNHYHNTLSLHHHHQNDVAIAQRESLFEKSLTPSDVGKLNRLVIPKQHAKEYFPLN
NNNNNGGSGDDVATTEKGMLLSFEDESGKCKWKFRYSYWNSSQSYVLTKGWSRYVK
DKHLDAGDWFFQRRHFDLHRLFIGWRRRGEASSPAVSVVSQEALVNTTAYWSGL
TTPYRQVHASTTYPNIHQEYSHYGAVVDHAQSIPPWAGSSRTVRLFGVNLECHGDA
15 VEPPPRPDVYNDQHIYYSTPHPMNISFAGEALEQVGDGRG

AtNGAL3 nucleic acid sequence SEQ ID NO. 4 (cDNA) at5g06250

ATGTCAGTCAACCATTACTCCACAGACCACCACCACACTCTCTTGTGGCAGCAAC
20 AGCAACACCGCCACACCACCGACACATCGGAGACAACCACCACCGCCACATGGC
TCCACGACGACCTAAAAGAGTCACTCTTCGAGAAGTCTCTCACACCAAGCGACGT
CGGGAAACTCAACCGCCTCGTCATACCAAAACAACACGCAGAGAAATACTTCCCT
CTCAATGCCGTCCTAGTCTCCTCTGCTGCTGCTGACACGTCATCTTCGGAGAAAG
GGATGCTTCTAAGCTTTGAAGACGAGTCAGGCAAGTCATGGAGGTTTCAGATACTC
25 TTAAGTGAACAGCAGTCAAAGCTATGTCTTGACTAAAGGATGGAGCAGATTTGTCA
AAGACAAACAGCTCGATCCAGGCGACGTTGTTTTCTTCCAACGACACCGTTCTGA
TTCTAGGAGACTCTTCATTGGCTGGCGCAGACGTGGACAAGGCTCCTCATCCTCC
GTCGCGGCCACTAACTCCGCCGTGAATACGAGTTCTATGGGAGCTCTTTCTTATC
ATCAAATCCACGCCACTAGTAATTACTCTAATCCTCCCTCTCACTCAGAGTATTCC
30 CACTATGGAGCCGCCGTAGCAACAGCGGCTGAGACTCACAGCACACCGTCGTCT
TCCGTCGTCGGGAGCTCAAGGACGGTGAGGCTTTTCGGTGTGAATCTGGAGTGT
CAAATGGATGAAAACGACGGAGATGATTCTGTTGCAGTTGCCACCACCGTTGAAT
CTCCCGACGGTTACTACGGCCAAAACATGTACTATTATTACTCTCATCCTCATAAC
ATGGTAATTTTAACTCTTTTATAA

35

AtNGAL3 amino acid SEQ ID NO.5

MSVNHYSTDHHHTLLWQQQQHRHTTDTSETTTTATWLHDDLKESLFEKSLTPSDVG
 KLNRLVIPKQHAKEYFPLNAVLVSSAAADTSSSEKGMLLSFEDES GKSWRFRYSYWN
 SSQSYVLTKGWSRFVKDKQLDPGDWFFQRHRSDSRRLFIGWRRRGQGSSSSVAAT
 5 NSAVNTSSMGALSYHQIHATSNYSNPPSHSEYSHYGA AVATAAETHSTPSSSWGSS
 RTVRLFGVNLECQMDENDGDDSVAVATTVESPDGYYGQNMYYYYSHPHNMVILTLL

Oryza sativa

10 Osl2g0157000 LOC_Osl2g06080.1
 Cover 73% identity 53%

SEQ ID NO: 49

15 MAMHAGHAWWGVAMYTNNHYHHYRHKTS DVGKNRVKHARYGGGDSGKGS DSGKWRRYSYWTSSSYVTKG
 WSRVYVKRDAGDVVHRVRGGAADRGCRRRGSAAAVRVTANGGWSMCYSTSGSSYDTSANSYAYHRSVDDHSD
 HAGSRADAKSSSAASARRRGVND CGADATAMYGYMHHSYAAVSTVNYWSV

CDS SEQ ID NO: 50

20 ATGCCCATGCACCCTCTCGCCCAGGGGCACCCCCAGGCGTGCCCATGGGGTGTAGCCATG
 TACACCAACCTGCACTACCACCACCACTACGAGAGGGAGCACCTGTTGAGAAGCCGCTG
 ACGCCGAGCGACGTGGCAAGCTCAACAGGCTGGTGATCCCCAAGCAGCACGCCGAGAGG
 TACTTCCCCTCGGCGGCGGCGACTCCGGTGAGAAGGGCCTCCTCTCTCTTCGAGGAC
 GAGTCCGGCAAGCCATGGCGGTTCCGCTACTCCTACTGGACCAGCAGCCAGAGCTACGTG
 25 CTCACCAAGGGCTGGAGCCGCTACGTCAAGGAGAAGCGCCTCGACGCCGCGCAGCTCGTC
 CACTTCGAGCGCGTCCGCGGCTCGGCGCCGCGCCGACCGCCTCTTCATCGGCTGCAGGCGC
 CGCGGCGAGAGCGCGCCGCGCCGCGCCGCGCGCTTCGCGTCACGCCGCGAGCCGCTGCC
 CTCAACGGCGGCGAGCAGCAGCCGTGGAGCCCAATGTGTTACAGCACGTGCGGGCTCGTCC
 TACGACCCTACCAGCCCTGCCAATTCATATGCCTACCATCGCTCCGTAGACCAAGATCAC
 30 AGCGACATACTACACGCAGGAGAGTCGAGAGAGAAGCAGACGCCAAGAGCAGCAGCGCG
 GCGTCGGCGCCGCGCGCTCGAGGCGGCTCAGGCTGTTGCGCGTTAACCTCGACTGCGGC
 CCGGAGCCGAGGCGGATCAGGCGACGGCAATGTACGGCTACATGCACCACCAGAGCCCC
 TACGCCGCAGTGTCTACAGTGCCAAATTACTGGTCAGTATTTTTTCAGTTTTAA

35 Oslg0156000
 LOC_Oslg05740.1
 Cover 81% identity 47%

40 SEQ ID NO: 51
 MAM NHPLFSQEQPQSWPWGVAMYAN FHYHHHYEKEHMFELTPSDVGKLNRLVIPKQHA
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 VVHFERVRGSFGVGDRLFIGCRRRGDAAAQTPAPPPAVRVAPAAQNAGEQQPWSPMCYS
 TSGGGSYPTSPANSYAYRRAADHDHGDMMHHADES PRDTSFSA GSAPSRRRLRFGVNL
 45 DCGPEPEADTTAAATMYGYMHQQSSYAAMSAVPSYWGNS

CDS SEQ ID NO: 52

ATGCCCATGAACCACCCTCTCTTCTCCCAGGAGCAACCCAGTCTGGCCATGGGGTGTG
 GCCATGTACGCCAACTTCCACTACCACCACCACTACGAGAAGGAGCACATGTTTGAGAAG

CCCCTGACGCCAGTGACGTGGGGAAGCTGAACCGGCTGGTGATCCCCAAGCAGCACGCC
 GAGAGGTACTTCCCCCTCGGCGCCGCGCAGCGCCGACAAGGGCCTGATCCTGTCTGTTT
 GAGGACGAGGCCGCGCGCCGTGGCGTTTCAGGTACTCCTACTGGACGAGCAGCCAGAGC
 TACGTGCTCACCAAGGGCTGGAGCCGCTACGTCAAGGAGAAGCGCCTCGACGCCGGCGAC
 5 GTCTGCACTTCGAGAGGGTGC GCGGCTCCTTCGGCGTCGGCGACCGTCTCTTCATCGGC
 TGCAGGCGCCGCGCGCAGCGCCGCCGCGCAAACACCCGCACCGCCGCCGCCGTGCGC
 GTCGCCCCGGCTGCACAGAACGCCGCGCAGCAGCAGCCGTGGAGCCCAATGTGTTACAGC
 ACGTCGGGCGGCGGCTCATACCCTACCAGCCCAGCCAACTCCTACGCCTACCGCCGCGCA
 GCAGATCATGATCACGGGGACATGCACCATGCAGACGAGTCTCCGCGCGACACGGACAGC
 10 CCAAGCTTCAGTGACGGCTCGGCGCCATCGAGGCGGCTCAGGCTGTTTCGGCGTCAACCTC
 GACTGCGGGCCAGAGCCGAGGCAGACACCACGGCAGCGGCAACAATGTACGGCTACATG
 CACCAGCAGAGCTCCTATGCTGCCATGTCTGCAGTACCCAGTTACTGGGGCAATTCATAA

 15
 os02g0683500 LOC_Os02g45850
 Cover 47% identity 62%

 20 SEQ ID NO: 53
 MEFTTSSRFSKEEEDDEQDEAGRREIPFMTATAEAPAPTSSSSSPAHHAASASASASAS
 GSSTPFRSDDGAGASGSGGGGGGGGAEVVEKEHM FDKVVTSPDVGKLNRLVI PKQYAEK
 YFPLDAAAN EKGLLLNFEDRAGKPWRFRYSYWNSSQSVMKGSRFVKEKRLDAGDTVS
 FSRGIGDEAARHLRFI DWKRRADTRDPLRLPRGLPLPMLTSHYAPWGIGGGGGFFVQPS
 25 PPATLYEHLRLRQLDFRAFNPAAAMGRQVLLFGSARI PPQAPLLARAPSPLHHHYTLQPS
 GDGVRAAGSPVVLDSVPVI ESPTTAAKRVRFLFGVNLDNPHAGGGGGAAAGESSN HGNAIS
 LQTPAWM RRDPTLRLLLELPPH HHHGAESSAASSPSSSSSSSKRDAHSALDLDL

 CDS SEQ ID NO: 54
 30 ATGGAGTTCACTACAAGCAGTAGGTTTTCTAAAGAAGAGGAGGACGAGGAGCAGGATGAG
 GCGGGAAGGCGAGAGATCCCTTTCATGACGCCACGGCCGAAGCCGCGCCTGCGCCCACG
 TCGTCGTCGTCGTCCTGCTCATCACGCGGCTTCCGCGTCGGCGTCGGCGTCTGCGTCA
 GGGAGCAGCACTCCCTTTCGCTCCGACGATGGCGCCGGGGCGTCTGGAGCGGCGGGCGG
 GCGGCGGCGGCGGAGAAAGCGGAGGTGGTGGAGAAGGAGCACATGTTGACAAGGTGGTG
 35 ACGCCGAGCGACGTTGGGAAGCTGAACCGGCTGGTGATCCCGAAGCAGTACGCCGAGAAG
 TACTTCCCCTGGACGCGGCGGCGAACGAGAAGGGCCTCCTGCTCAACTTCGAGGACCGC
 GCGGGGAAGCCATGGCGGTTCCGCTACTCCTACTGGAACAGCAGCCAGAGCTACGTGATG
 ACCAAGGGGTGGAGCCGCTTCGTCAAGGAGAAGCGCCTCGACGCCGGGGACACCGTCTCC
 TTCTCCCGCGGCATCGGCGACGAGGCGGCGCGGCACCGCCTTTCATCGACTGGAAGCGC
 40 CGCGCCGACACCCGCGACCCGCTCCGGCTGCCCGCGGGCTGCCGCTCCCGATGCCGCTC
 ACGTCGCACTACGCCCCGTGGGGGATCGGCGGCGGAGGGGGATTCTTCGTGCAGCCCTCG
 CCGCCGGCCACGCTCTACGAGACCGCCTCAGGCAAGGCCTCGACTTCCGCGCCTTCAAC
 CCCGCCGCCGCGATGGGAGGCAGGTCTCTGTTTCGGCTCGGCGAGGATTCTCCGCAA
 GCACCACTGCTGGCGCGCGCGCCGTGCGCGCTGCACCACCACTACACGCTGCAGCCGAGC
 45 GGCGATGGTGTAAAGGGCGGCGGGCTCACCGGTGGTGTCTGACTCGGTTCCGGTCATCGAG
 AGCCCCACGACGGCCGCGAAGCGCGTGC GGCTGTTTCGGCGTGAACCTCGACAACCCGCAT
 GCCGGCGGCGGCGGCGGCGCGCCGCCGCGGCGAGTCGAGCAATCATGGCAATGCACTGTCA
 TTGCAGACGCCCCGCTGGATGAGGAGGGATCCAACACTGCGGCTGCTGGAATTGCCCTCCT
 CACCACCACCATGGCGCCGAGTCGTCCGCTGCATCGTCTCCGTCGTCTGCTCTCCTCC
 50 AAGAGGGACGCGCATTCGGCCTTGATCTCGATCTGTAG

os04g0581400 LOC_Os04g49230
Cover 46% identity 64%

CDS SEQ ID NO: 55

5 ATGGAGTTTGCTACAACGAGTAGTAGGTTTTCCAAGGAAGAGGAGGAGGAGGAAGGG
GAACAGGAGATGGAGCAGGAGCAGGATGAAGAGGAGGAGGAGGCGGAGGCCTCGCCCCGC
GAGATCCCCCTTCATGACGTCGGCGGCGGCGGCGGCCACCGCTCATCGTCCTCCCCGACA
TCGGTCTCCCCCTTCGCCACCGCTTCCGCGGCGGCGTCCACGTCGGCGTCGGGCTCTCCC
TTCCGGTGCAGGCGACGGTGCGGGAGCGTCGGGGAGTGCGGCGGCGGCGTGCGGGCAGGAC
10 GTGGAGGTGATCGAGAAGGAGCACATGTTTCGACAAGGTGGTGACGCCGAGCGACGTGGGG
AAGCTGAACCGGCTGGTGATCCCGAAGCAGCAGCGCGAGAAGTACTTCCCGCTGGACTCG
GCGGCGAACGAGAAGGGCCTTCTCCTCAGCTTCGAGGACCGAACCGGCAAGCTATGGCGC
TTCCGCTACTCCTACTGGAACAGCAGCCAGAGCTACGTCATGACCAAGGGTTGGAGCCGC
TTCGTCAAGGAGAAGCGCCTCGACGCCGGGACACCGTCTCCTTCTGCCGCGGCGCCGCC
15 GAGGCCACCCGCGACCGCCTTTCATCGACTGGAAGCGCCGCGCCGACGTCCGCGACCCG
CACCGCTTCCAGCGCCTACCGCTCCCCATGACCTCGCCCTACGGCCCCGTGGGGCGGCGGC
GCGGGCGCTTTCATGCCGCCCGCGCCGCCCGCCACGCTCTACGAGCATCACCGCTTTC
GCCAGGGCTTCGACTTCCGCAACATCAACCCCGCTGTGCCGCGAGGCAGCTCGTCTTCT
TCGGCTCCCCAGGGACGGGGATTATCAGCACCCGCCCTTGCCACCGCCGCCGTCGCCAC
20 CTCCGCCTCTCACTCACTCCACATTACGGTGCACACCCGAGCCCCGTAG

SEQ ID NO: 56

M EFATTSSRFSKEEEEEEGEQEMEQQDEEEEEAEASPREIPFMTSAAAAATASSSPT
SVSPSATASAAASTSASGSPFRSSDGAGASGSGGGGGEDVEVIEKEH MFDKVVTPSDVG
25 KLNRLVIPKQHAKEYFPLDSAANEKGLLLSFEDRTGKLWRFYSYWNSSQSYVMTKGWSR
FVKEKRLDAGDVTVFCRGAAEATRDRLFI DWKRRADVRDPHRFQRLPLPMTSPYGPWGGG
AGASSCRPRRPRRSTITAFARASTSATSTPLCRRGSSSSSAPQGRGFISTRPCHRRRRH
LRLLTNSTLRCTTRAP

30

os03g0120900 LOC_Os03g02900
Cover 47% identity 63%

CDS SEQ ID NO: 57

35 ATGGAGTTCATCACGCCAATCGTGAGGCCGGCATCGGCGGCGGCGGGCGGCGGCGAGGTG
CAGGAGAGTGGTGGGAGGAGCTTGCGGCGGCTGGAGAAGGAGCACATGTTGACAAGGTG
GTGACGCCGAGCGACGTGGGGAAGCTGAACCGGCTGGTGATCCCGAAGCAGCAGCGGAG
AAGTACTTCCCGCTGGACGCGGCGTCCAACGAGAAGGGGCTCCTGCTCAGCTTCGAGGAC
CGCACGGGGAAGCCATGGCGGTTCCGCTACTCCTACTGGAACAGCAGCCAGAGCTACGTG
40 ATGACCAAGGGGTGGAGCCGCTTCGTCAAGGAGAAGCGACTCGACGCCGGGACACCGTC
TCCTTCGGCCGCGCGTCGCGCAGGCCGCGCGCGGGAGGCTTTCATCGACTGGCGCCGC
CGCCCCGACGTGTCGCGCGCTCCAGCCGCCACGCACCGCTTCGCCCACCACCTCCCT
TCCTCCATCCCCCTTCGCTCCCTGGGCGCACCAACACGGACACGGAGCCGCCGCCGCCGCC
GCCGCCGCCGCCGCGCCAGGTTTCTCCTGCCTCCCTCCTCGACTCCCATCTACGACCAC
45 CACCGCCGACACGCCACGCCGTGCGGTACGACGCGTACGCCGCGGCCACCAGCAGGCAG
GTGCTGTTCTACCGGCCGTTGCCGCCGAGCAGCAGCATCATCCCGCGGTGGTGCTGGAG
TCGGTGCCGGTGCGCATGACGGCGGGGCACGCGGAGCCGCCGTCGGCTCCGTGAAGCGA
GTTCCGGCTGTTCCGGGTGAACCTCGACTGCGCGAATTCCGAACAAGACCACGCCGGCGTG
GTCGGGAAGACGGCGCCGCCGCTGCCATCGCCGCCGTCATCATCGTCATCTTCTCTCC
50 GGGAAAGCGAGGTGCTCCTTGAACCTTGAACCTTGTGA

SEQ ID NO: 58

5 M EFITPIVRPASAAAGGGEVQESGGRSLAAVEKEHM FDKVVTPSDVGKLNRLVI PKQHAE
KYFPLDAASNEKGLLLSFEDRTGKPWFRFRYSYWNSSQSYVMTKGWSRFVKEKRLDAGDTV
SFGRGVGEAARGRLFDWRRRPDVVAALQPPTHFAHHLPSI PFAPWAHHHGHGAAAAA
AAAAGARFLLPPSSTPIYDH HRRHAHAVGYDAYAAATSRQVLFYRPLPPQQH HPAVVLE
SVPVRMTAGHAEPSPAPSKRVRLFVNLDCANSEQDHAGVVGKTAPPPLPSPSSSSSSS
GKARCSLN LDL

10

os01g0693400

Cover 47% identity 63%

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CDS SEQ ID NO: 59

ATGGACAGCTCCAGCTGCCTGGTGGATGATACCAACAGCGCGGCTCGTCCACGGACAAG
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GCGAGCGCGGTGGTGGACGCGGCCGAGCCTGGCGCGGAGGCGGACTCCGGGTCCGGGGGA
CGTGTGTGCGGCGGCGGCGGCGGCGGTGCCGGCGGTGCGGGAGGGAAGCTGCCGTCTGCC
20 AAGTTCAAGGGCGTCTGTGCCGACGCCAACGGGAGGTGGGGCGCGCAGATCTACGAGCGG
CACCAGCGGGTGTGGCTCGGCACGTTCCGCCGGGAGGACGACGCCGCGCGCGCCTACGAC
GTCCGCCGCGCAGCGCTTCCGCGGCCGCGACGCCGTACCAACTTCCGCCCGCTCGCCGAG
GCCGACCCGGACGCCGCCGCGGAGCTTCGTTCTCGCCACGCGCTCCAAGGCCGAGGTC
GTGACATGCTCCGCAAGCACACCTACTTCGACGAGCTCGCGCAGAGCAAGCGCACCTTC
25 GCCGCCTCCACGCCGTGGCCGCGACCAACCGCCTCCCTCTCAACGGCCACCTCTCG
TCGCCCCGCTCCCCCTTCGCGCCCGCCGCGGCGCGGACCACTGTTGACAAGACGGTC
ACCCGAGCGACGTGGGCAAGCTGAACAGGCTCGTCATACGAAGCAGCACGCCGAGAAG
CACTTCCCGCTACAGCTCCCGTCCGCCGCGGCGGAGAGCAAGGGTGTCTCTCACTTC
GAGGACGCCGCCGCAAGGTGTGGCGGTTCCGGTACTCGTACTGGAACAGCAGCCAGAGC
30 TACGTGCTAACCAAGGGCTGGAGCCGCTTCGTCAAGGAGAAGGGTCTCCACGCCGGCGAC
GTCGTGGCTTCTACCGCTCCGCCGCCAGTGCCGGCGACGACGGCAAGCTCTTCATCGAC
TGCAAGTTAGTACGGTCGACCGGCGCCGCCCTCGCGTCGCCCCGCTGATCAGCCAGCGCCG
TCGCCGGTGAAGGCCGTGAGGCTCTTCGGCGTGACCTGCTACGGCGCCGGCGCCGGTC
GAACAGATGGCCGGGTGCAAGAGAGCCAGGACTTGGCGGCGACGACGCCTCCACAAGCG
35 GCGGCGTTCAAGAAGCAATGCATAGAGCTGGCACTAGTATAG

SEQ ID NO: 49

60M DSSSCLVDDTNSGGSSTDKLRALAAAAAETAPLERMSGASAVVDAAEPGAEDSGSGG
40 RVCGGGGGGAGGAGGKLPSSKFKGVVPQPNGRWGAQIYERHQRVWLGTGAGEDDAARAYD
VAAQFRFRGDAVTNFRPLAEADPDAEAELRFLATRSKAEVDM LRKHTYFDELAQSKRTF
AASTPSAATTTASLSNGHLSSPRSPFAPAAARDHLDKTVTPSDVGKLN RLVI PKQHAEK
HFPLQLPSAGGESKGVLLNFEDAAGKVWFRFRYSYWNSSQSYVLTGWSRFVKEKGLHAGD
VVGFYRSAASAGDDGKLFI DCKLVRSTGAALASPADQPAPSPVKAVRLFVLDLLTAPAPV
45 EQMAGCKRARDLAATTPPQAAAFKKQCI ELALV

50

Osl0g0537100

LOC_Osl0g39190

Cover 47% identity 60%

CDS SEQ ID NO: 61

ATGGAGTTCACCCCAATTTGCCGCCGACGAGGGTCGCCGGCGGTGAGGAGGATTCCGAG

5 AGGGGGGCGGCGGCGTGGGCGGTGGTGGAGAAGGAGCACATGTTTGAGAAGGTCGTGACG
 CCGAGCGACGTGGGGAAGCTGAACCGATTGGTCATCCCCAAGCAGCACGCCGAGAGGTAC
 TTCCCGCTCGACGCCGCGGCGGCGCGCGCGCGGTGGTGGCGGCGGTGGCGGCGGC
 GGGGGGAAGGGGCTGGTGCTGAGCTTCGAGGACAGGACGGGGAAGGCGTGAGGTTCCGG
 10 TACTCGTACTGGAACAGCAGCCAGAGCTACGTGATGACCAAAGGGTGGAGCCGCTTCGTC
 AAGGAGAAGCGCCTCGGCGCCGCGGACACCGTGTCTGTCGGCCGCGGCCTCGGCGACGCC
 GCGCGCGGCCCTCTTCATCGACTTCCGCCGCCGCCGAGGACGCCGCGCAGCTTCATG
 TTCCCGCCGACGGCGGCGCGCGCGTCCGCACTCGCACCACCATCATCAGCGACACCACCCG
 CCGTCCCCGTCCGTGCCCTTTGCCCGTGGCGAGACTACACCACCGCCTATGGCGGCGGC
 15 TACGGCTACGGCTACGGCGGCGGCTCCACCCCGGCGTCCAGCCGCCACGTGCTGTTCTC
 CGGCCGACAGTGCCGCGCGCTGTGGTGTCAAGTCGGTCCGCGTGCACGTCGCGGCCACC
 TCGGCGGTGCAGGAGGCGGCGACGACGACAAGGCCGAAGCGTGTCCGGCTGTTCCGGGTG
 AACCTCGACTGCCCCGGCGGCCATGGACGACGACGACATCGCCGAGCGGCGAGCCGG
 ACGGACGCGTCGTCTCTCTGACGCTCCCCTCGCCGTCGTCTCGACGTCGTCTCGACG
 GCGGGGAAGAAGATGTGCTCCTTGATCTTGGGTTGTGA

20 SEQ ID NO: 62
 MEFTPIPPTRVAGGEEDSERGAAWAVVEKEH MFEKVVTPSDVGKLNRLVI PKQHAERY
 FPLDAAAGAGGGGGGGGGGGGKGLVLSFEDRTGKAWRFRYSYWNSSQSYVMTKGWSRFV
 KEKRLGAGDVSFGRGLDAARGRLFIDFRRRRQDAGSFMFPPTAAPPSHHHHQRHHP
 PLPSVPLCPWRDYTTAYGGGYGYGYGGGSTPASSRHVFLRPQVPAAVVLKSVPVHVAAT
 25 SAVQEAATTPRKRVRFLGVNLDCPAAMDDDDDIAGAASRTAASSLLQLPSPSSSTSST
 AGKKMCSLDLGL

Glycine max

30 L.OC100795470
 Cover 75% identity 53%

35 SEQ ID NO: 63
 Msinhysmdlpeptlwwphphhqqqltmdpdlrlnlnsddgngndndndenqtttgeqeilddkepmf^ kpltpsdvlglnr
 lviqkhaekyfpplsgdsggseckgllsfedesgkcwrfryswnssqsyvltkgwsryvdkrldagdvvlferhrvdaqrfigwrrrrqsd
 aalppahvsrrsgggdgsnknegwtrgfysahpypthlhqhpy spyqqqhdcldhagrgsqgqnqmrpvgnnsssssssrvlrl
 fgvdmeccpehdsgpstpqcsynsnmlpstqgtdhshnfyyqqpsnsnpsphhmmvhhqpyyy

40 CDS SEQ ID NO: 64
 ATGTCCATAAACCACTACTCCATGGACCTTCCCGAACCGACACTCTGGTGGCCACACCCA
 CACCACCAACAACAACAACTAACCTTAATGGATCCTGACCCTCTCCGTCTCAACCTCAAT
 AGCGACGATGGCAATGGCAATGACAACGACAACGACGAAAAATCAAACAACCACAACAGGA
 GGAGAACAAAGAAATATTAGACGATAAAGAACCGATGTTTCGAGAAGCCCTTAACCCCGAGC
 45 GACGTGGGGAAGCTGAACCGTCTCGTAATCCCGAAGCAGCACGCGGAGAAGTACTTCCCA
 CTGAGTGGTGA CTGCGGGCGGGAGCGAGTGCAAGGGGCTGTTACTGAGTTTCGAGGACGAG
 TCGGGGAAGTGTG GCGCTTCCGCTACTCGTACTG GAACAG CAGCCAGAGCTACGTGCTC
 ACCAAAGGGTGGAGCCGCTACGTCAAGGACAAGCGCCTTGACGCGGGCGACGTCGTTTTG
 TTCGAGCGTCACCGCGTCGACGCGCAGCGCTCTTCATCGGGTGGAGGCGCAGGCGGCAG
 50 AGCGATGCCGCTTGCCGCTGCGCACGTTAGCAGTAGGAAGAGTGGTGGTGGTATGGG
 AATAGTAATAAGAATGAGGGGTGGACCAGAGGGTTCTATTCTGCGCATCATCCTTATCCT
 ACGCATCATCTTCATCATCATCAGCCCTCGCCATACCAACAACAACATGACTGTCTTCAT

GCAGGTAGAGGGTCCCAAGGTCAGAACCAAGGATGAGACCAGTGGGAAACAACAGTTCT
AGCTCTAGTTCGAGTTCAAGGGTACTTAGGCTGTTGCGGGTTCGACATGGAATGCCAACCC
GAACATGATGATTCTGGTCCCTCCACACCCCAATGCTCCTACAATAGTAACAACATGTTG
CCATCAACACAGGGCACAGATCATTCCCATCACAATTTCTACCAACAGCAACCTTCTAAT
5 TCCAATCCTTCCCCTCATCACATGATGGTACATCACCAACCATACTACTACTAG

LOC100818164

Cover 50% identity 73%

10

SEQ ID NO: 65

MSTNHYTM DLPEPTLWWPHPHQQQLTLI DPDPLPLNLN NDDNDNGDDNDNDENQVT
TGGEIEI INNKEPMFEKPLTPSDVGKLNRLVIPKQHAKEYFPLSGGDSGSSECKGLLLSF
EDES GK CWRFRYSYWNSSQSYVLTKGWSRYVKDKRLDAGDVVLFQRHRADAQRLFIGWRR
15 RRQSDALPPPAHVSSRKSGGDGNSKNEGDVGVGWTRGFYPAHHPYPTHHHHPSPYHHQQ
DDSLHAVRGSQQQNQRTRPVGNSSSSSSSSRVLRVLFVNMECQPEHDDSGPSTPQCSYN
TNNI LPSTQGTDI HSHLNFYQQQTSNSKPPPHH MMI RHQPYYY

20

SEQ ID NO: 66

ATGTCGACAAACCACTACACCATGGACCTTCCCGAACCAACACTCTGGTGGCCACACCCA
CACCAACAACAATAACCTTAATAGATCCAGACCCTCTCCCTCTGAACCTCAACAACGAC
GACAACGACAATGGCGACGACAACGACAACGACGAAAACCAACAGTTACAACAACCACA
ACAGGAGGAGAAGAAGAAATAATAACAATAAAGAACCGATGTTTCGAGAAGCCGTAACC
25 CCGAGCGACGTGGGGAAGCTGAACCGCCTCGTAATCCCGAAGCAGCACGCTGAGAAGTAC
TTTCCACTGAGTGGTGGTGACTCGGGCAGTAGCGAGTGCAAGGGGCTGTTACTGAGTTTC
GAGGACGAGTCGGGGAAGTGCTGGCGCTTCCGCTACTCGTACTGGAACAGCAGCCAGAGC
TACGTGCTCACCAAGGGTGAGCCGTTACGTGAAGGACAAGCGCCTCGATGCGGGAGAT
GTCGTTTTATTCCAGCGCCACCGCGCCGACGCGCAGCGCCTCTTCATCGGCTGGAGGCGC
30 AGGCGGCAGAGCGACGCCCTGCCGCCGCTGCGCACGTTAGCAGCAGGAAGAGTGGTGGT
GATGGGAATAGTAGTAAGAATGAGGGTGATGTGGGCGTGGGCTGGACCAGAGGGTTCTAT
CCTGCGCATCATCCTTATCCTACGCATCATCATCATCCCTCGCCATACCATCACCAACAA
GATGACTCTTTCATGCAGTTAGAGGGTCCCAAGGTCAGAACCAAGGACGAGACCAGTG
GGAAACAG CAGTTCTAGTTCGAGTTCGAGTTCAAGG GTACTTAGGCTATTCGGGTCAAC
35 ATGGAATGCCAACCCGAACATGATGATTCTGGACCCTCCACACCCCAATGCTCCTACAAT
ACTAACAACATATTGCCATCCACACAGGGCACAGATATTCATTCCCATCTCAATTTCTAC
CAACAACAACAACTTCTAATTCCAAGCCTCCCCCTCATCACATGATGATACGTACCAAA
CCATACTACTACTAG

40

LOC100802734

Cover 77% identity 53%

45

SEQ ID NO: 67

MSIINHYPSETTLYWTNDQQQQAAMWLSNSHTPRFN LNDEEEEEEDDVIVSDKATNNLTQ
EEEKVAMFEKPLTPSDVGKLNRLVIPKQHAKEH FPLDSSAAKGLLLSFEDES GK CWRFRY
SYWNSSQSYVLTKGWSRYVKDKRLHAGDVVLFHRHRSPLQRRFFISCSRRQPNPVP
TRSSASFYSAHPPYPAHHFFPYQPHSLHAPGGGSQQQNETTPGGNSSSSSGSRVLRVLF
50 VNMECQPDNHN DSQNSTPECSYTHLYHHQTSSYSSSSNPHHHMVPPQP

SEQ ID NO: 68

5
 10
 15

ATGTCATCGATAAACCACTATTACCGGAAACAACACTATACTGGACCAACGACCAACAG
 CAACAAGCCGCCATGTGGCTGAGTAATTCACACCCCCGCGTTTCAATCTGAACGACGAG
 GAGGAGGAGGAGGAAGACGACGTTATCGTTTCGGACAAGGCTACTAATAACTTGACGCAA
 GAGGAGGAGAAGGTAGCCATGTTTCGAGAAGCCGTTGACGCCGAGCGACGTCGGAAGCTG
 AACCGGCTCGTGATTCCGAAACAGCACGCGGAGAAGCACTTCCCTCTCGACTCGTCGGCG
 GCGAAGGGGCTGTTGCTGAGTTTCGAGGACGAGTCCGGGAAGTGTTGGCGCTTCCGTTAC
 TCTTATTGGAACAGTAG ccAGAGTTACGTTTTGACCAAAG GATGGAGCCGTTACGTCAA
 GACAAACGCCTCCACGCTGGCGACGTCGTTTTGTCCACAGACACCGCTCCCTCCCTCAA
 CGCTTCTTCATCTCCTGACGCCGCCGAACCCAACCCGGTCCCCGCTCACGTTAGCACC
 ACCAGATCCTCCGCTTCTTCTACTCTGCGCACCCACCTTATCCTGCGCACCACTTCCCC
 TTCCCATACCAACCTCACTCTCTTCATGCACCAGGTGGAGGGTCCCAAGGACAGAACGAA
 ACGACACCGGGAGGGAACAGTAGTTCAAGTGGCAGTGGCAGGGTGCTGAGGCTCTTTGGT
 GTGAACATGGAATGCCAACCTGATAATCATAATGATTCCCAGAACTCCACACCAGAATGC
 TCCTACACCCACTTATACCACCATCAAACCTCTTCTTATTCTTCTTCAAACCTCAC
 CATCACATGGTACCTCAACAACCATAA

LOC100781489

Cover 49% identity 64%

20 SEQ ID NO: 69
M ELMQQVKGNYSDSREEEEEEAAITRESESSLHQQDTASNFGKKLDLMDLSLGSSKE
EEEEGNLQQGGGGVVHHAHQVVEKEHMFVKVATPSDVGKLNRLVIPKQHAEKYFPLDSST
NEKGLLLNFEDRNGKVWFRFYSYWNSSQSYVMTKGWSRFVKEKKLDAGDIVSFQRGLGDL
YRHRLYIDWKRRPDHAHAHPPH HHDPLFLPSI RLYSLPPTMPPRYHHDHFFH HHLNYYNNL
25 FTFQQHQYQQLGAATTTT HNNYGYQNSGSGSLYLRSSMSMGGGDQN LQGRGSNIVPMI I
DSVPVNVAAHHN NRHGNGGITSGGTNCSGKRLRLFGVN MECASSAEDSKELSSGSAAHVT
TAASSSSLHHQRLRVPVPVPLEDPLSSAAAAARFGDHKGASTGTSLFLDLPQLQYHRH

[illegible]

LOC100776987

Cover 46% identity 62%

5 SEQ ID NO: 71
MDAISCLDESTTTESLSISQAKPSSTIMSSEKASPSPPPPNRLCRVSGASAVVDSDDGGG
GGGSTEVESRKLPSISKYKGVVPQPNGRWGSQIYEKHQRVWLGTFN EEDEAARAYDVAVQR
FRGKDAVTNFKPLSGTDDDDGESEFLNSHSKSEIVDMLRKHTYNDELEQSKRSRGFVRRR
10 GSAAGAGNGNSISGACVM KAREQLFQKAVTPSDVGKLNRLVIPQHAEKHFPLQSAANGV
SATATAAKGVLLNFEDVGGKVWRFRYSYWNSSQSYVLTKGWSRFVKEKNLKAGDTVCFQR
STGPDRQLYIDWKTRNVVNEALFGPVVEPIQMVRLFGVNI LKLPGSDSIANNNNNASGCC
NGKRREM ELFSLECSKKPKIIGAL

15 CDS SEQ ID NO: 72
ATGGATGCAATTAGTTGCCTGGATGAGAGCACCAACCGAGTCACTCTCCATAAGTCAG
GCCAAGCCTTCTTCGACGATTATGTCGTCCGAGAAGGCTTCTCCTTCCCGCCGCCGCCG
AACAGGCTGTGCCGCGTCGGTAGCGGTGCTAGCGCAGTCGTGGATTCCGACGGCGGCGGC
GGGGGTGGCAGCACCGAGGTGGAGTCGCGGAAGCTCCCCTCGTCCAAGTATAAGGGCGTC
20 GTGCCCCAGCCCAACGGCCGCTGGGGCTCGCAGATTTACGAGAAGCACCGCGCGTGTGG
CTGGGAACGTTCAACGAGGAAGACGAGGCGGCGCGTGCCTACGACGTGCGCGTGACGCGA
TTCCGCGGCAAGGACGCCGTCACAACTTCAAGCCGCTCTCCGGCACCGACGACGACGAC
GGGGAATCGGAGTTTCTCAACTCGCATTCGAAATCCGAGATCGTCGACATGCTGCGTAAG
CATACTACAATGACGAGCTGGAACAAAGCAAGCGCAGCCGCGGCTTCGTACGTGCGCGC
25 GGCTCCGCCGCCGGCGCGCGGAAACGGAACCTCAATCTCCGGCGCGTGTGTTATGAAGGCG
CGTGAGCAGCTATTCCAGAAGGCCGTTACGCCGAGCGACGTTGGGAACTGAACCGTTTG
GTGATACCGAAGCAGCAGCGGAGAGCACTTTCTTTACAGAGCGCTGCTAACGGCGTT
AGCGCGACGGCGACGGCGGCGAAGGGCGTTTTGTTGAACTTGAAGACGTTGGAGGGAAA
GTGTGGCGGTTTCGTTACTCGTATTGGAACAGTAGCCAGAGTTACGTCTTGACCAAAGGT
30 TGGAGCCGGTTCGTTAAGGAGAAGAATCTGAAAGCCGGTGACACGGTTTGTGTTTCAACGG
TCCACTGGACCGGACAGGCAGCTTTACATCGATTGGAAGACGAGGAATGTTGTTAACGAG
GTCGCGTTGTTCGGACCGGTTGTGCAACCGATCCAGATGGTTCCGGCTCTTTGGTGTTAAC
ATTTTGAACTACCCGTTTCAGATTCTATCGCCAATAACAATAATGCAAGTGGGTGCTGC
AATGGCAAGAGAAGAAATGGAACCTTTTCATTAGAGTGTAGCAAGAAACCTAAGATT
35 ATTGGTGCTTTGTAG

Locl00778733

Cover 44% identity 64%

40 SEQ ID NO: 73
MELMQEVKGYSDGREEEEEEEAAEEI ITREESSRLLHQHQEAAGSNFI INNN HHHHQHH
HHHTTKQLDFMDLSLGSSKDEGNLQGSSSSVYAAH HHAASASSSANGN NNNSSSSNLQQQ
QQQPAEKEHM FDKVVTSDVGKLNRLVI PKQHAKEYFPLDSSANEKGLLLNFEDRNGKLW
45 RFRYSYWNSSQSYVMTKGWSRFVKEKKLDAGDMVSFQRGVGELYRHRLYI DWWRRPDH HH
HHH HGPDHSTTLFTPFLLI PNQPHH LMSI RWGATGRLYSLPSPTPPRHHEHLNYYNNAMYH
PFHHHGAGSGI NATTH HYNHYHEMSSTTTSGSAGSVFYHRSTPPISMPLADHQLNTRQQ
QQQQQQQEGAGNVSLSPMI IDSVPAHHLHHQHH HGGKSSGPSSTSTSPSTAGKRLRLFG
VNMECASSTSEDPKCFSLSSSSMANSNSQPPLQLLREDTLSSSSARFGDQRGVGEPSM L
50 FDLDPSLQYRQ

SEQ ID NO: 74

ATGGAGTTGATGCAAGAAAGTAAAGGGTATTCTGATGGCAGAGAGGAGGAGGAGGAGGAA
GAGGAAGCAGCAGAAGAAATCATCACAAGAGAAGAAAGCAGCAGGTTGTTACACCAGCAC
CAGGAGG CAGCAGGTTCCAATTTTCATCATCAACAATAATCATCATCATCAACATCAC
CACCACCACACAACAAAGCAGCTAGACTTCATGGACTTGTCACCTGGTAGCAGCAAGGAT
5 GAAGGGAATTTGCAAGGATCATCTTCTTCTGTCTATGCTCATCATCATGCAGCAAGT
GCTAGTTCTTCTGCCAATGGTAACAACAAC AACAG CAGCAGCAGCAACTTG CAGCAACAG
CAGCAGCAGCCTGCTGAGAAGGAGCACATGTTTGATAAAGTAGTGACACCAAGTGATGTG
GGGAAGCTGAACCGGTTGGTGATACCAAAGCAGCATGCTGAGAAGTATTTCCCTCTTGAT
TCCTCAGCCAATGAGAAGGGTCTGTTGCTGAATTTTGAGGACAGGAATGGTAAGTTGTGG
10 AGGTT CAGGTA CTCTATTGGAACAGCAGCCAGAGCTATGTGATGACCAAAGGTTGGAGC
CGTTTTGTAAAGGAGAAGAAGCTTGATGCTGGTGACATGGTGTCTTCCAGCGTGGTGTT
GGGGAGTTGTATAGGCATAGGTTGTACATAGATTGGTGGAGAAGGCCTGATCATCATCAC
CATCACCATCATGGCCCTGACCATTCAACCACACTCTTCACACCTTTCTTAATTCCTCAAT
CAGCCTCATCACTTAATGTCCATCAGATGGGGTGCCACTGGCAGATTGTA CTCTCCCTCCCT
15 TCCCCAACCCACCACGCCACCATGAACACCTCAATTACAACAATAACGCCATGTATCAT
CCCTTTCATCACCATGGTGCTGGAAGTGAATTAATGCTACTACTCATCACTACAACAAC
TATCATGAGATGAGTAGTACTACTACTTCAGGATCTGCAGGCTCAGTCTTTTACCACAGG
TCAACACCCCAATATCAATGCCATTGGCTGACCACCAAACCTTGAACACAAGGCAGCAG
CAACAACAACAACAACAAGAGGGAGCTGGCAATGTTTCTCTTTCCCTATGATCATT
20 GATTCTGTTCCAGTTGCTCACCACCTCCATCATCAACAACACCATGGTGGCAAGAGTAGT
GGTCCTAGTAGTACTAGTACTAGTCTAGCAGTGCAGGGAAAAGACTAAGGCTATTTGGG
GTCAACATGGAATGTGCTTCTTCAACATCAGAAGACCCCAATGCTTCAGCTTGTGTCC
TCATCTTCAATGGCTAATTCCAATTCACAACCACACTTCAGCTTTTGAGGGAAGATACA
CTTTTCGTCATCATCGGCAAGGTTTGGGGATCAGAGAGGAGTAGGGGAACCTTCAATGCTT
25 TTTGATCTGGACCCTTCTTTGCAATACCGGCAGTGA

LOC732601

Cover 44% identity 62%

30

SEQ ID NO: 75

MDGGCVTDETTTSSDSLSPPPSRVGSVASAVVDPDGCCVSGEASRKLPSKYKGVVPQ
PNGRWGAQIYEKHQRVWLGTNEEDEAARAYDIAALRFRGPDAVTN FKPPAASDDAESEF
LNSHSKF EIVDM LRKHTYDDELQQSTRGGRRRLDADTASSGVFDAKAREQLFEKTVTPSD
35 VGKLNRLVIPKQHAKEHFPLSGSGDESSPCVAGASAAKGMLLNFDVGGKVWRFRYSYWN
SSQSYVLTKGWSRFVKEKNLRAGDAVQFFKSTGPDRQLYDCKARSGEVNNNAGGLFVPI
GPVVEPVQMVRFLFVNLLKLPVPGSDGVGKRKEMELFAFECCCKLKVIGAL

CDS SEQ ID NO: 76

ATGGATGGAGGCTGTGTACAGACGAAACCACCACATCCAGCGACTCTCTTTCCGTTCCG
CCGCCCAGCCGCTCGGCAGCGTTGCAAGCGCCGTCGTCGACCCCGACGTTGTTGCGTT
TCCGGCGAGGCCGAATCCCGGAACTCCCTTCGTCGAAATACAAAGCGTGGTGCCGCAA
CCGAACGGTCGCTGGGGAGCTCAGATTTACGAGAAGCACCAGCGCGTGTGGCTCGGCACT
TTCAACGAGGAAGACGAAGCCGCCAGAGCCTACGACATCGCCGCGCTGCGCTTCCGCGGC
45 CCCGACGCCGTCACCAACTTCAAGCCTCCCGCCGCTCCGACGACGCCGAGTCCGAGTTC
CTCAACTCGCATTCCAAGTTCGAGATCGTCGACATGCTCCGCAAGCACACCTACGACGAC
GAGCTCCAGCAGAGCACGCGCGGTGGTAGGCGCCGCTCGACGCTGACACCGCGTCGAGC
GGTGTGTTTCGACGCGAAAGCGCGTGAGCAGCTGTTTCGAGAAAACGGTTACGCCGAGCGAC
GTCGGGAAGCTGAATCGATTAGTGATACCGAAGCAGCACGCGGAGAAGCACTTTCCGTTA
50 AGCGGATCCGGCGACGAAAGCTCGCCGTGCGTGCGGGGGCTTCGGCGGCGAAGGGAATG
TTGTTGAACTTTGAGGACGTTGGAGGGAAAGTGTGGCGGTTTCGTTACTCTTATTGGAAC
AGTAGCCAGAGCTACGTGCTTACCAAAG GATGGAGCCG GTTCGTTAAGGA GAAG AATCTT
CGAGCCGGTGACGCGGTTCAAGTTCGACCCGACCGGACCGGCAGCTATATATA

GACTGCAAGGCGAGGAGTGGTGAGGTTAACAATAATGCTGGCGGTTTGTGTTGTTCCGATT
GGACCGGTCTGTTAGCCGGTTCAGATGGTTCGGCTTTTCGGGGTCAACCTTTTGAACTA
CCCGTACCCGGTTCGGATGGTGTAGGGAAGAGAAAAGAGATGGAAGTGTGTCATTTGAA
TGTGCAAGAAGTTAAAAGTAATTGGAGCTTTGTAA

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Locl00801107

Cover 44% identity 61%

10

SEQ ID NO: 77

MDAISCMDSTTTESLSISLSPSSSEKAKPSSMITSSSEKVSLSPPPSNRLCRVSGGASA
VVDPDGGGSGAEVESRKLPSKYKGVVPQPNGRWGAQIYEKQVRVWLGTFNEDDEAARAY
DIAAQRFRGKDAVTNFKPLAGADDDGESEFLNSHSHKPEIVDM LRKHTYNDELEQSKRSR
GVVRRRGSAAAGTANSISGACFTKAREQLFEKAVTPSDVGKLNRLVIPKQHAKEHFPLQS
SNGVSATTIAAVTATPTAAKGVLN FEDVGGKVWRFYSYWNSSQSYVLTGWSRFVKEK
NLKAGDTCVCFH RSTGPDQKLYIDWKTRNVNNEVALFGPVGPVVEPIQMVRLEFGVNI LKL
PGSDTIVGN NNNASGCCNGKRREMELEFSLECSKKPKI IGAL

15

20

CDS SEQ ID NO: 78

ATGGATGCAATTAGTTGCATGGATGAGAGCACCACCACTGAGTCACTCTCTATAAGTCTT
TCTCCGACGTCATCGTCGGAGAAAGCGAAGCCTTCTTCGATGATTACATCGTCGGAGAAG
GTTTCTCTGTCCCCGCCGCCGTCAAACAGACTATGCCGTGTTGGAAGCGGCGCGAGCGCA
GTCGTGGATCCTGATGGCGGCGGCAGCGGCGCTGAGGTAGAGTCGCGGAAACTCCCCTCG
TCGAAGTACAAGGGCGTGGTGCCCCAGCCCAACGGCCGCTGGGGTGCGCAGATTTACGAG
AAGCACCAGCGCGTGTGGCTTGGAACGTTCAACGAGGAAGACGAGGCGGCGCGTGCCTAC
GACATCGCCGCGCAGCGGTTCCGCGGCAAGGACGCCGTACGAACTTCAAGCCGCTCGCC
GGCGCCGACGACGACGACGAGGAATCGGAGTTTCTCAACTCGCATTCCAAACCCGAGATC
GTCGACATGCTGCGAAAGCACACGTACAATGACGAGCTGGAGCAGAGCAAGCGCAGCCGC
GGCGTCGTCCGGCGGCGAGGCTCCGCCGCCGCCGCCACCGCAAACCTCAATTTCCGGCGCG
TGCTTTACTAAGGCACGTGAGCAGCTATTCGAGAAGGCTGTTACGCCGAGCGACGTTGGG
AAATTGAACCGTTTTGGTGATACCGAAG CAGCACGCGGAGAAG CACTTTCCGTTACAGAGC
TCTAACGGCGTTAGCGCGACGACGATAGCGGCGGTGACGCGGACGCCGACGGCGGCGAAG
GGCGTTTTGTGAACTTCGAAGACGTTGGAGGGAAAGTGTGGCGGTTTCGTTACTCGTAT
TGGAACAGTAGCCAGAGTTACGTCTTAACCAAAGGTTGGAGCCGTTTCGTTAAGGAGAAG
AATCTGAAAGCTGGTGACACGGTTTGTTCACCGGTCCACTGGACCGGACAAGCAGCTT
TACATCGATTGGAAGACGAGGAATGTTGTTAACAACGAGGTCGCGTTGTTCCGACCGGTC
GGACCGGTTGTCGAACCGATCCAGATGGTTCGGCTCTTTGGGGTTAACATTTTGAACTA
CCCGGTTTCAGATACTATTGTTGGCAATAACAATAATGCAAGTGGGTGCTGCAATGGCAAG
AGAAGAGAAAAGTGAAGTGTCTCGTTAGAGTGTAGCAAGAAACCTAAGATTATTGGTGCT
TTGTAA

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LOC100789009

Cover 44% identity 62%

45

SEQ ID NO: 79

M DGGSVTDETTTTNSLSVPANLSPPLSLVSGGATAVVYPDGCCVSGEASRKLPSKY
KGVVPQPNGRWGAQIYEKQVRVWLGTFNEDDEAARAYDIAAH RFRGRDAVTNFKPLAGAD
DAEAEFLSTHSKSEIVDM LRKHTYDNEQQSTRGGRRRRDAETASSGAFDAKAREQLFEK
TVTQSDVGKLNRLVIPKQHAKEHFPLSGSGGGALPCMAAAAGAKGMLLNFEDEVGGKVWRF
RYSYWNSSQSYVLTGWSRFVKEKNLRAGDAVQFFKSTGLDRQLYDCKARSGKVN NNA
GLFI PVGPVVEPVQMVRLFGVDLLKLPVPGSDGIGVCDGKRKEMELFAFECSSKLLKVI
GAL

50

SEQ ID NO: 80

5 ATGGATGGAGGCAGTGTACAGACGAAACCACCACAACCAGCAACTCTCTTTTCGGTTCGG
GCCAATCTATCTCCGCCGCTCTCAGCCTTGTCCGGCAGCGGCGCAACCGCCGTCGTCTAC
CCCAGCGTTGTTGCGTCTCCGGCGAAGCCGAATCCCGGAAACTCCCGTCTCGAAATAC
AAAGGCGTGGTGCCGCAACCGAACGGTCGTTGGGGAGCTCAGATTTACGAGAAGCACCAG
CGCGTGTGGCTCGGCACCTTCAACGAGGAAGACGAAGCCGCCAGAGCCTACGACATCGCC
10 GCGCATCGCTTCCGCGGCCGCGACGCCGTCACTAACTTCAAGCCTCTCGCCGGCGCCGAC
GACGCCGAAGCCGAGTTCTCAGCACGCATTCCAAGTCCGAGATCGTCGACATGCTCCGC
AAGCACACCTACGACAACGAGCTCCAGCAGAGCACCCGCGGCGGCAGGCGCCGCCGGGAC
GCCGAAACCGCGTCGAGCGGCGCGTTTCGACGCGAAGGCGCGTGAGCAGCTGTTTCGAGAAA
ACCGTTACGCAGAGCGACGTCGGGAAGCTGAACCGATTAGTGATACCAAAGCAGCACGCG
15 GAGAAGCACTTTCGTTAAGCGGATCCGGCGGCGGAGCCTTGCCGTGCATGGCGGCGGCT
GCGGGGGCGAAGGGAATGTTGCTGAACTTTGAGGACGTTGGAGGGAAAGTGTGGCGGTTTC
CGTTACTCGTATTGGAACAGTAGCCAGAGCTACGTGCTTACCAAAGGATGGAGCCGGTTC
GTTAAGGAGAAGAATCTTCGAGCTGGTGACGCGGTTCAAGTCTTCAAGTCGACCGGACTG
GACCGGCAACTATATAGACTGCAAGGCGAGGAGTGGTAAGGTTAACAATAATGCTGCC
20 GGTTTGTATTCCGGTTGGACCGTTGTTGAGCCGTTTCAGATGGTACGGCTTTTCGGG
GTCGACCTTTTGAACTACCCGTACCCGTTTCGGATGGTATTGGGGTTGGCTGTGACGGG
AAGAGAAAAGAGATGGAGCTGTTTGCAATTTGAATGTAGCAAGAAGTTAAAAGTAATTGGA
GCTTTGTAA

25 LocI02660503
Cover 36% identity 57%

SEQ ID NO: 81

30 migvekvcticmrievntekgralmdcwqisgvhessdcseikfadvvkrarheennaaqkfgvvsqqngnwgai yahqqriwl
gtfkseraamaydsasiklrsgchmfpwndqvtqepqfshysaetvlnmirdgtypskfatflktrtqkgvakhighlgddeeefcct
qlfqkelpsdvglnrlvipkxhavsypvyvggsadesgsdvdeavfydklmrlwkfrycywkssqsyvfrgwnrfvdkklkakdviafft
wgksgegeafalidviynnnaeedskgdkvlgnglqlagseegededanigkdfnaqkglrlfgyvit

35 CDS SEQ ID NO: 82

atgattggagttgagaaagtgaacaattgtatgagaatagaggtgaatactgaaaagggaagaagggtttaatggactgttgcaaatatcag
gagttcatgaaagttcagattgtatgcgaaatcaaattgcattcgacgcagtagtaaaacgcgcgaggtcatgaagagaataatgcagcagcac
agaagttcaaaggcgttgtgtctcaaaaaatgggaactgggtgcacagatatatgcacaccagcagagaatctggttggggaccttcaaat
40 ctgaaagagaggctgcaatggcttatgacagcgccagcataaaactagaagcggagagtgccacagaaacttccatggaacgaccaaaca
gttcaagagcctcagttcgaagccattacagcgagaaacagtgctaaacatgattagagatggcacctatccatcaaaattgtacatttctc
aaaactcgtaaacccaaaaaggcgttgcaaacacataggtctgaagggtgatgacgaggaacagtttggcaccacaaacttttcagaagg
aattaacaccaagtgtatgtggcaagctcaacaggctgtcatccaaagaagcatgcagttagctatttcttacgttgggtgagtgctgatg
agagtggtagtgtgacgtggaggctgtgtttatgacaaactatgcgattgtggaagttccgatactgtattggaagagcagccaaagttacg
45 tgttcaccagaggctggaatcggttgtgaaggataagaagttgaaggctaaagatgtcattgcgtttttacgtggggaaaaagtgaggaga
gggagaagcgtttgcattgatcgatgaattataataataatgcagaagaagacagcaaggaggagacaccaacaagtttgggaaccaatta
caattagctggcagtgagaaggtgaagatgaagatgcaaacattggaaaggatttcaatgcacaaaagggtctgaggctcttgggtgtgtga
tcacctaa

50 Hordeum vulgare

MLOC_66387

Cover 47% identity 64%

SEQ ID NO: 83

5 M EFTATSSRFSKGEVEVEEQEEASMREIPFMTCAAATCAAAPPSASASASTPASASGSS
PPFRSGDDAGASGSGAGDGSRSNVAEAVEKEHM FDKVVTSPDVGKLNRLVI PKQYAEKYF
PLDSAANEKGLLLNFEDSAGKPWRFRYSYWNSSQSYVMTKGWSRFVKEKRLDAGDTVFSF
RGAGEAARHRLFI DWKRRADTRDPLRLPLPLM PLTSHYSPWGLGAGARGFFM PPSPPA
TLYEHRLRQGDFRGMNPSYPTMGRQVI LFGSAARM PPHGPAPLLVPRPPPPLHFTVQQQ
10 GSDAGGSVTAGSPVVLDSVPVI ESPTTATKKRVRLFVNLDNPQHPGDGGGESSNYGSAL
PLQMPASAWRPRDHTLRLLEFSPSHGAEASSPSSSSSSSKREAHSGLDLDL

SEQ ID NO: 84

15 ATGGAGTTTACTGCGACAAGCAGTAGGTTTTCTAAAGGAGAGGAGGAGGTGGAGGAGGAG
CAGGAGGAGGCGTCGATGCGCGAGATCCCTTTTCATGACGCCCGCGGCCGCCACCTGCGCC
GCGGCGCGCCTTCTGCTTCTGCGTCGGCCTCGACACCCGCGTCAGCGTCTGGAAGTAGC
CCTCCCTTTTCGATCTGGGGATGACGCCGAGCGTCGGGGAGCGGGGCCGCGACGGCAGC
CGCAGCAACGTGGCGGAGGCCGTGGAGAAGGAGCACATGTTGACAAAGTGGTGACGCCG
20 AGCGACGTGGGGAAGCTTAACCGGCTGGTCATCCCCAAGCAGTACGCCGAGAAGTACTTC
CCGCTGGACTCGGCGGCCAACGAGAAGGGCCTTCTGCTCAACTTCGAGGACAGCGCCGGG
AAG CCATGGCGCTCCGCTATTCTACTGGAACAGCAGCCAGAGCTACGTCATGACCAA
GGCTGGAGCCGCTTCGTAAGGAGAAGCGCCTCGACGCTGGGGACACCGTCTCCTTCTCC
CGCGGCGCCGGTGAGGCCGCGCGCCACCGCCTTTCATCGACTGGAAGCGCCGAGCCGAC
25 ACCAGAGACCCGCTCCGCTTGCCCCGCTCCCGCTCCCGATGCCGCTGACGTGCGACTAC
AGCCCGTGGGGCCTCGGCGCCGCGGCCAGAGATTCTTCATGCCTCCCTCGCCGCCAGCC
ACGCTCTACGAGCACCGTCTCCGTCAAGGCTTCGACTTCGCGGCATGAACCCAGTTAC
CCCACAATGGGAGACAGGTCATCCTTTTCGGCTCGGCCGCCAGGATGCCTCCGCACGGA
CCAGCACCACTCCTCGTGCCGCGCCCGCCGCCGCGCTGCACTTCACGGTGCAGCAACAA
GGCAGCGACGCCGCGGAAGTGTAACCGCAGGATCCCCAGTGGTGCTCGACTCAGTGCCG
30 GTAATCGAAAGCCCCACGACGGCAACGAAGAAGCGCGTGCGCTTGTTGCGCGTGAACCTG
GACAACCCGCGAGCATCCCGGTGATGGCGGGGGCGAATCGAGCAATTATGGCAGTGCACTG
CCATTGAGATGCCCGCATCAGCATGGCGGCCAAGGGACCATACGCTGAGGCTGCTCGAA
TTCCCTCGCACGGTGCCGAGGCGTCTCTCCATCGTCGTCGCTCTTCAAGAGGGAG
GCGCATTGCGGCTTGATCTCGATCTGTGA
35

M LOC44012

40 Cover 55% identity 63%

SEQ ID NO: 85

45 M LRKHTYFDELAQSKRAFAASAALSAPTTSGDAGGSASPPSPAAREHLFDKTVTPSDVG
KLNRLVIPQNAEKHFPLQLPAGGGESKGLLLNFEDDAGKVWRFRYSYWNSSQSYVLTKG
WSRFVKEKGLGAGDVVGFYRSAAGRTGEDSKFFI DCRLRPNTNTAAEADPVDQSSAPVQK
AVRLFVLDLLAAPEQGM PGGCKRARDLVKPPPKVAFKKQCI ELALA

SEQ ID NO: 86

50 ATGCTCCGCAAGCACACCTACTTCGACGAGCTCGCCAGAGCAAGCGCGCCTTCGCCGCG
TCGGCCGCGCTCTCCGCGCCACCACTCGGGCGACGCCGGCGGCAGCGCCTCGCCGCC
TCCCCGGCCGCGTGCGCGAGCACCTCTTCGACAAGACCGTCACGCCAGCGACGTCCGG
AAGCTGAACAGGCTGGTGATACCGAAGCAGAACGCCGAGAAGCACTTCCCGCTGCAGCTC

5 CCGGCCGGCGGGCGGCGAGAGCAAGGGCCTGCTCCTCAACTTCGAGGACGATGCGGGCAAG
GTGTGGCGGTTCCGCTACTCGTACTGGAACAGCAGCCAGAGCTACGTCCTACCAAGGGC
TGGAGCCGCTTCGTGAAGGAGAAGGGCCTCGGGCGCGGAGACGTCGTCGGGTTCTACCGC
TCCGCCGCCGGGAGGACCGGCGAAGACAGCAAGTTCTTCATTGACTGCAGGCTGCGGCCG
AACACCAACACCGCCGCCGAAGCAGACCCCGTGGACCAGTCGTCGGCGCCCGTGCAGAAG
GCCGTGAGACTCTTCGGCGTCGATCTTCTCGCGGCGCCGGAGCAGGGCATGCCGGGCGGG
TGCAAGAGGGCCAGAGACTTGGTGAAGCCGCCGCTCCGAAAGTGGCGTTCAAGAAGCAA
TGCATAGAGCTGGCGCTAGCGTAG

10

MLOC_57250

Cover 50% identity 57%

15

SEQ ID NO: 87

MYCSRGRIDPAEEGQVMGGLGVRDASWALFKVLEQSDVQVGQNRLLLTKEAVWGGPI PKL
FPELEELRGDGLNAENRVAVKILDADGCEGDANFRYLNSSKAYRVMGPQWSRLVKETGMC
KGDRLDLYAATATAASSCSGARAAVAPAPPGAIVKAAGF

20

CDS SEQ ID NO: 88

ATGTATTGTTCCCGCGGCCGCATCGATCCCGCGGAAGAAGGGCAGGTGATGGGCGGCCTC
GGCGTGCGCGACGCCAGCTGGGCGCTGTTCAAGGTGTTGGAGCAGTCCGACGTCCAGGTG
GGGCGAACC GGCTGCTCCTACCAAGGAGGCGGTGTGGGGCGGCCCTATCCCAAGCTT
TTCCCGGAGCTGGAGGAGCTCCGCGGCGACGGCCTCAACGCCGAGAACAGGGTCGCGGTC
25 AAGATCCTCGACGCCGACGGCTGCGAGGGGACGCCAACTTCCGCTACCTCAACTCCAGC
AAGGCGTACCGGGTCATGGGGCCTCAGTGGAGCCGGCTCGTGAAGGAGACCGGCATGTGC
AAGGGAGACCGCCTCGATCTGTACGCGGAACGGCGACCGCTGCCTCTTCGTGTTCTGGA
GCCAGGGCGGCTGTGGCGCCGGCGATACCTCCCGAGCAATCGTGAAGGCAGCCGGGTTCTAA

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MLOC_38822

Cover 47% identity 56%

35

SEQ ID NO: 89

MLRKH IYPDELAQHKAFFFAAASSPTSSSSPLASPAPSAARREHLFDKTVTPSDVGK
LNRLVIPKQHAEKHFPLQLPSASAAVPGECKGVLLNFDDATGKVWRFYRYSWNSSQSYVL
TKGWSRFVKEGLHAGDAVEFYRAASGNQLFI DCKLRSKSTTTTTSVNSEAAPSPAPVT
RTVRLFGVDLLIAPAARHAHEHEDYGMAKTNKRTMEASVAAPTPAHAVWKKRCVDFALTY
RLATTPQCPRSRDQLEGVQAAGSTFAL

40

CDS SEQ ID NO: 90

ATGCTGCGCAAGCACATCTATCCCGACGAGCTCGCGCAGCACAAGCGCGCCTTCTTCTTC
GCCGCGGCGTGTCCCTACGTCGTGTCGTACCTCTCGCCTCGCCGGCTCCTCAGCC
GCGGCGGCGCGGCGGAGCACCTGTTGACAAGACGGTCACGCCAGCGACGTGGGGAAG
45 CTGAACCGGCTGGTGATCCCAAGCAGCAGCCGAGAAAGCACTTCCCGCTGCAGCTCCCT
TCTGCCAGCGCCGCGTGCAGGCGAGTGCAAGGGCGTGCTGCTCAACTTCGATGACGCG
ACCGGCAAGGTGTGGAGGTTCCGGTACTCCTACTGGAACAGCAGCCAGAGCTACGTGCTC
ACCAAGGGGTGGAGCCGCTTCGTGAAGGAGAAGGGCCTTACGCCGGCGACGCCGTGAG
TTCTACCGCGCCGCTCCGGCAACAACAGCTCTTCATCGACTGCAAGCTCCGGTCCAAG
50 AGCACCACGACGACACCTCCGTCAACTCGGAGGCCGCCCATCGCCGGCACCCGTGACG
AGGACAGTGGCACTCTTCGGGGTCGACCTTCTCATCGCGCCGGCGGCGAGGCACGCGCAT
GAGCACGAGGACTACGGCATGGCCAAGACAAAGAGAACCATGGAGGCCAGCGTAGCG
GCGCCTACTCCGGCGCACGCGGTGTGGAAGAAGCGGTGCGTAGACTTCGCGCTGACCTAC

CGACTTGCCACCACCCACAGTGCCCGAGGTCAAGAGATCAACTAGAAGGAGTACAAGCA
GCTGGGAGTACATTGCTCTATAG

5 MLOC_7940
Cover 49% identity 52%

SEQ ID NO: 91

MGVEI LSSTGEHSSQYSSGAASTATTESGVGGRPPTAPSLPVSIADESATSRSSASAQSTS
10 SRFKGVVPQPNGRWGAQIYERHARVWLGTFPDEDSAARAYDVAALRYRGREAATNFPCAA
AEAELAFLAHSAEIVDM LRKHTYTDELRLRGRGMGARAQPTPSWAREPLFEKAVT
PSDVGKLNRLVVPKQHAKEH FPLKRTPETTTTTGKGVLLNFEDGEGKVWFRYSYWNSSQ
SYVLTGWSRFRVREKGLGAGDSIVFSCSAYGQEKQFFIDCKKNKMTSCPADDRGAATAS
PPVSEPTKGEQVRVRLFGVDIAGEKRGRAAPVEQELFKRQCVAHSQHSPALGAFVL

CDS SEQ ID NO: 92

ATGGGGGTGGAGATCCTGAGCTCAACGGGGGAACACTCCTCCCAGTACTCTTCCGGAGCC
20 GCGTCCACGGCGACGACGGAGTCAGGCGTGGGCGGACGGCCCGCGACTGCGCCGAGCCTA
CCTGTTTCCATCGCCGACGAGTCGGCGACCTCGCGGTGGCATCGGCGCAGTCGACGTCTG
TCGCGGTTCAAGGGCGTGGTGCCGCAGCCCAACGGGCGGTGGGGCGCCCAGATCTACGAG
CGCCACGCCCCTGCTGCTCGGCACGTTCCTCGGACGAAGACTCTGCGGCGCGCGCCTAC
GACGTGGCCGCGCTCCGGTACCGGGGCCGCGAGGCCGCCACCAACTTCCCGTGC GCGGCC
25 GCGGAGGCGGAGCTCGCCTTCTGGCGGCACACTCCAAGGCCGAGATCGTCGACATGCTC
CGGAAGCACACCTACACCGACGAGCTCCGCCAGGGCCTGCGGCGCGGCCGCGCATGGGG
GCGCGCGCGCAGCCGACGCCGTCTGGGCGCGGGAGCCCCTTTTCGAGAAGGCCGTGACC
CCGAGCGACGTGGGCAAGCTCAACCGCCTCGTTGTGCCGAAGCAGCACGCCGAGAAGCAC
TTCCCCCTGAAACGCACGCCGAGACGACAACGACCACCGGCAAGGGGGTGCTTCTCAAC
30 TTCGAGGATGGCGAGGGGAAAGTGTGGAGTTCCGGTACTCGTATTGGAACAGCAGCCAG
AGCTACGTGCTACCAAGGGATGGAGCCGCTTCGTTCCGGGAGAAGGGCCTCGGTGCCGGC
GACTCCATCGTGTCTCTCTGCTCGGCGTACGGTCAGGAGAAGCAGTTCTTCATCGACTGC
AAGAAGAACAAGACGATGACGAGCTGCCCCGCCGATGACCGCGGCGCCGCAACAGCGTCG
CCGCCAGTGTGAGAGCAACAAAAGGAGAACAAAGTCCGTGTTGTGAGGCTGTTCCGGCGTC
35 GACATCGCCGGAGAGAAGAGGGGGCGAGCGGCGCCGGTGGAGCAGGAGTTGTTCAAGAGG
CAATGCGTGGCACACAGCCAGCACTCTCCAGCCCTAGGTGCCTTCGTCTTATAG

MLOC_56567

Cover 42% identity 59%

SEQ ID NO: 93

MGVEI LSSMVEHSFYSSGASSATAESGAVGTPPRHLSLPVAIADESLSRSASSRFGV
VPQPNGRWGAQIYERHARVWLGTFPDQDSAARAYDVASLRYRGDAAFNPCVVVEAELA
FLAAHSAEIVDM LRKQTYADELRQGLRRGRGMGVRAQPMPSWARVPLFEKAVTPSDVGK
45 LNRLVVPKQHAKEHFPLKRSPETTTTTGNGVLLNFEDGQGVWFRYSYWNSSQSYVLTG
GWSRFRVREKGLGAGDSIMFSCSAYGQEKQFFIDCKKNNTTVNGGKSASPLQVMEIAKAEQV
RVVRLFGVDIAGVKRERAATAEQGPQGWFKRQCMAHGQHSPALGDFAL

SEQ ID NO: 94

ATGGGGGTGGAGATCCTGAGCTCCATGGTGGAGCACTCCTTCCAGTACTCTTCGGGCGCG
50 TCCTCGGCCACCGCGGAGTCAGGCGCGCTCGGAACACCGCCGAGGCATCTGAGCCTACCT
GTCGCCATCGCCGACGAGTCCCTGACCTCACGGTCGGCGTCGTCTCGGTTCAAGGGCGT

GTGCCGCAGCCCAACGGGCGGTGGGGCGCCCAGATCTACGAGCGCCACGCTCGCGTCTGG
CTCGGCACGTTCCAGACCAGGACTCGGCGGCGCGCCTACGACGTTGCCTCGCTCAGG
TACCGCGGCGGCGACGCCGCTTCAACTTCCCGTGCCTGGTGGAGGCGGAGCTCGCC
TTCCTGGCGGCGCACTCCAAGGCTGAGATCGTTGACATGCTCCGGAAGCAGACCTACGCC
5 GATGAACTCCGCCAGGACTACGGCGCGGCCGTGGCATGGGGGTGCGCGCGCAGCCGATG
CCGTCGTGGGCGCGGGTTCCCCCTTTTCGAGAAGGCCGTGACCCCTAGCGATGTGGCAAG
CTCAATCGCCTGGTGGTGCCGAAGCAGCACGCCGAGAAGCACTTCCCCCTGAAGCGCAGC
CCGAGACGACGACCACCACCGGCAACGGCGTACTGCTCAACTTTGAGGACGGCCAGGGA
AAAGTGTG GAGGTTCCG GTACTCATATTGAACAGCAGCCAGAGCTACGTGCTACCAAA
10 GGCTGGAGCCGCTTCGTCCGGGAGAAGGGCCTCGGCGCCGGTGAATCATCATGTTCTCC
TGCTCGGCGTACGGGCGAGGAGAAGCAGTTCTTCATCGACTGCAAGAAGAACACGACCGTG
AACGGAGGCAAATCGGCGTCGCCGCTGCAGGTGATGGAGATTGCCAAAGCAGAACAAGTC
CGCGTCGTTAGACTGTTCCGGTGTGACATCGCCGGGGTGAAGAGGGAGCGAGCGGCGACG
GCGGAGCAAGGCCCGCAGGGGTGGTTCAAGAGGCAATGCATGGCACACGGCCAGCACTCT
15 CCTGCCCTAGGTGACTTCGCCTTATAG

MLOC_75135

Cover 43% identity 57%

20 SEQ ID NO: 95
MGMEILSSTVEHCSQYSSSASTATTESGAAGRSTTALSLPVAITDESVTSRASAPASS
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EAELAFLTAHASKAEIVDMLRKHTYADELRQGLRRGRGMGARAQPTPSWARVPLFEKAVTP
SDVGKLNRLVVPKQHAKEH FPLKCTAETTTTTGNGVLLNFEDGEGKVWRFRYSYWNSSQS
25 YVLTKGWSSSFVREKGLGAGDSIVFSSSAYGQEKQLFINCKKNTTMNGGKTALPLPVVETA
KGEQDHVVKLFGVDIAGVKRVRAATGELGPPELFRQSVAHGCGRM NYICYSIGTIGPLM
LN

30 SEQ ID NO: 96
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GTTGCCATCACCGACGAGTCCGTTACCTCGCGGTGCGCATCGGCGCAGCCGGCGTCATCA
CGGTTCAAGGGCGTGGTGCCGCGAGCCCAACGGGCGGTGGGGCTCCCAGATCTACGAGCGC
CACGCTCGCGTCTGGCTCGGCACCTTCCCGGATCAGGACTCGGCGGCGCGTGCCTACGAC
35 GTTGCCCTCGCTCAGGTACCGGGGCCGCGATGCCGCCACCAACTTCCCGTGCGCCGCTGCG
GAAGCGGAGCTCGCCTTCCTGACCGCGCACTCCAAGGCCGAGATCGTCGACATGCTCCGG
AAGCACACCTACGCCGACGAACTCCGCCAGGGCCTGCGGCGCGGCCGCGGCATGGGTGCG
CGCGCGCAGCCGACGCCGTCGTGGGCGCGGGTCCCCCTTTTCGAGAAGGCTGTGACCCCT
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40 CCCCTGAAGTGACCGCAGAGACGACGACCACCACCGCAACGGCGTGCTGCTAACTTC
GAGGATGGTGAGGGGAAGGTGTGGAGGTTCCGGTACTCGTATTGGAACAGTAGCCAGAGC
TACGTGCTACCAAAGGCTGGAGCAGCTTCGTCCGGGAGAAGGGCCTCGGCGCAGGCGAC
TCCATCGTCTTCTCCTCCTCGGCGTACGGGCGAGGAGAAGCAGTTATTCATCAACTGCAA
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45 AAAGGAGAACAAGACCACGTCGTAAAGTTGTTCCGGTGTGACATCGCCGGTGTGAAGAGG
GTGCGAGCGGCGACGGGGGAGCTAGGCCCGCCGGAGTTGTTCAAGAGACAATCCGTGGCA
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50

MLOC_63261

Cover 49% identity 51%

SEQ ID NO: 97

MASSKPTNPEVDNDMECSSPESGAEDAVESSSPVAAPSSRFKGVVPQPNGRWGAQIYEKH
SRVWLGTFGDEEAAACAYDVAALRFRGRDAVTNHQRLPAAEGAGWSSTSELAFLADHKA
EIVDMLRKHTYDDELRRQGLRRGHGRAQTPAWAREFLFEKALTPSDVGKLNRLVVPKQHA
EKHFPPTTAAAGSDGKGLLLNFEDGQGKVVFRFRYSYWNSSQSYVLTKGWSRFVQEKGLC
AGDVTFTSRSAAYVMNDTDEQLFI DYKQSSKNDEAADVATADENEAGHVAVKLFQVDIGWA
GMAGSSGG

SEQ ID NO: 98

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TCGCGGGTGTGGCTTGGCAGTTTCGGGACGAGGAAGCCGCCGCTGCGCCTACGACGTG
GCCGCGCTCCGCTTCCGCGGCCGCGACGCCGTACCAACCACCAGCGCCTGCCGGCGGCG
GAGGGGGCCGGCTGGTCTCCACGAGCGAGCTCGCCTTCTCGCCGACCACTCCAAGGCC
GAGATCGTCGACATGCTCCGGAAGCACACCTACGACGACGAGCTCCGGCAGGGCCTGCGC
CGCGGCCACGGGCGCGCGCAGCCACGCCGGCGTGGGCGCGAGAGTTCCTCTTCGAGAAG
GCCCTGACCCCGAGCGACGTCCGGAAGCTCAACCGCCTGGTCGTTCCGAAGCAGCACGCC
GAGAAGCACTTCCCCCGACGACGGCGGCGGCCGCCGAAGCGACGGCAAGGGCTTGCTG
CTCAACTTCGAGGACGGCCAAGGGAAGGTGTGGAGGTTCCGGTACTCATACTGGAACAGC
AGCCAGAGCTACGTGCTACCAAGGGCTGGAGCCGCTTCGTCCAAGAAAAGGGCCTCTGC
GCCGGCGACACCGTGACGTTCTCCCGGTGCGCGTACGTGATGAATGACACGGATGAGCAG
CTCTTCATCGACTACAAGCAGAGTAGCAAGAACGACGAAGCGGCCGACGTAGCCACTGCC
GATGAGAATGAGGCCGGCCATGTCGCCGTGAAGCTCTTCGGGTCGACATTGGCTGGGCT
GGGATGGCGGGATCATCAGGTGGGTGA

MLOC_64708

Cover 49% identity 51%

SEQ ID NO: 99

MLFDSSVSASLGTMLPLVKKLDMLLAPARGYSTLCKRI KEVM HLLKHDVEEISSYLDEL
EVEDPPPMACWML NEARDLSYDMEDI DSLLFVPPGHFI KKKKKKKKKGKKKMVI KKRLK
WCKQIVFTKQVSDHGI KTSKIIHVNVPRLPNPKVAKI ILQFRIYVQEI ERYDKYRLHH
CSTLRRRLSTGSMLSVPIPYEEAAQIVTDGRMNEFISSLAANNAADQQQLKVVSVLGS
CLGKTTLANVLYDRIGMQFECRAFI RVSKKPDMLRFRDLSQFHQKQPLPTSCNELGIS
DNI IKHLQDKRYLVI DDLWDLSVWDII KYAFPKGNHGSRII ITTQI EDVALTCCCDHSE
HVFEM KPLN IGHRSRELFNRLFGSESDCLEEFKRVSNIEIVDICGGLPLATI NIASHLANQ
ETEVSLLDLDTRDRLRSCLWSNSTSERTKQVLNLSYSNLPDYLKTCCLLYLHMYPVGSI I
WKDDLKQLVAEGFIATREGKDQDQEM IEKAAGLCFDALI DRRFIQPIYTKYNNKVLST
VHEVVHDLIAKSAEENFIVVADHNRKN IALSHKVRRLSLIFGDTIYAKTPAN ITKSQIR
SFRFFGLFECM PCITEFKVLRVLNLQLSGHRGDN DPI DLTGISELFQLRYLKITSQVCIK
LPNQMQKLQYLETLDI MDAPRVTAVPWDI INLPHLLHLTPVDTYLLDWISSMTDSVISL
WTLGKLNLYQH LHLTSSSTRPSYHLERSVEALGYLIGGHGKLTIVVAHVSSAQNTVVRG
APEVTISWDRMSPPPLLQRFECPHSCFIFYRI PKWVTELGN LcI LKIAVKELHM ICLGTL
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VFNAIPRMDQNLVFFHHSRPAM HQRGGAVI IVEHMPGLRVISAKFGGAASDLEYASRTVV
SNH PSN PTI NMQLVCYSSNGKRSRKRKQPYDVVKGQPDYAKRLERPAEKRISTPTKSS
LRLHVPEITPKMQITDNNVQRREH MFDTVLTRGDVGMNLNRLVVPKKHAEKYFPLDSSST
RTSKAIVLSFEDPAGKSWFFHYSYRSSSQNYVMFKGWTGFVKEKFLEAGDTVFSRGGVGE
ATRGRFLIDCQNEQRYM FERVLTASDMESDGCSLMVPVNLVWPHPLRKT I KGRHAVLQF
EDGSGNGKVPWFQFEASGQYYLMKGLNYFVNDRLAAGYTVSFYRAGTRLFVDSGRKDDK

VALGTRSRERIYPKIVRSQ

Brassica rapa

5

LOC103849927

Cover 99% ident 80%

CDS SEQ ID NO: 100

10

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TGCAGAGCTTTTCATTGAGTGTCCAAAAAGCCTGATATGAAGAGACTTTTCCGTGACTTG
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CTTTTTGGTTCTGAAAGTGACTGTCTTGAAGAATTCAAACGAGTTTCAAACGAAATTGTT
GATATATGTGGTGTTTACCGCTAGCAACAATCAACATAGCTAGTCAATTTGGCAAACAG
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CCTGATTATCTGAAGACATGTTTGCTGTATCTTCATATGTATCCAGTGGGCTCCATAATC
TGGAAGGATGATCTGGTGAAGCAATTGGTGGCTGAAGGGTTTATTGCTACAAGAGAAGGG
AAAGACCAAGACCAAGAAATGATAGAGAAAGCTGCAGGACTCTGTTTCGATGCACTTATT
GATAGAAGATTATCCAGCCTATATATACCAAGTACAACAATAAGGTGTTGCTCTGCACG
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CCTTCATACCATCTGGAGAGAAGTGTGGAGGCTCTGGGTTATTTGATCGGAGGACATGGC
AAGCTGAAAACCTATAGTAGTCGCTCATGTCTCCTCTCTCTCAAAATACTGTGGTTCGTGGC
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50

GGTATAGCTTGGCTGAAATTTGAGGCTGATGCAATGCCTAGTCTATGGAACTGATGCTA
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5 AGTAACCATCCAAG CAATCCTACAATCAACATGCAATTGGTGTGTTATAGTTCCAATGGT
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CAGAGGAGGGGAGCACATGTTTCGATACGGTCTGACTCGGGGGGACGTGGGGATGCTGAAC
10 CGGCTGGTGGTACCGAAGAAGCACGCGGAGAACTTCCCGCTGGACAGTTCCTCCACC
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15 GTGCTGACGGCGAGTGATATGGAGTCGATGGCTGCTCGCTGATGGTCCCAGTGAACCTG
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20 GTAGCCTTGGGAACCAG AAG CCGCGAAAG GATCTATCCTAAGATCGTGCGGTGCGCAGTAG

LOC103849927

25 SEQ ID NO: 101
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30 Bra034828
Cover 100% identity 79%

35 SEQ ID NO: 102
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HRFDI HRLFIGWRRRGEASSSSAVSAVTQDPRANTTAYWNLTPYRQVHASTSSYPNNI
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40 M NISFAGEAM EQVGDGRG

CDS SEQ ID NO: 103
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ATACCAAAACAACACGCCGAGAGATACCTCCCTCTCAATAATTGCGGCGGCGGCGGCGAC
45 GTGACGGCGGAGTCGACGGAGAAAGGGGTGCTTCTCAGCTTCGAGGACGAGTCGGGAAAA
TCTTGAAATTAGATACTCATATTGGAACAGTAGTCAAAGCTACGTGTTGACCAAGGA
TGGAGCAGGTACGTCAAAGACAAGCACCTCAACGCAGGGGACGTGTTTTATTTCACCG
CACCGTTTTGATATTCATAGACTCTTCATTGGCTGGAGGAGACGCGGAGAGGCTTCTTCC
TCTTCGCCGTTTCCGCCGTGACTCAAGATCCTCGAGCTAACACGACGGCGTACTGGAAC
50 GGTTTGACTACACCTTATCGTCAAGTACACGCGTCAACTAGTTCTTACCCTAACAAACATC
CACCAAGAGTATTACATTATGGCCCTGTTGCTGAGACACCGACGGTAGCTGCAGGGAGC
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CCACCGTGTCTGACGCTACAACGGCCAACACATTTACTATTACTCAACTCCACATCCC

ATGAATATCTCATTTGCTGGAGAAGCAATGGAGCAGGTAGGAGATGGACGAGGTTGA

5 Bra005886
Cover 100% identity 79%

SEQ ID NO: 104

10 MSVN HYSTDH HQVHH HHTLFLQNLHTTDTSEPTTTAATSLREDQKEYLFEKSLTPSDVGK
LNRLVIPKQHAKEYFPLNTI ISNNAEEKGMLLSFEDESGKCWRFRYSYWNSSQSYVLTKG
WSRYVKDKQLDPADVFFQRQRSDSRRLFIGWRRRGQGSSSAANTTSYSSSMTAPPYSNY
SNRPAHSEYSHYGA AVATATETHFPSSSAVGSSRTVRLFGVNLECMDEDEGDDSVATA
AAECPRQDSYYDQN MYNYTPHSSAS

15 CDS 105

ATGTCAGTCAACCATTACTCCACGGACCACCACCAGGTCCACCACCACCACACTCTCTTC
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CGCGAAGACCAGAAAGAGTATCTCTTCGAGAAATCTCTCACACCAAGCGACGTTGGCAAA
20 CTCAACCGTCTCGTTATACCAAAACAGCACGCGGAGAAGTACTTCCCTCTCAACACCATC
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25 TCCGCCGCGAATACGACGTCGTATTCTAGTTCCATGACTGCTCCACCGTATAGTAATTAC
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GAGACGCACTTCATACCATCGTCTTCCGCCGTCGGGAGCTCGAGGACGGTGAGGCTTTTT
GGTGTGAATTTGGAGTGTCAAATGGATGAAGACGAAGGAGATGATTCGGTTGCCACGGCA
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30 ACTCCTCACTCCTCAGCCTCATAA

Bra005301
Cover 100% identity 58%

35 SEQ ID NO: 106

MSI NQYSSDFNYHSLMWQQQHRHHHHQNDVAEEKEALFEKPLTPSDVGKLNRLVIPKQH
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AGDVI LFHRHRVDGGRFFIGWRRRGNSSSSSDSYRHLQSNASLQYYPHAGVQAVESQRGN
40 SKTLRLFGVNMECQLDSDLPDPSTPDGSTICTPSHDQFH LYPQQHYPPPYMDISFTGDV
HQTRSPQG

CDS SEQ ID NO: 107

45 ATGTCAATAAACCAATACTCAAGCGATTTCAACTACCACTCTCTCATGTGGCAACAACAG
CAGCACCGCCACCACCACCATCAAAACGACGTCGCGGAGGAAAAAGAGCTCTTTTCGAG
AAACCCTTAACCCCAAGTGACGTCGGAAGAACTCAACCGCCTCGTCATCCCAAAACAGCAC
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50 AGCCAGAGTTATGTCTTGACCAAAGGATGGAGCAGATACGTCAAGGAGAAGCAGCTCGAC
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GATCCATCTACACCAGACGGTTCACCATATGTCCGACCAGTCACGACCAGTTTCATCTC
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CACCAGACGAGAAGCCCACAAGGATAA

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Bra017262

Cover 92% identity 56%

10

SEQ ID NO: 108

MSI NQYSSEFYHSLMWQQQQQHQQNEVVVEEKEALFEKPLTPSDVGKLNRLVIPQHA
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DVVLFHRH RADGGRFFIGWRRRGDSSSSSDSYRNLQSNSSLQYYPHAGAAQAVENQRGNSK
TLRLFGVNM ECQIDSDWSEPSTPDGFTTCPTNHDQFPIYPEH FPPPYMDVSFTGDVHQT
SSQQG

15

CDS SEQ ID NO: 109

20

ATGTCAATAAATCAATATTCAAGCGAGTTCTACTACCATTCTCTCATGTGGCAACAACAG
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AGATACTTCCCTCTCGCCGCCGCCGCGGTAGACGCCGTGGAGAAGGGATTACTCCTCTGC
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AGTTACGTCTTGACCAAGGATGGAGCAGATATGTTAAAGAGAAGCAACTTGACGCCGGC
GACGTTGTTCTCTTTTCATCGCCACCGTGCTGACGGTGGAAGATTCTTCATTGGCTGGAGA
AGACGCGGCGACTCTTCTCCTCCTCCGACTCTTATCGCAATCTTCAATCTAATTCCTCG
CTCCAATATTATCCTCATCAGGGGCTCAAGCGGTG GAGAACCAG AGAGGTAAGTCCAAG
ACATTGAGACTTTTTGGAGTGAACATGGAGTGCCAGATAGACTCAGACTGGTCCGAGCCA
TCCACACCTGACGGTTTTACCACATGTCCAACCAATCACGACCAGTTTCCTATCTACCT
GAACACTTTCCTCCTCCGTACTACATGGACGTAAGTTTCACAGGAGATGTGCACCAGACG
AGTAGCCAACAAGGATAG

25

30

Bra000434

35

Cover 96% identity 47%

SEQ ID NO: 110

40

MMTNLSLAREGEEEEEEAGAKKPTEEVEREHM FDKVVTSPDVGKLNRLVI PKQHAERYFP
LDSSTNEKGLI LNFEDLTGKSWRFRYSYWNSSQSYVMTKGWSRFVKDKKLDAGDIVSFLR
CVGDTGRDSRLFI DWRRRPKVPDYTTSTSHFPAGAMFPRFYSFQTATTSTSYNPYNHQQP
RHHHSGYCYPI PREFGYGYVRSVDQRAVVADPLVI ESVPVM MHGGARVNQAAVGTAGK
RLRLFGVDMCEGSGGTNSTEESSSSGSLPRGGASPSSMFQLRLGNSEDDH LFKKG
KSSLPFN LDQ

45

SEQ ID NO: 111

50

ATGATGACAAATTTGTCTCTTGCAAGAGAAGGAGAAGAAGAAGAAGAGGCAGGAGCA
AAGAAGCCCACAGAAGAAGTGAGAGAGAGCACATGTTGACAAAGTGGTGACTCCAAGT
GACGTGCGGAACTAAACCGACTCGTGATCCCAAAGCAACACGCGGAGAGATACTTCCT
TTAGATTATCCACAAACGAGAAGGGTTTGATTCTAAACTTCGAAGATCTCACGGGAAAG
TCATGGAGGTTCCGTTACTCTTACTGGAACAGCAGTCAGAGCTATGTCATGACTAAAGT
TGGAGCCGTTTCGTTAAAGACAAGAAGCTAGACGCTGGAGATATTGTCTCTTTCCTGAGA
TGTGTGCGGAGACACAGGAAGGGACAGCCGCTTGTTTATCGATTGGAGGAGACGACCTAAA

5 GTCCCTGACTACACGACATCGACTTCTCACTTTCCTG CCG GAGCTATGTTCCCTAGGTTT
TACAGTTTTTCAGACAGCAACTACTTCCACAAGTTACAATCCCTATAATCATCAGCAGCCA
CGTCATCATCACAGTGGTTACTGTTATCCTCAAATCCCGAGAGAATTTGGATATGGGTAT
GTCGTTAGGTCAGTAGATCAGAGGGCGGTGGTGGCTGATCCGTTAGTGATCGAATCTGTG
10 CCGGTGATGATGCACGGAGGAGCTCGAGTGAACCAGGCGGCTGTTGGAACGGCCGGGAAA
AGGCTGAGGCTTTTTGGAGTCGATATGGAATGTGGCGAGAGTGGAGGAACAAACAGTACG
GAGGAAGAATCTTCATCTTCCGGTGGGAGTTTGCCACGTGGCGGTGCTTCTCCGTCTCC
TCTATGTTTCAGCTGAGGCTTGGAACAGCAGTGAAGATGATCACTTATTTAAGAAAGGA
AAGTCTTCATTGCCTTTTAATTTGGATCAATAA

Bra040478

Cover 96% identity 48%

SEQ ID NO: 112

15 MMTNLSLAREGEAQVKKPI EEVEREHMFDKVVTPSDVGKLNRLVIPKQHAERYFPLDSSS
NEKLLLNFDLTGKSWRFRYSYWNSSQSYVMTKGWSRFVKDKLDAGDIVSFQRCVGDS
RLFI DWRRRPKVPDPTSTAHFAAGAMFPRFYSFPTATTSTCYDLNHQPPRHHHIGYGY
PQI PREFGYGYFVRSDQRAVVADPLVI ESVPVMMRGGARVSQEVVGTAGKRLRFLGVDM
20 EEESSSSGSLPRAGGGGASSSSSLFQLRLGSSCEDDH FSKKGKSSLPFDLDQ

SEQ ID NO: 113

25 ATGATGACCAACTTGTCTCTTGCAAGGGAAGGAGAAGCACAAAGTAAAGAAGCCCATAGAA
GAAGTTGAGAGAGAGCACATGTTTCGACAAAGTGGTGACTCCAAGCGACGTAGGGAAACTA
AACAGACTCGTGATCCCAAAGCAACACGCAGAGAGATACTTCCCTCTAGATTCATCCTCA
AACGAGAAAAGGTTTGCTTCTAACTTTGAAGATCTAACAGGAAAGTCATGGAGGTTCCGT
TACTCTTACTGGAACAGTAGCCAGAGCTATGTCATGACTAAAGGTTGGAGTCGTTTCGTT
AAAGACAAGAAGCTTGACGCCGAGATATTGTCTCTTCCAGAGATGTGTCGGAGACAGC
30 CGCTTGTTTATCGATTGGAGGAGACGACCTAAAGTCCCTGACTATCCGACATCGACTGCT
CACTTTGCTG CAGGAG CTATGTTCCCTAGGTTTTACAGTTTTCCG ACAGCAACTACTTCG
ACATGTTACGATCTGTACAATCATCAGCCGCCACGTCATCATCACATTGTTACGTTTAT
CCACAGATTCCGAGAGAATTTGGATACGGGTATTTCTAGGTGAGTGGACCAGAGAGCG
GTGGTGGCTGATCCGTTGGTGATCGAATCTGTGCCGGTGATGATGCGCGGAGGAGCTCGA
GTTAGTCAG GAG GTTGTG GAACGGCCGGAAGAGG CTGAGG CTTTTTGAGTCG ATATG
35 GAGGAAGAATCTTCATCTTCCGGTGGGAGTTTGCCGCGTGCCGAGGTGGCGGTGCTTCT
TCATCTTCTCTTTGTTTCAGCTGAGACTTGGGAGCAGCTGTGAAGATGATCACTTCTCT
AAGAAAGGAAAGTCTTCATTGCCTTTTGATTTGGATCAATAA

Bra004501

Cover 74% identity 45%

SEQ ID NO: 114

45 M M MTNLSLSREGESEEEEEQEEAKKPM EEVEREHM FDKVVTPSDVGKLNRLVI PKQYAE
YFPLDSSTNEKLLLNFDLAGKSWRFRYSYWNSSQSYVMTKGWSRFVKDKLDAGDIVS
FQRCVGDSGRDSRLFI DWRRRPKVPDHPTSHFAAGSM FPRFYSFPTATSYNLYNQPP
RHHHHSYNYNYPQI PREFGYGYLVDQRAVVADPLVIESVPVMM HGGAQVSQAVVGTAGKRL
RFLGVDMEEESSSSGSLPRGDASPSSSLFQLRLGSSSEDDH FSKKGKSSLPFDLDQ

SEQ ID NO: 133

50 ATGATGATGACAACTTGTCTCTTCAAGAGAAGGAGAAGAGGAGGAAGAAGAAGAACAA
GAAGAGGCCAAGAAGCCCATGGAAGAAGTAGAGAGAGAGCACATGTTTCGACAAAGTGGTG
ACTCCAAGCGATGTTGGTAAACTAAACCGGCTCGTGATCCCAAAGCAATACGCAGAGAGA

5 TACTTCCCTTTAGATTCATCCACAAACGAGAAAGGTTTGCTTCTAAACTTCGAAGATCTC
GCAGGAAAGTCATGGAGGTTCCGTTACTCTTACTGGAACAGTAGTCAGAGCTATGTCATG
ACTAAAGGTTGGAGCCGTTTCGTTAAAGACAAAAAGCTAGACGCCGAGATATTGTCTCT
TTCCAGAGATGTGTGCGGAGATTGAGGAAGAGACAGCCGCTTGTATTGATTGGAGGAGA
10 AGACCTAAAGTTCCTGACCATCCGACATCGATTGCTCACTTTGCTGCCGGATCTATGTTT
CCTAGGTTTTACAGTTTTCCGACAGCAACTAGTTACAATCTTTACAACTATCAGCAGCCA
CGTCATCATCATCAGTGGTTATAATTATCCTCAAATTCCGAGAGAATTTGGATACGGG
TACTTGGTGGATCAAAGAGCCGTGGTGGCTGATCCGTTGGTGATTGAATCTGTGCCGGTG
ATGATGCACGGAGGAGCTCAAGTTAGTCAGGCGGTTGTTGGAACGGCCGGGAAGAGGCTG
15 AGGCTTTTTGGAGTCGATATGGAGGAAGAATCTTCATCTTCCGGTGGGAGTTTGCCACGT
GGTGACGCTTCTCCGCTCTCCTCTTTGTTTCAGCTGAGACTTGAAGCAGCAGTGAAGAT
GATCACTTCTCTAAGAAAGGAAAGTCCTCATTGCCTTTTGATTGGATCAATAA

15 Bra003482
Cover 79% identity 44%

20 SEQ ID NO: 115
MNQEEENPVEKASSM EREHM FEKVVTSPDVGKLNRLVIPKQHAERYFPLDNNSDSSKGLL
LNFEDRTGNSWRFYRYSYWNSSQSYVMTKGWSRFVKDKKLDAGDIVSFQRDPGNKDKLFI D
WRRRPKI PDHHHQFAGAMFPRFYSFHPQNLHYRYQQDLGIGYYVSSMERNDPTAVI ESV
PLI MQRRAAHVAI PSSRGEKRLRLFGVDM ECGGGGGSVNSTEESSSSGGGGGVSMASV
GSLQLRLVSSDDESLVAMEAASVDEDHHLFTKKGKSSLSFDLDRK

25 SEQ ID NO: 116
ATGAATCAAGAAGAAGAGAATCCTGTGGAAAAAGCCTCTTCAATGGAGAGAGAGCACATG
TTTGAAAAAGTAGTAACACCAAGCGACGTAGGCAAATAACCGACTCGTGATCCCAAAG
CAACACGCGGAGAGATACTTCCCTTTAGACAACAATTCTGACAGCAGCAAAGGTTTGCTT
CTAAACTTCGAAGACCGAACAGGAACTCATGGAGATTCCGTTACTCTTACTGGAACAGT
30 AGCCAGAGTTATGTCATGACAAAAGGTTGGAGCCGCTTCGTCAAAGACAAGAAGCTTGAT
GCTGGCGACATCGTTTCTTTTCAGAGAGATCCTGGTAATAAAGACAAGCTTTTCATTGAT
TGGAGGAGACGACCAAAGATTCCAGATCATCATCAATTCGCTGGAGCTATGTTCCCT
AGGTTTTACTCTTCTCTCATCCTCAGAACCTTTATCATCGATATCAACAAGATCTTGGA
ATTGGGTATTATGTGAGTTCAATGGAGAGAAATGATCCAACGGCTGTAATTGAATCTGTG
35 CCGTTGATAATGCAAAGGAGAGCAGCACACGTGGCTGCTATACCTTCATCAAGAGGAGAG
AAGAGGTTAAGGCTGTTTGGAGTGGACATGGAGTGCGGCGGCGGCGGAGGAAGTGTGAAT
AGCACGGAGGAAGAGTCGTCGCTTCCGGTGGTGGCGGCGGCGGCTTTCTATGGCTAGTGTT
GGTTCTCTTCCAATTGAGGCTAGTGAGCAGTGATGATGAGTCTTTGGTAGCAATGGAA
GCTGCAAGTGTGATGAGGATCATCACTTGTTTACAAAGAAAG GAAAGTCTTCTTTGTCT
40 TTCGATTTGGATAGAAAATGA

45 Bra007646
Cover 74% identity 45%

50 SEQ ID NO: 117
MNQEN KKPLEEASTSMEREN MFDKVVTPSDVGKLNRLVIPKQHAERYFPLDNSSTNNKGL
LLDFEDRTGSSWRFYRYSYWNSSQSYVMTKGWSRFVKDKKLDAGDIVSFQRDPNCKDKLYI
DWRRRPKI PDHHHQFAGAMFPRFYSFHPQM PTSFESSHNLHYHHRFQRDLGIGYYPTAVIE
SVPVI MQRREAQVAN MASSRGEKRLRLFGVDVECGGGGGSVNSTEESSSSGGGSMRGG
VSMAGVGSLLQLRLVSSDDESLVAMEGATVDEDH LFTTKKGKSSLSFDLDI

CDS SEQ ID NO: 118

5 ATGAATCAAGAAAACAAGAAGCCTTTGGAAGAAGCTTCGACTTCAATGGAGAGAGAGAAC
ATGTTTCGACAAAGTAGTAACACCAAGCGACGTAGGGAACTAAACCGACTCGTGATCCCA
AAGCAACACGCAGAGAGATACTTCCCTTTAGACAACCTCTCAACAAACAACAAAGGGTTG
CTTCTAGACTTTCGAAGACCGTACAGGAAGCTCATGGAGATTCCGTTACTCTTACTGGAAC
AGTAGCCAAAGTTATGTATGACAAAAGGTTGGAGCCGTTTTGTCAAAGACAAGAAGCTT
10 GATGCTGGTGACATCGTGCTTTTCAAAGAGATCCCTGTAATAAAGACAAGCTTTACATA
GATTGGAGGAGACGACCAAAGATTCCAGATCATCATCAGTTCGCCGGAGCTATGTTCCCT
AGGTTTTACTCTTTCCCTCACCCTCAGATGCCGACAAGTTTTGAAAGTAGTCACAACCTT
TATCATCATCGGTTTCAACGAGATCTTGAATTGGGTATTATCCAACGGCTGTGATTGAA
TCTGTGCCGGTGATAATGCAAAGGAGAGAAGCACAAAGTGGCTAATATGGCTTCATCAAGA
GGAGAGAAGAGGTTAAGGCTGTTTGGAGTGGACGTGGAGTGC GGCGGCGGAGGAGGAGGA
15 AGTGTGAATAGCACGGAGGAAGAGTCGTCGTCTTCCGGTGGTAGTATGTCACGTGGCGGC
GTTTCTATGGCTGGTGTGGTTCTCTCCTTCAGTTGAGGTTAGTGAGCAGTGATGATGAG
TCTTTAGTAGCGATGGAAGGTGCTACTGTGATGAGGATCATCACTTGTTTACAACCTAAG
AAAGGAAAGTCTTCTTTGTCTTTGATTGGATATATGA

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Bra014415

Cover 48% identity 60%

SEQ ID NO: 119

25

MERKSNLDERSENI DSQNKMMNLEERPVQEASSMEREHM FDKVVTSPDVGKLNRLVI PK
QHAERYFPLDNNSSDN KGLLLNFEDRIGILWSFRYSYWNSSQSVMKGSRFVKDKKL
DAGDIVSFHRGSCNKDKLFI DWKRRPKI PDHQVVGAM FPRFYSYPYPIQASYERHNLHY
RYQRDIGIGYYVRSM ERYDPTAVIESVPVI MQRRAHVATMASSRGEKRLRLFGVDM ECVR
GGRGGGGSVNSTEEESSTSGGSISRGGVSMAGVGSPLQLRLVSSDQDDQSLVARGAARVD
EDHH LFTKKGKSSLSFDLKD

30

CDS SEQ ID NO: 120

35

ATGGAGAGGAAGTCCAATGATCTTGAGAGATCTGAGAATATTGATTCTCAAACAAGAAG
ATGAATCTAGAAGAAGAGAGGCCTGTACAAGAAGCTTCTTCGATGGAGAGAGAGCACATG
TTCGACAAAGTAGTAACACCAAGCGACGTTGGGAACTAAACCGGCTGGTGATCCCAAAG
CAACACGCAGAGCGATACTTCCCTTTAGACAATAATTCCTCAGACAACAACAAAGGTTTG
CTTCTAAACTTCAAGATCGAATAGGAATCTTATGGAGTTTCCGTTACTCCTACTGGAAC
AGTAGCCAAAGTTATGTAATGACTAAAGGCTGGAGCCGTTTCGTCAAAGACAAGAAGCTT
GATGCTGGCGACATAGTTTCTTTTCATAGAGGTTCTTGTAAATAAAGACAAGCTTTTCATT
GATTGGAAGAGACGACCAAAGATTCTGATCACCAAGTCGTCGGAGCTATGTTCCCTAGG
40 TTTTACTCTTACCCTTATCCTCAGATACAGGCTAGTTATGAACGTCAACCTTTATCAT
CGATATCAACGAGATATAGGAATTGGGTATTATGTGAGGTCAATGGAGAGATATGATCCA
ACGGCTGTAATTGAATCTGTGCCGGTGATAATGCAAAGGAGAGCACATGTGGCTACTATG
GCTTCATCAAGAGGAGAGAAGAGGTTAAGGCTTTTTGGAGTGGATATGGAGTGCCTCAGA
GGCGGCCGAGGAGGAGGAGGAAGTGTGAATAGCACGGAGGAAGAGTCTTCGACTTCCGGT
45 GGTAGTATCTCACGTGGCGGCGTTTCTATGGCTGGTGTGGCTCTCCACTCCAGTTGAGG
TTAGTGAGCAGTGACGGTGATGATCAGTCTCTAGTAGCTAGGGGAGCTGCTAGGGTTGAT
GAGGATCATCACTTGTTTACAAAGAAAGGAAAGTCTTCTTTGTCTTTGATTTGGATAAA
TGA

50

Bra038346

Cover 51% identity 57%

SEQ ID NO: 121

MVFSCI DESSSTSESFSPATATATATATKFSAPPLPPLRLNMRSGGSNVVLDSKNGVDI

5 DSRKLSSSKYKGVVPQPNGRWGAQIYVKHQRVWLGTFCDEEEAAHSYDIAARKFRGRDAV
VNFKTFLEASEDDNGELCFLEAHSKAEIVDM LRKHTYADELAQSNKRSGANTNTNTTQSH
VSRTRVLFKVVTPSDVGKLNRLVI PKQHAKEYFPLPSLSVTGKGLI NFEDVTGKVWRF
RYSYWNSSQSYVLTGWSRFVKEKNLRAGDVVTFERSTGSDRQLYIDWKI RSGPSKNPVQ
VVVRLFGVDI FNVTSAKPSNVVDACGGKRSRDVDM FALRCSKKHAIINAL

CDS SEQ ID NO: 122

10 ATGGTATTTCAGTTGCATAGACGAGAGCTCTTCCACTTCAGAATCTTTTTACCCGCAACC
GCAACCGCAACCGCAACCGCCACAAAGTTCTCTGCTCCTCCGCTTCCACCGTTACGCCTC
AACCGGATGAGAAGCGGTGGAAGCAACGTCGTGTTGGATTCAAAGAATGGCGTAGATATT
GATTCACGGAAGCTATCGTCGTCAAAGTACAAAGGCGTGGTTCCTCAGCCCAACGGAAGA
TGGGGAGCTCAGATTTACGTGAAGCACCAGCGAGTTTGGCTGGGCACCTTCTGCGATGAA
15 GAGGAAGCTGCTCACTCCTACGACATAGCCGCCCGTAAATTCCGTGGCCGTGACGCCGTT
GTCAACTTCAAACCTTCTCGCCTCAGAGGACGACAACGGCGAGTTATGTTTCCTTGAA
GCTCACTCCAAGGCCGAGATCGTCGACATGTTGAGGAAACACACTTACGCTGACGAGCTT
GCGCAGAGCAATAAACGCAGCGGAGCGAATACGAATACGAATACGACTCAAAGCCACACC
GTTTCGAGAACACGTGAAGTGCTTTTCGAGAAGTTGTACGCCTAGCGACGTTGGTAAG
CTAAACCGCCTCGTGATACCTAAACAGCACGCGGAGAAATATTTCCGTTACCGTCACTG
20 TCGGTGACTAAAGGCGTTCTGATCAACTTCGAAGACGTGACGGGTAAGGTGTGGCGGTT
CGTTACTCATACTGGAACAGTAGTCAAAGTTACGTGTTGACCAAGGGATGGAGTCGGTTC
GTTAAGGAGAAGAATCTCCGAGCCGGTGATGTCGTTACTTTTCGAGAGATCGACCGGTTCA
GACCGGCAGCTTTATATTGATTGGAAAATCCGGTCTGGTCCGAGCAAAAACCTGTTTACG
GTTGTGTTAGGCTTTTCGAGTTGACATCTTCAACGTGACAAGCGCGAAGCCGAGCAAC
25 GTTGTAGACGCGTGCGGTGGAAAGAGATCTCGGGATGTTGATATGTTTGCCTACGGTGT
TCCAAAAACACGCTATAATCAATGCTTTGTGA

Zea mays

GRMZM2G053008

Cover 74% identity 47%

30
SEQ ID NO: 123
MAAPSSPLTAPPEPVTPPSPWTITDGAISGTLPAAEFAVHYPGYPSSPARAARTLGGL
PGLAKVRSSDPGARLELRFRPEDPYCHPAFGQSRSTGLLLRLSKRKGAAPCAHVVARV
RTAYYFEGMADFQHVVPVHAAQTRKRKHSQNDNENFGSDKTGHDEADGDVM MLVPPLF
35 SVKDRPTKIALVPSSNAISKTMH RGVVQERWEMNVGPTLALPFNTQVVPEKINWEDHI RK
NSVEWGWQMAVCKLFDERPVWPRQSLYERFLDDNVHVSQNQFKRLLFRAGYYFSTGPFK
FWI RRGYDPRKDSQSIYQRI DFRM PPELRYLLRLKNSESARKWADMCKLETMPQSFIYL
QLYELKDDFIQAEI RKPSYQSVCSRSTGWFSKPM IKTLRLQVSI RLLSLHNEEAKNLLR
NAHELI ERSKKQEALSRSELSI EYNDADQVSAHTGTEDQVGPNNSDSEDVDDEEEEL
40 EGYDSPPMADDI HEFTLGDSYAFGEGFSNGYLEEVLRSLPLQEDGQKKLCDAPINADASD

CDS SEQ ID NO: 124

45 ATGGCCGCCTCGCCCTCTTCACCCTTGACAGCGCCGCCAGAGCCGGTGACCCCGCCGTCC
CCATGGACCATCACAGACGGAGCCATCTCTGGCACGCTCCCAGCAGCCGAGGCCTTCGCA
GTGCACTACCCGGGCTACCCCTCCTCTCCCGCCCGCGCCCGCCGACCCCTCGGCGGTCTC
CCCGGCCTCGCCAAAGTCCGGAGTTCCGATCCCGGCGCCCGCCTCGAGCTCCGCTTCCG
CCCGAGGACCCCTACTGCCATCCAGCCTTTGGCCAGTCCCGCGCCTCCACTGGCCTTCTG
CTGCGCCTCTCCAAGCGCAAAGGAGCTGCGGCACCTTGTGCCATGTGGTCTGCTCGTGT
50 CGGACTGCTTACTACTTCAAGGTATGGCAGATTTTCAACATGTTGTTCCAGTGATGCT
GCACAAACAAGAAAAAGAAAACACTCAGATTCTAAAAATGATAATGAGAATTTGGTAGT
GATAAGACAGGACATGATGAAGCAGATGGAGATGTCATGATGTTGGTACCCCTCTCTTT
TCAGTGAAG GATAGGCCAACAAAG ATAGCGCTTGATCATCGTCCAATGCCATATCTAAA

ACCATGCACAGGGGAGTTGTACAAGAACGGTGGGAGATGAATGTTGGACCAACTCTGGCG
CTTCCGTTCAACACTCAAGTTGTCCCGGAGAAGATTAATTGGGAAGACCACATTAGAAAAG
AATTCTGTAGAATGGGGTTGGCAAATGGCTGTTTGCAAATTGTTTGATGAGCGCCCTGTG
TGGCCAAGGCAATCACTTTATGAGCGGTTCTTGATGATAATGTGCATGTCTCTCAAAAC
5 CAATTCAAAAGGCTTCTGTTTAGAGCTGGATACTACTTCTCTACTGGACCCTTTGAAAA
TTTTGGATCAGAAGAGGATATGACCCTCGTAAAGACTCTGAGTCACAAATATATCAGAGA
ATTGATTTTCGCATGCCCTCCCGAGCTACGATATCTTCTAAGGCTGAAGAATTCTGAGTCT
CGAAAGTGGGCAGATATGTGCAAGCTTGAACAATGCCATCACAGAGTTTCATCTACCTG
CAATTATATGAACTGAAGGATGATTTTATTCAAGCAGAAATTCGAAAACCTTCTTATCAA
10 TCAGTTTGTTACGTTCTACAGGATGGTTTTCTAAGCCAATGATCAAAACCCTGAGGTTG
CAAGTGAGCATAAGGCTCCTCTCTTTATTGCATAATGAAGAGGCTAAAACTTGTTGAGG
AATGCCCATGAGCTTATTGAAAGGTCCAAGAAGCAGGAAGCCCTTCGAGATCTGAGCTG
TCAATAGAATATAATGATGCTGATCAAGTTTCTGCCGCACATACTGGAAGTGAAGGATCAA
GTCGGCCCTAACAACTCTGATAGTGAAGATGTGGATGATGAAGAAGAGGAAGAGGAATTG
15 GAGGGTTATGATTCTCCACCTATGGCAGATGATATTCATGAGTTCACCTTAGGTGATTCC
TATGCATTTGGTGAAGGCTTCTGAATGGATACCTCGAAGAAGTACTGCGCAGCTTGCCA
TTGCAGGAAGACGGCCAAAAGAAATTATGTGATGCTCCTATCAACGCTGATGCAAGTGAT
GGAGAGTTTGAAATTTACGAACAGCCCAGTGATGATGAAGATTCTGATGGCTAG

20 GRMZM2G102059_T01
Cover 47% identity 62%
SEQ ID NO: 125
M EFASSSSRFSREDEEEEEQEEEEEEEEASPREI PFMTAAATADTGAAASSSSPSAAASS
25 GPAAAPRSSDGAGASGSGGGGSDDVQVI EKEHM FDKVVTSPDVGKLNRLVIPQHAKEYF
PLDAAAN EKGQLLSFEDRAGKLWFRYSYWNSSQSYVMTKGWSRFVKEKRLDAGDTSFC
RGAGDTARDRLFI DWKRRADSRDPH RM PRLPLMAPVASPYGPWGGGGGGGAGGFFM PPA
PPATLYEHHRFRQALDFRNI NAAAAPARQLLFFGSAGM PPRASMPQQQPPPPPHPLHS
IM LVQPSAPPTASVPMLLDSVPLVNSPTAASKRVRLFVNLNPNQPGTSAESSQDANAL
30 SLRTPGWQRPGPLRFFESPQRGAESSAASSPSSSSSSSKREAHSSLDL

CDS SEQ ID NO: 126
ATGGAGTTCGCGAGCTCTTCGAGTAGGTTTTCCAGGGAGGAGGACGAGGAGGAAGAGCAG
35 GAGGAAGAGGAGGAGGAGGAGGAGGCGTCTCCGCGCGAGATCCCCTTCATGACAGCGGCA
GCGACGGCCGACACCGGAGCCGCGCCTCCTCGTCTCGCCTTCCGCGCGGCGCCTCATCG
GGTCTGTGCTGCCCCCGCTCGAGCGACGGCGCCGGGCGTCCGGGAGCGGCGGCGGCG
GGGAGCGACGACGTGCAGGTGATCGAGAAGGAGCACATGTTTCGACAAGGTGGTGACGCC
AGCGACGTGGGGAAGCTCAACCGGCTGGTGATCCCGAAGCAGCACGCGGAGAAGTACTTC
40 CCGCTGGACGCGGCGGCCAACGAGAAGGGCCAGCTGCTCAGCTTCGAGGACCGCGCCGGT
AAGCTCTGGCGCTTCCGCTACTCTACTGGAACAGCAGCCAGAGCTACGTCATGACCAAG
GGCTGGAGCCGCTTCGTCAAGGAGAAGCGCCTCGACGCCGGCGACACCGTCTCCTTCTGC
CGCGGCGCGGCGACACCGCGCGGGACCGCCTTTTCATCGACTGGAAGCGCCGCGCCGAC
TCCCGCGACCCGACCGCATGCCGCGCCTCCGCTCCCCATGGCGCCCGTGCCTCGCCC
45 TACGGCCCCGTGGGGCGGCGGCGGCGGCGGCGGCGGCGGCGGCGGTTTCTTCATGCCGCCGCG
CCGCCCGCCAACTCTACGAGCACCAACCGCTTCCGCCAGGCCCTCGACTTCGCAACATC
AACGCCGCGGCCGCGCCGGCCAGGCAGCTCCTCTTCTCGGCTCAGCCGGCATGCCCCCG
CGCGCGTCCATGCCGCGAGCAGCAGCAGCCGCTCCGCCCCCGCACCCGCTCTGCACAGC
ATTATGTTGGTGCAACCCAGCCCCGCGCCGCCACGGCCAGCGTGCCCATGCTTCTCGAC
50 TCGGTACCGCTCGTCAACAGCCCAACGGCAGCGTCGAAGCGCGTCCGCTGTTGGGGTC
AACCTCGACAACCCGCAACCAGGCACAAGTGCGGAGTCAAGCCAAGATGCCAACGCATTG
TCGCTGAGGACACCGGGATGGCAAAGGCCGGGGCGGTTGAGGTTCTTGAATCGCCTCAA
CGCGGCGCCGAGTCATCTGCAGCCTCCTCGCGTCGTCATCGTCTCTCAAGAGAGAA

5 SEQ ID NO: 127

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AACCTCGACTGCGCCGCCGCCGCCGGCTCAGAGGAGGAGAACGTCGGCGGGTGGAGGACT
AGTGCGCCGCCGACGCAGCAGGCGTCTCCTCCTCATCCTACTCTCCGGGAAAGCGAGG
TGCTCCTTGAACCTTGACTTGTGA

5 GRMZM2G024948_T01
Cover 46% identity 63%
SEQ ID NO: 131

M DQFAASGRFSREEEADDEEQEDASNSMREISFMPAAAASSSSAAASASASASTSASACAS
GSSSAPFRSASASGDAAGASGSGPADADAEAEAVEKEHMFDPKVVTPSDVGKLNRLVI PK
10 QYAEKYFPLDAAANEKGLLLSFEDSAGKHWRFRYSYWNSSQSYVMTKGWSRFVKEKRLVA
GDTVSFSRAAAEDARHRLFI DWKRRVDTRGPLRFSGLLALPM PLPSSHYGGPHHYSWPWGFG
GGGGGGGGFFMPPSPATLYEHRLRQGLDFRSMTTTTYPAPTVGRQLLFFGSARM PPHHAP
PPQPRPFSPLH HYTVQPSAAGVTAASRPVLLDSVPVI ESPTTAAKRVRLFVGNLDNN PD
GGGEASHQGDALSLQMPGWQRTPTLRLLLEPRHGGESSAASSPSSSSSSSKREARSALDL
15 DL

CDS SEQ ID NO: 132

ATGGACCAGTTCCGCCGAGCGGGAGGTTCTCTAGAGAGGAGGAGCGGACGAGGAGCAG
20 GAGGATGCGTCCAATTCCATGCGCGAGATCTCCTTCATGCCGCCGGCTGCGGCCTCGTCA
TCTTCGGCGGCTGCTTCCGCGTCCGCGTCCGCCTCCACCAGCGCATCCGCGTGTGCATCG
GGAAGCAGCAGCGCCCCCTCCGCTCCGCTCCGCGTCCGGGGGATGCCGCCGGAGCGTCCG
GGGAGCGGCGGCCAGCGGACGCGGACGCGGAGGCGGAGGCGGTGGAGAAGGAGCACATG
TTCGACAAGGTGGTCACGCCGAGCGACGTGGGGAAGCTCAACCGGCTGGTGATCCCGAAG
25 CAGTACGCGGAGAAGTACTTCCCGCTGGACGCGGCGGCCAACGAGAAGGGCCTCCTCCTC
AGCTTCGAGGACAGCGCCGGCAAGCACTGGCGCTTCCGCTACTCCTACTGGAACAGCAGC
CAGAGCTACGTCATGACCAAGGGCTGGAGCCGCTTCGTCAAGGAGAAGCGCCTCGTCGCC
GGGGACACCGTCTCCTTCTCCGCGCCGCCGCCGAGGACGCGCGCCACCGCCTCTTCATC
GACTGGAAGCGCGGGTTCGACACCCGCGGCCCGCTTCGTTTCTCCGGCCTCGCGTGCCG
30 ATGCCGCTGCCGTGTCGCACTACGGCGGGCCCCACCACTACAGCCCGTGGGGCTTCGGC
GGCGGCGGCGGCGGCGGCGGCGGATTCTTCATGCCGCCCTCGCCGCCCGCCACGCTCTAC
GAGCACCGCCTCAGACAGGGCCTCGACTTCCGACGATGACGACGACCTACCCCGCGCCG
ACCGTGGGGAGGACGCTCCTGTTTTTCGGCTCGGCCAGGATGCCTCCTCATCACGCGCCG
CCGCCCCAGCCGCGCCCGTTCTCGCTGCCGCTGCATCACTACAGGTGCAACCGAGCGCC
35 GCCGGCGTCACCGCCGCGTCACGGCCGTCCTTCTTGACTCGGTGCCGGTCATCGAGAGC
CCGACGACCGCCGCGAAGCGCGTGCGGCTGTTCCGGCGTCAACCTGGACAACAACCCAGAT
GGCGGCGGCGAGGCTAGCCATCAGGGCGATGATTGTCATTGCAGATGCCCGGGTGGCAG
CAAAGGACTCCAACCTAAGGCTACTAGAATTGCCTCGCCATGGCGGGGAGTCTCCGCG
GCGTCGTCTCCGTGTCGTCGTCTTCTCCTCAAGAGGGAGGCGGTTTCAGCTTTGGATCTC
40 GATCTGTGA

GRMZM2G328742_T01

45 Cover 55% identity 64%
SEQ ID NO: 134
MATN HLSQGHQHPQAWPWGVAMYTNLHYH HQQH HHYEKEH LFEKPLTPSDVGKLNRLVI
PKQHAERYFPLSSSGAGDKGLILCFEDDDDDDEAAAAANKPWFRYSYWTSSQSYVLTKGWS
RYVKEKQLDAGDVVRFQRM RGFGM PDRLFISHSRRGETTATAATTVPAAAARVVVAPA
QSAGADHQQQQPSWSPMCYSTSGSYPTSSPANSQHAYHRHSADH DHSNNMQHAGES
50 QSDRDNRSCSAASAPPPSRRLRLFVN LDCGPGPEPETPTAMYGYMHQSPYAYNNWGS
YQHDEEI

CDS 135

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CCCAAGCAGCACGCCGAGAGGTACTTCCCTCTCAGCAGCAGCGGCGCCGGCGACAAAGGC
5 CTCATCCTGTGCTTCGAGGACGACGACGACGACGAGGCTGCCGCCGCCAACAAAGCCGTGG
CGGTTCCGCTACTCGTACTGGACCAGCAGCCAGAGCTACGTGCTCACCAGGGGCTGGAGC
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10 CAGAGCGCTGGCGCAGACCACCAGCAGCAGCAGCAGCCGTGCGCTTGGAGCCCAATGTGC
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15 ACGGCGATGTACGGCTACATGCACCAAAGCCCCTACGCTTACAACAACCTGGGGCAGTCCA
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- 20 GRMZM2G142999_T01
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35 CTGGTGCTCAGCTTCGAGGACCGGGCGGGGAAGGCGTGCGCTTCCGCTACTCGTACTGG
AACAGCAGCCAGAGCTACGTGATGACCAAAGGTTGGAGCCGCTTCGTGAAGGAGAAGCGC
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GCCTCGGCCGCCGCGGCGTCCGCATCGGCGTCCGGCGGGCGGCGGGAAGGCGGGGCTGGTG
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10 GCCGGGGACACGGTATTGTTTCGCGCGCGGCGCGGGCGCCACGCGCGGGCCGCTTCTTCATC
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CGCCTCCTGCCGCTCCCGTCGGTGCCCATCTGCCCGTGGCAGGGCTACGGCGCCTCCGCT
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20 *Tricum aeseirum*

TRAES3BF098300010CFD_t1

Cover: 42% ident 60%

25 SEQ ID NO: 140
MGVEI LSSMVEHSFYSSGVSTATTESGTAGTPPRPLSLPVAIADESVTSSRSASSRFKGVVPQPNGRWGAQIYERH
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RGMGARAQPTPSWAREPLFEKAVTPSDVGKLNRLVVPKQHAKEH FPLKRTPETPTTTGKGVLLNFEDGEGKVWR
FRYSYWNSSQSYVLTGKWSRFVREKGLGAGDSILFSCSLYEQEKQFFIDCKKNTSMNGKSASPLPVGVTTKGEQV
30 RVVRLFGVDISGVKRGRAATATAEQGLQELFKRQCVAPGQHSALGAFAL

CDS SEQ ID NO: 141

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Cover 47% ident 55%

SEQ ID NO: 142

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DPELGFLADHSAEIVDMLRKHTYDDELRLQGLRRGRGRAQPTPAWARELLFEKAVTPSDV
GKLNRLVVPKQQAIEKHFPTTAAATGSNGKGVLLNFEDGEGKVWFRYSYWNSSQSYVLT
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15 CDS SEQ ID NO: 143

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30 TACCGGTGCGCGTACGGGAATGACACGGAGGATCAGCTCTTCATCGACTACAAGAAGATG
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TGA

35

TRAES3BF062600010CFD_tI
Cover 43% ident 58%

40 SEQ ID NO: 144

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DPELGFLADHSAEIVDMLRKHTYDDELRLQGLRRGRGRAQPTPAWARELLFEKAVTPSDV
GKLNRLVVPKQQAIEKHFPTTAAATGSNGKGVLLNFEDGEGKVWFRYSYWNSSQSYVLT
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GCCGCCGCGCGCCTACGACGTGGCCGCGCTCCGCTTCCGCGGCCCGGACGCCGTCATC

5 AACCACCAGCGACCGACGGCCGCGGAGGAGGCCGGCTCGTCGTCTCCAGGAGCGAGCTG
GATCCAGAGCTCGGCTTCCTCGCCGACCACTCCAAGGCCGAGATCGTCGACATGCTCCGG
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10 GGCAAGCTCAACCGCCTCGTGGTGCCGAAGCAGCAGGCCGAGAAGCACTTCCCTCCGACC
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15 AACAAGAATGACGATGCTGCGGACGCGGCGATTTCCGATGAGAATGAGACAGGCCATGTC
GCCGTCAAGCTCTTCGGCGTTGACATTGCCGGTGGAGGGATGGCGGGATCATCAGGTGGC
TGA

CLAIMS:

1. A plant wherein said plant does not produce a functional NGAL2 polypeptide or does not produce functional NGAL2 and NGAL3 polypeptides.
- 5 2. A plant according to claim 1 wherein the expression of a nucleic acid sequence encoding a NGAL2 polypeptide or the activity of a NGAL2 polypeptide is reduced or abolished.
3. A plant according to claim 1 or 2 wherein the expression of a nucleic acid sequence encoding a NGAL3 polypeptide or the activity of a NGAL3 polypeptide is reduced or abolished.
- 10 4. A plant according to a preceding claim wherein the NGAL2 polypeptide comprises a sequence comprising or consisting of SEQ ID NO: 3, a functional variant or homologue thereof.
5. A plant according to a preceding claim wherein the *SOD7* nucleic acid sequence encoding a NGAL2 polypeptide comprises a nucleic acid sequence comprising or consisting of SEQ ID NO: 1 or 2, a functional variant or homologue thereof.
- 15 6. A plant according to claim 5 wherein the functional variant or homologue comprises a sequence comprising or consisting of SEQ ID No 6 or 7.
- 20 7. A plant according to a preceding claim wherein the NGAL3 polypeptide comprises a sequence comprising or consisting of SEQ ID NO: 5, a functional variant or homologue thereof.
8. A plant according to a preceding claim wherein the *NGAL3* nucleic acid sequence encoding a NGAL3 polypeptide comprises a nucleic acid sequence comprising or consisting of SEQ ID NO: 4, a functional variant or homologue thereof.
- 25 9. A plant according to claim 8 wherein the functional variant or homologue thereof comprises a sequence comprising or consisting of SEQ ID NO: 6 or 7.
10. A plant according to a preceding claim wherein the endogenous *SOD7* nucleic acid sequence or its promoter carries a functional mutation.
- 30 11. A plant according to any of claims 1-9 wherein said plant comprises an RNA interference construct that reduces the expression of *SOD7*, a functional variant or homolog thereof.
- 35 12. A plant according to a preceding claim wherein the endogenous *NGAL3* nucleic acid sequence or its promoter carries a functional mutation.

13. A plant according to any of claims 2 to 11 wherein said plant comprises an RNA interference construct that reduces the expression of *NGAL3*, a functional variant or homolog thereof.
14. A plant according to a preceding claim wherein said plant is a crop plant.
- 5 15. A plant according to a preceding claim wherein said plant is a monocotyledonous plant or dicotyledonous plant.
16. A plant according to a preceding claim wherein said plant is selected from maize, rice, wheat, oilseed rape/canola, sorghum, soybean, sunflower, alfalfa, potato, tomato, tobacco, grape, barley, pea, bean, field bean, lettuce, cotton, 10 sugar cane, sugar beet, broccoli or other vegetable brassicas or poplar.
17. A method for altering a plant phenotype comprising reducing or abolishing the expression of a nucleic acid sequence encoding a *NGAL2* polypeptide or reducing or abolishing the activity of a *NGAL2* or reducing or abolishing the expression of a nucleic acid sequences encoding *NGAL2* and *NGAL3* 15 polypeptides or reducing or abolishing the activity of a *NGAL2* and *NGAL3* polypeptide relative to a control plant.
18. A method for making a plant with an altered phenotype comprising reducing or abolishing the expression of a nucleic acid sequence encoding a *NGAL2* polypeptide or reducing or abolishing the activity of a *NGAL2* or reducing or 20 abolishing the expression of a nucleic acid sequences encoding *NGAL2* and *NGAL3* polypeptides or reducing or abolishing the activity of a *NGAL2* and *NGAL3* polypeptide relative to a control plant..
19. A method according to claim 17 or 18 comprising reducing or abolishing the expression of a nucleic acid sequence encoding a *NGAL2* polypeptide and 25 reducing or abolishing expression of a nucleic acid sequence encoding a *NGAL3* polypeptide.
20. A method according to claim 17 or 18 comprising reducing or abolishing activity of a *NGAL2* polypeptide and reducing or abolishing activity of a *NGAL3* polypeptide.
- 30 21. A method according to claim 17 or 18 comprising reducing or abolishing expression of a nucleic acid sequence encoding a *NGAL2* polypeptide and reducing or abolishing activity of a *NGAL3* polypeptide.
22. A method according to claim 17 or 18 comprising reducing or abolishing expression of a nucleic acid sequence encoding a *NGAL3* polypeptide and 35 reduced or abolished activity of a *NGAL2* polypeptide.

23. A method according to any of claims 17 to 22 wherein the *NGAL2* polypeptide comprises a sequence comprising or consisting of SEQ D NO: 1 or 2, a functional variant or homologue thereof.
- 5 24. A method according to any of claims 17 to 22 wherein the *SOD7* nucleic acid sequence encoding a *NGAL2* polypeptide comprises a nucleic acid sequence comprising or consisting of SEQ ID NO: 1 or 2, a functional variant or homologue thereof.
25. A method according to claim 24 wherein the functional variant or homologue comprises a nucleic acid sequence as shown in SEQ Id NO:49-145.
- 10 26. A method according to any of claims 17-25 wherein the *NGAL3* polypeptide comprises a sequence comprising or consisting of SEQ ID NO: 5, a functional variant or homologue thereof.
27. A method according to claim 17 to 26 wherein the *NGAL3* nucleic acid sequence encoding a *NGAL3* polypeptide comprises a nucleic acid sequence comprising or consisting of SEQ ID NO: 4, a functional variant or homologue thereof.
- 15 28. A method according to claim 27 wherein the functional variant or homologue comprises a nucleic acid sequence comprising or consisting of SEQ ID NOs:49-145.
- 20 29. A method according to any of claims 17 to 28 wherein said method comprises introducing a functional mutation in a nucleic acid sequence encoding a *NGAL2* and/or *NGAL3* protein or peptide in a plant or in its corresponding promoter.
30. A method according to claim 29 wherein said mutation is introduced using T-DNA insertion, chemical mutagenesis or genome editing.
- 25 31. A method according to claim 30 comprising using TILLING.
32. A method according to any of claims 17 to 28 comprising silencing of the *SOD7* and/or *NGAL3* nucleic acid sequence.
33. A method according to claim 32 comprising introducing a RNAi, shRNA, snRNA, dsRNA, siRNA, miRNA, ta-siRNA or co-suppression molecule which targets the *SOD7* or *NGAL3* nucleic acid sequence gene into a plant.
- 30 34. A method according to any of claims 17 to 33 wherein said phenotype is characterised by increased seed size relative to a control plant.
35. A plant obtained or obtainable by the method of any of claims 17 to 34.
36. An isolated nucleic acid comprising a sequence comprising or consisting of SEQ ID NO: 1 or 2 or a functional variant or homologue thereof.
- 35 37. A vector comprising an isolated nucleic acid according to claim 36.

38. A silencing nucleic acid construct targeting sequence comprising or consisting of SEQ ID NO: 1, 2 or 3 or a functional variant, part or homologue thereof.

Figure 1

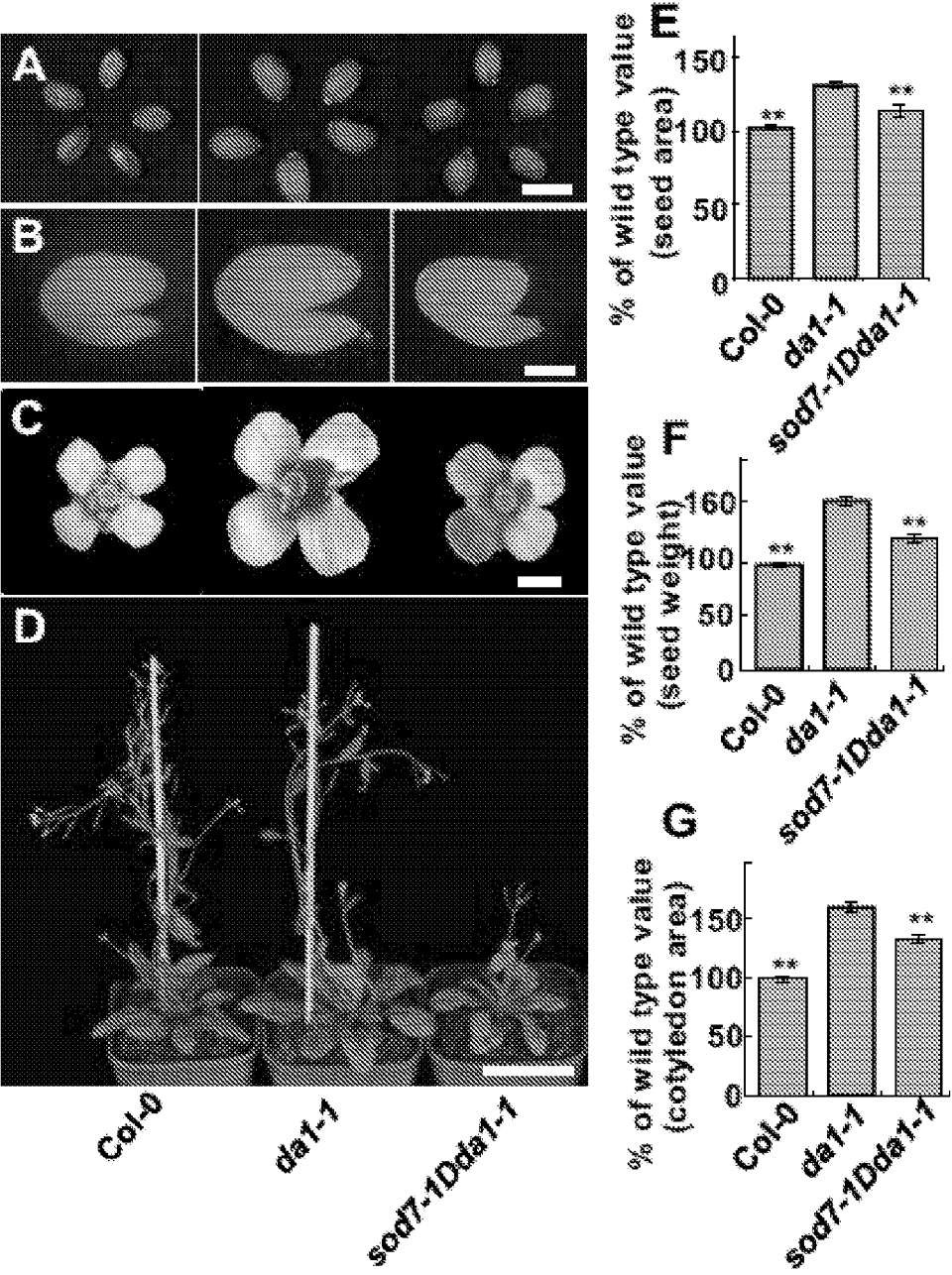


Figure 2

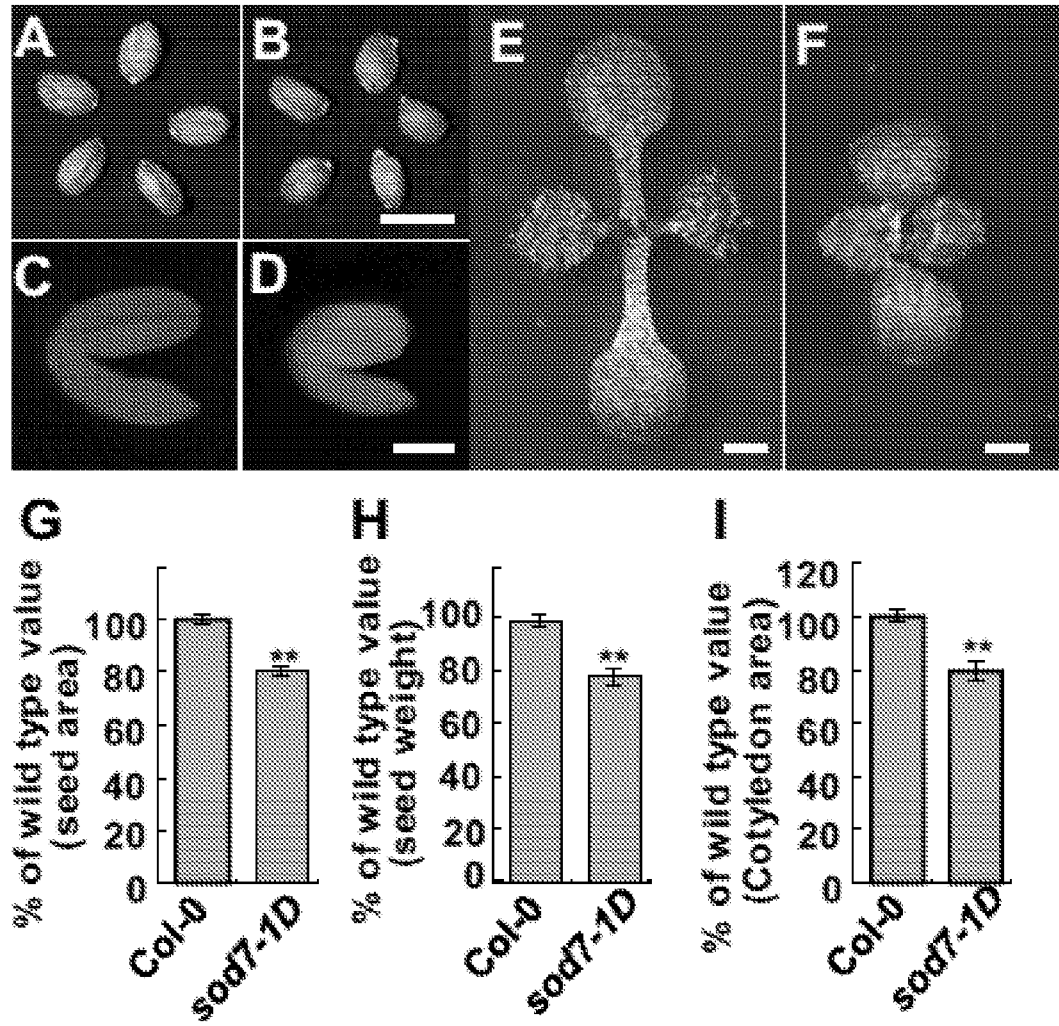


Figure 3

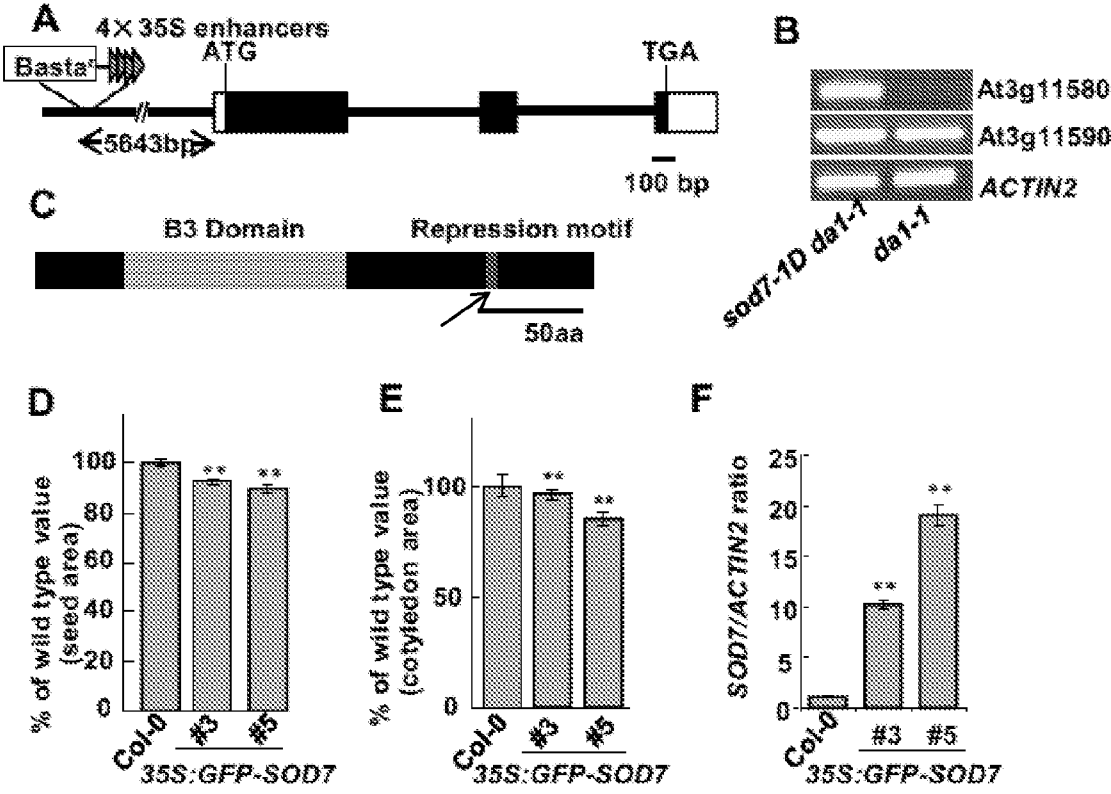


Figure 4

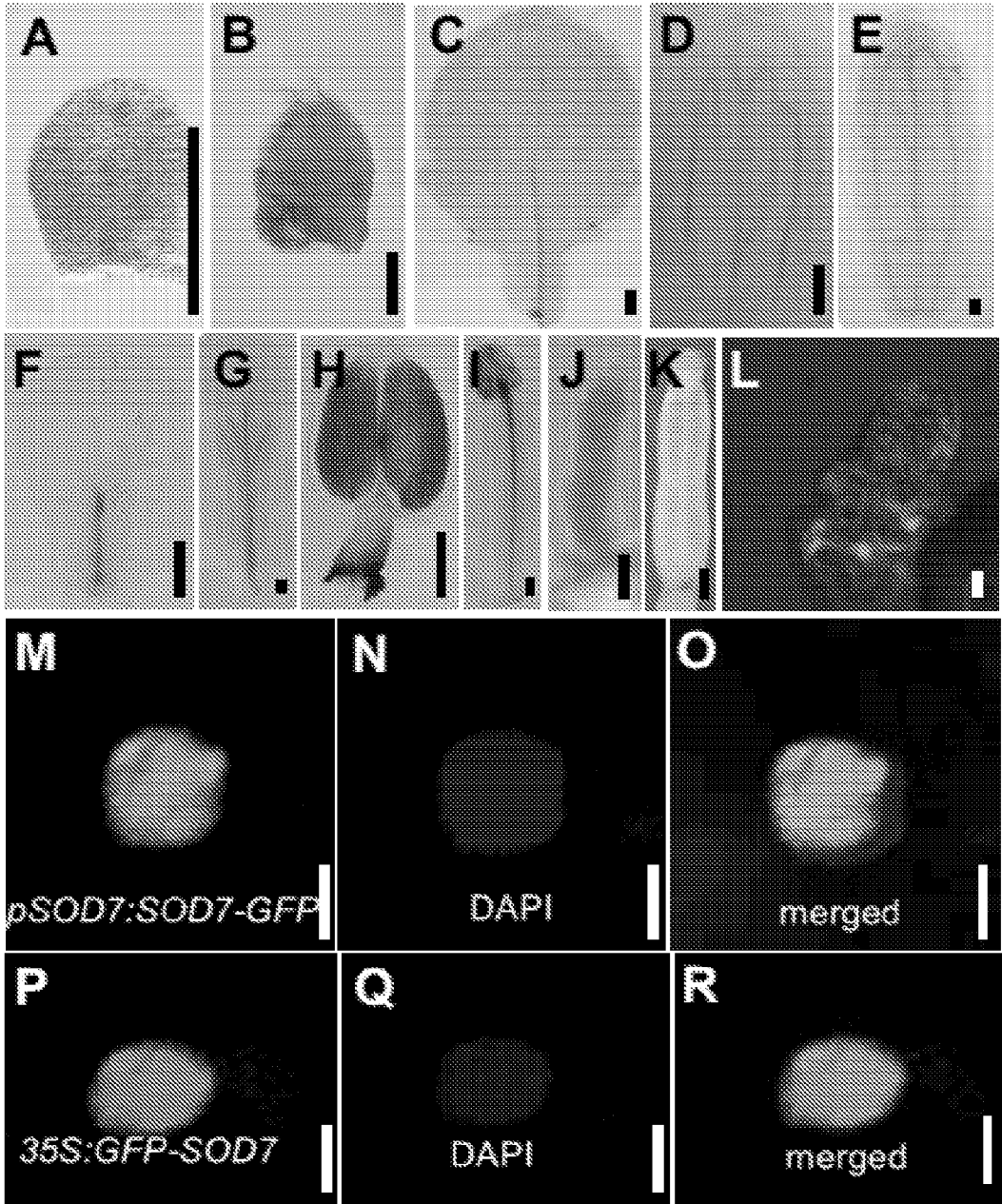


Figure 5

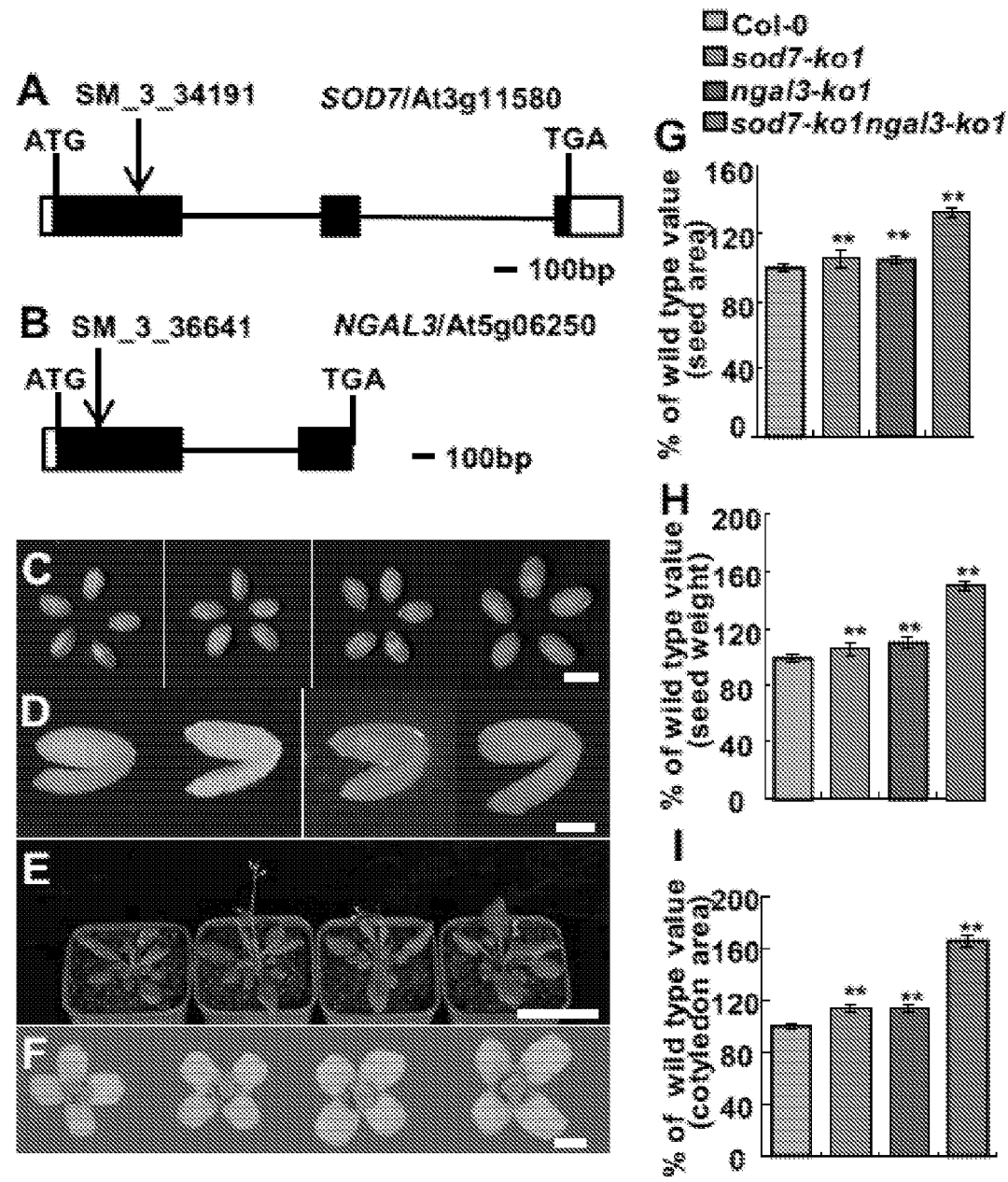


Figure 6

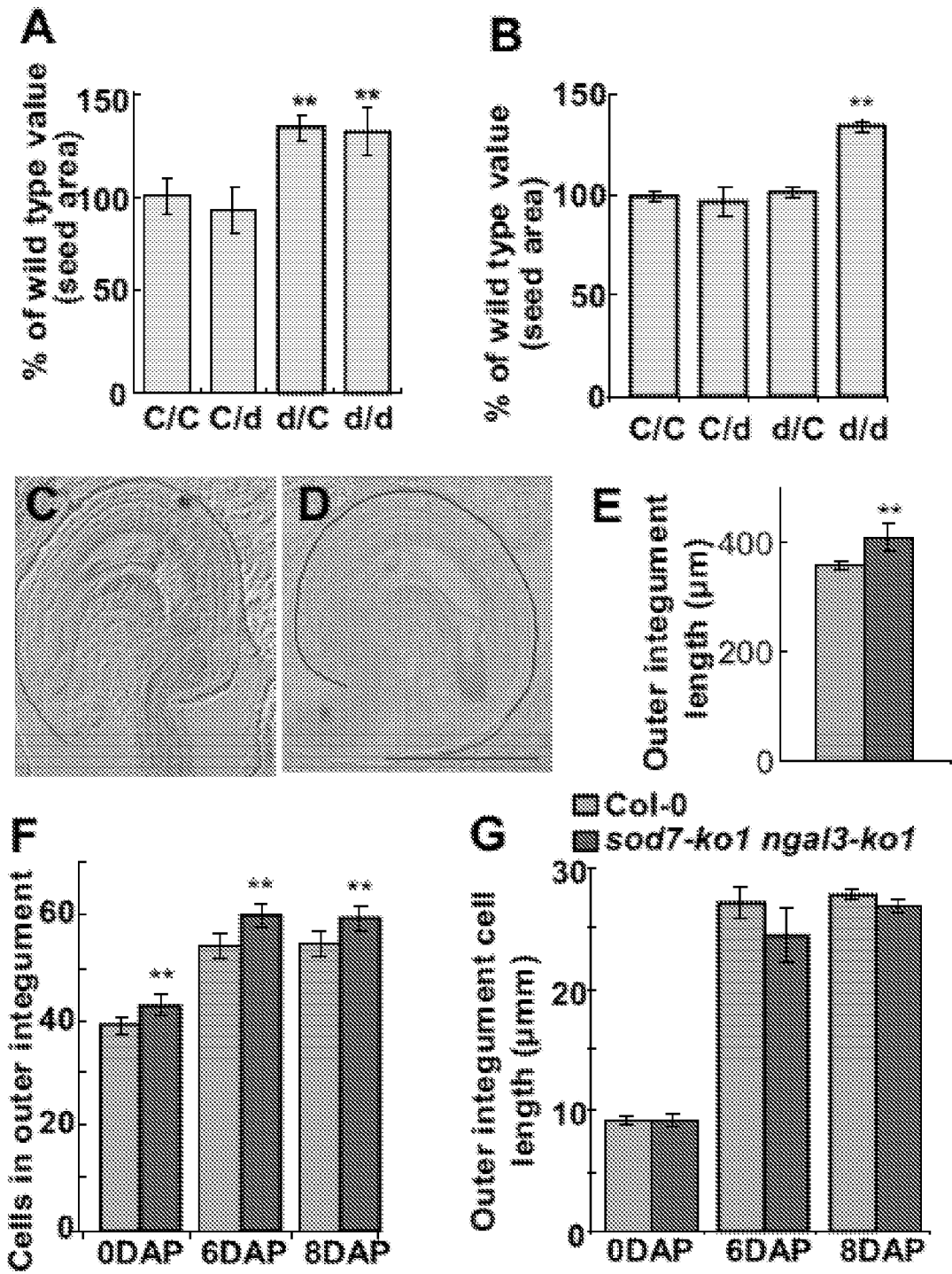


Figure 7

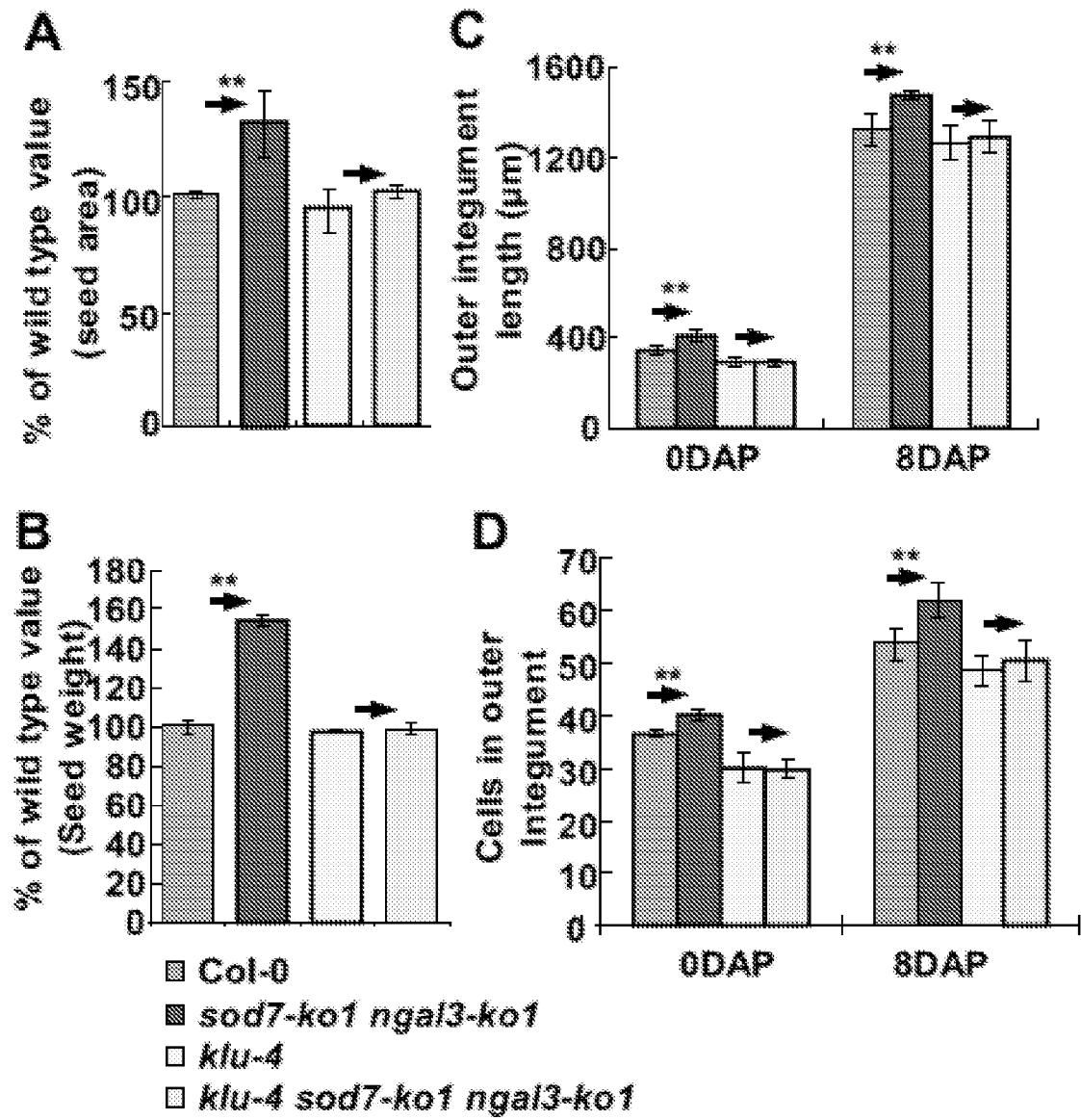


Figure 8

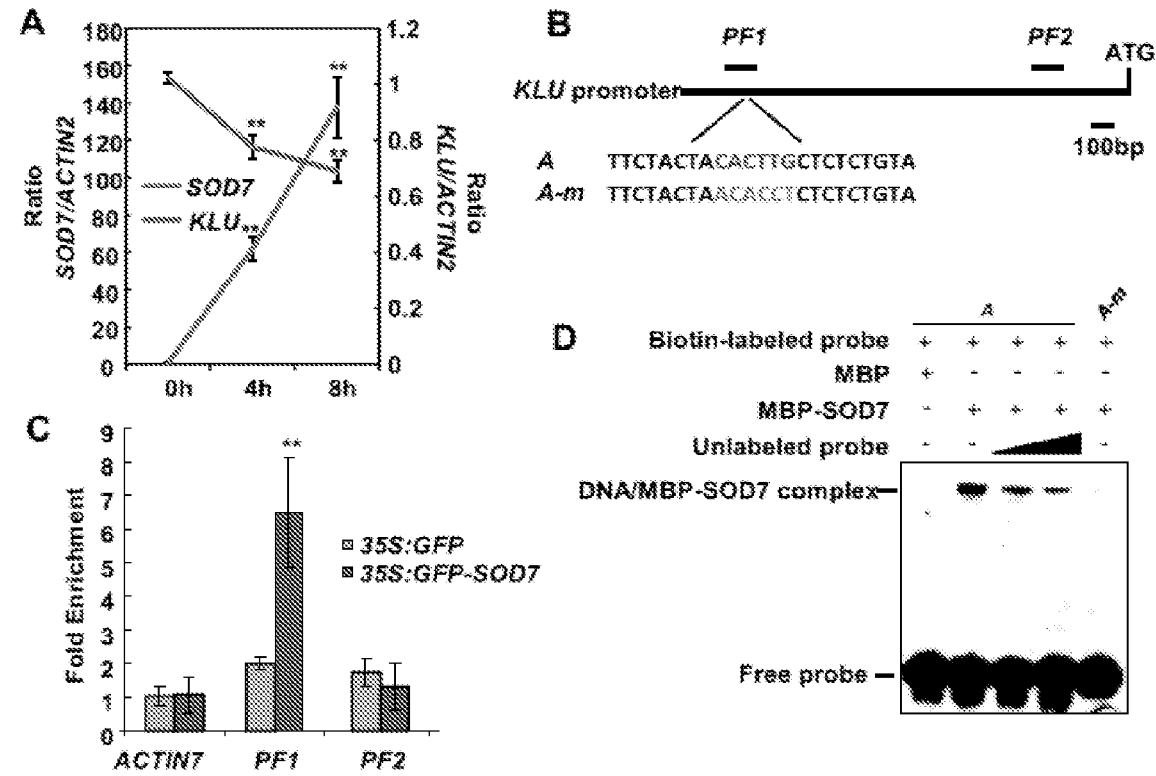
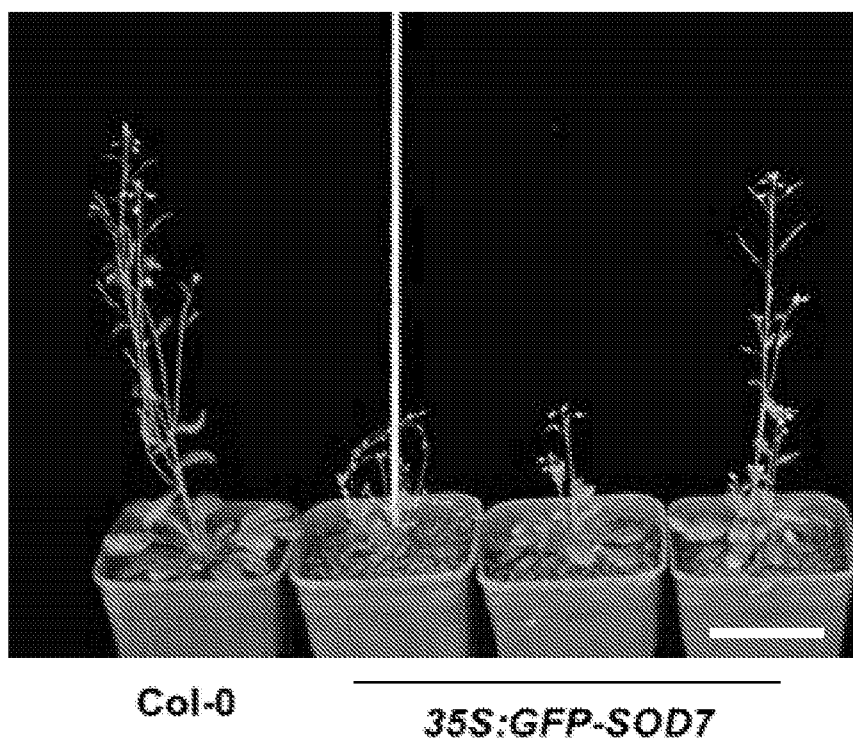


Figure 9



10/29

Figure 10

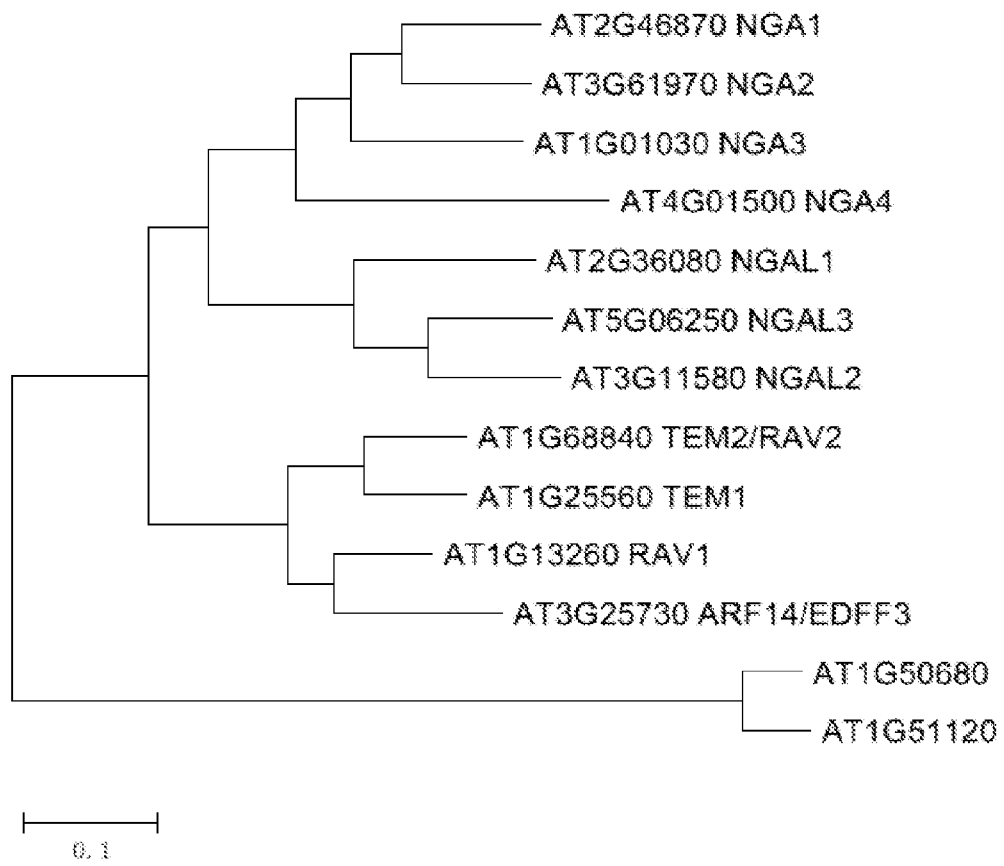


Figure 11

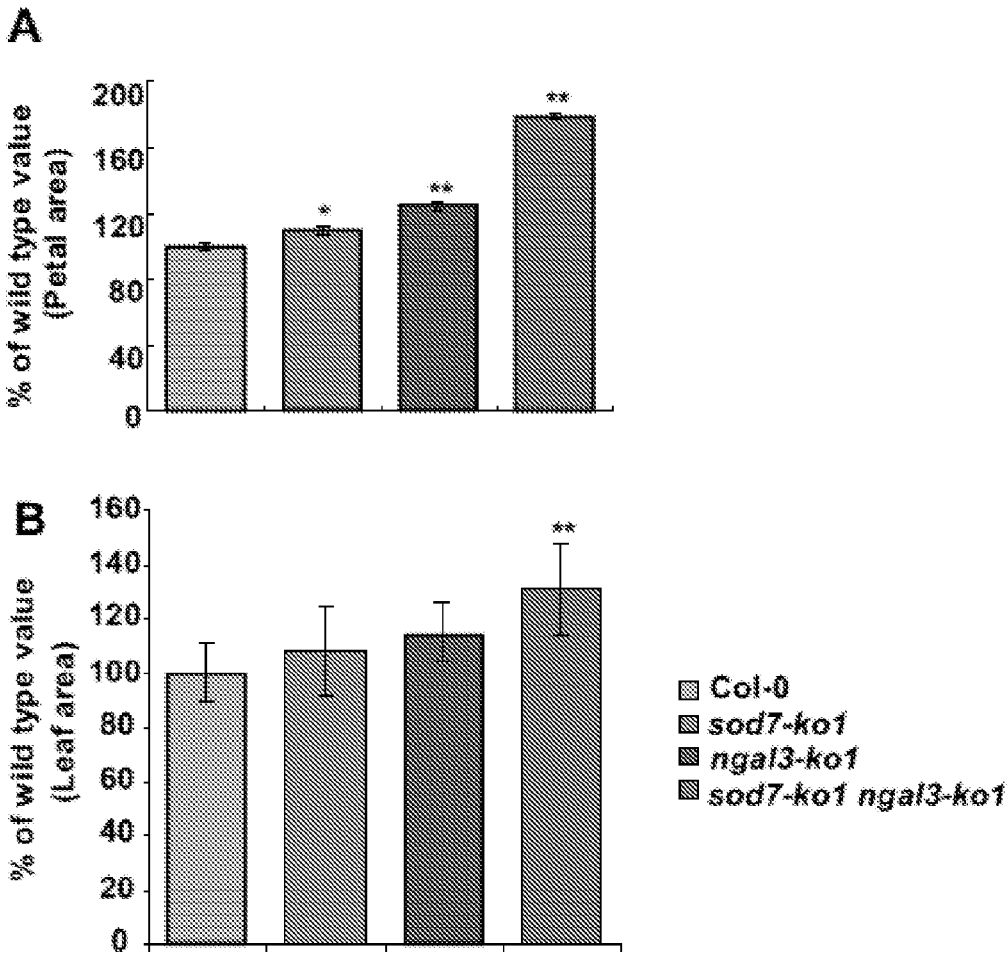


Figure 12

A

SOD7	1	FEKSLTPSDVGKLNRLVIPKQHAKEYFPLNNNNNNNGSGDDVA	EKG	LLSFEDES
Brassica rapa2	1	FEKSLTPSDVGKLNRLVIPKQHAKEYFPLNN--CGGGGDVTAE	EKG	LLSFEDES
Glycine max.At3g11580-likel		FEKPLTPSDVGKLNRLVIPKQHAKEYFPLS-----DSGG	ECKGLLSFEDES	
Glycine max At5g06250-likel		FEKPLTPSDVGKLNRLVIPKQHAKEYFPLS--G-----DSGG	ECKGLLSFEDES	
Glycine max At2g36080-likel		FEKPLTPSDVGKLNRLVIPKQHAKEYFPLDSS-----GGD	AAAKGLLSFEDES	
Oryza sativa	1	FEKPLTPSDVGKLNRLVIPKQHAKEYFPLG-----GD	GE	KGLLSFEDES
At5g06250/NAGL3	1	FEKSLTPSDVGKLNRLVIPKQHAKEYFPLNVLVS-SAAADT	EKG	LLSFEDES
Hordeum vulgare1	1	EKVLTPSDVGKLNRLVIPKQHAKEYFPLD-----AANE	EKGLLSFEDRG	
Zea mays Os02g0683500	1	EKVLTPSDVGKLNRLVIPKQHAKEYFPLD-----AANE	EKGLLSFEDRA	
Zea mays Os02g0683500-likel		EKVLTPSDVGKLNRLVIPKQHAKEYFPLD-----AANE	EKG	LLSFEDES
Hordeum vulgare2	1	EKVLTPSDVGKLNRLVIPKQHAKEYFPLD-----	KGLLSFEDRA	
Gossypium hirsutum RAV	1	FEKALTPSDVGKLNRLVIPKQHAKEYFPLQS-----GA	SKG	LLNFEDV
Triticum aestivum	1	FEKALTPSDVGKLNRLVIPKQHAKEYFPLK RTP-----ETP	GKG	LLNFEDGE

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56	GKSWFRYSYWNSSQSYVLTGWSR	VK	KHLNAGDVVLFQRHR--FDIHLRLFIGWRRRGE	114	SEQ ID NO 261
50	GKQWFRYSYWNSSQSYVLTGWSR	VK	KRLDAGDVVLFQRHR--VDAQRLFIGWRRR--	106	SEQ ID NO 262
51	GKQWFRYSYWNSSQSYVLTGWSR	VK	KRLDAGDVVLFQRHR--DAQRLFIGWRRR--	107	SEQ ID NO 263
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57	GKSWFRYSYWNSSQSYVLTGWSR	VK	KOLDPGDVVFQRHR--SDSRRLFIGWRRRQ	115	SEQ ID NO 266
47	GKQWFRYSYWNSSQSYVLTGWSR	VK	KRLDAGDVVFQRHR--TRDRLFIGWRRRGE	107	SEQ ID NO 267
47	GKQWFRYSYWNSSQSYVLTGWSR	VK	KRLDAGDVVFQRHR--ARDRLFIGWRRRGE	105	SEQ ID NO 268
47	GKQWFRYSYWNSSQSYVLTGWSR	VK	KRLDAGDVVFQRHR--ARDRLFIGWRRRGE	107	SEQ ID NO 269
47	GKQWFRYSYWNSSQSYVLTGWSR	VK	KRLDAGDVVFQRHR--ARGRLFIGWRRRGE	107	SEQ ID NO 270
48	GKQWFRYSYWNSSQSYVLTGWSR	VK	KRLDAGDVVFQRHR--TEKQLFIGWRRRGE	104	SEQ ID NO 271
52	GKQWFRYSYWNSSQSYVLTGWSR	VK	KRLDAGDVVFQRHR--YEQLFIGWRRRGE	102	SEQ ID NO 272

B

SEQ ID NO 164	LOC_Os04g49230	1	DRLHIDWKRR
SEQ ID No 165	Bra007646	1	RLFGVD--
SEQ ID No 166	sGmLoc100795470	1	RLFGVD--
SEQ ID No 167	Bra000434	1	RLFGVD--
SEQ ID No 168	Bra040478	1	RLFGVD--
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SEQ ID No 177	Bra017262	1	RLFGVN--
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SEQ ID No 180	HvMLOC_56567	1	VRLFGVD	A-
SEQ ID No 181	Bra038346	1	VRLFGVD	F-
SEQ ID No 182	TRAES3BF098300010CFD_t1	1	VRLFGVD	S-
SEQ ID No 183	GmLoc100776987	1	VRLFGVN	L-
SEQ ID No 184	GmLoc100801107	1	VRLFGVN	L-
SEQ ID No 185	os01g0693400	1	VRLFGVDLL	-
SEQ ID No 186	GmLoc100789009	1	VRLFGVDLL	-
SEQ ID No 187	HvMLOC44012	1	VRLFGVDLL	-
SEQ ID No 188	HvMLOC_38822	1	VRLFGVDLL	-
SEQ ID No 189	GmLoc732601	1	VRLFGVNLL	-
SEQ ID No 190	BrLOC103849927	1	VRLFGVNL	-
SEQ ID No 191	Bra034828	1	VRLFGVNL	-
SEQ ID No 192	Bra005886	1	VRLFGVNL	-
SEQ ID No 193	SOD7	1	VRLFGVNL	-
SEQ ID No 194	At5g06250/NGAL3	1	VRLFGVNL	-
SEQ ID No 195	LOC_Os11g05740.1	1	VRLFGVNL	-
SEQ ID No 196	GRMZM2G328742_T01	1	VRLFGVNL	-
SEQ ID No 197	os02g0683500	1	VRLFGVNL	-
SEQ ID No 198	LOC_Os03g02900	1	VRLFGVNL	-
SEQ ID No 199	Os10g0537100	1	VRLFGVNL	-
SEQ ID No 200	HvMLOC_66387	1	VRLFGVNL	-
SEQ ID No 201	GRMZM2G102059_T01	1	VRLFGVNL	-
SEQ ID No 202	GRMZM2G082227_T01	1	VRLFGVNL	-
SEQ ID No 203	GRMZM2G024948_T01	1	VRLFGVNL	-
SEQ ID No 204	GRMZM2G142999_T01	1	VRLFGVNL	-
SEQ ID No 205	GRMZM2G125095_T01	1	VRLFGVNL	-

Figure 13

GRMZM2G053008	SEQ ID NO 207	1	-----MAAS-----PSSPLTAPPEPV
HvMLOC_57250	SEQ ID NO 208	1	-----
Os12g0157000	SEQ ID NO 209	1	-----
GmLoc100778733	SEQ ID NO 210	1	-----
Bra004501	SEQ ID NO 211	1	-----
Bra000434	SEQ ID NO 212	1	-----
Bra040478	SEQ ID NO 213	1	-----
Bra014415	SEQ ID NO 214	1	-----
Bra003482	SEQ ID NO 215	1	-----
Bra007646	SEQ ID NO 216	1	-----
GlycinemaxLoc100781489	SEQ ID NO 217	1	-----
GRMZM2G024948_T01	SEQ ID NO 218	1	-----
os02g0683500	SEQ ID NO 219	1	-----
HvMLOC_66387	SEQ ID NO 220	1	-----
os04g0581400	SEQ ID NO 221	1	-----
GRMZM2G102059_T01	SEQ ID NO 222	1	-----
Os10g0537100	SEQ ID NO 223	1	-----
GRMZM2G142999_T01	SEQ ID NO 224	1	-----
GRMZM2G125095_T01	SEQ ID NO 225	1	-----
os03g0120900	SEQ ID NO 226	1	-----
GRMZM2G098443_T01	SEQ ID NO 227	1	-----
GRMZM2G082227_T01	SEQ ID NO 228	1	-----
Os11g0156000	SEQ ID NO 229	1	-----
GRMZM2G328742_T01	SEQ ID NO 230	1	-----
GmLoc100802734	SEQ ID NO 231	1	-----
GmLoc100795470	SEQ ID NO 232	1	-----
GmLoc100818164	SEQ ID NO 233	1	-----
Bra017262	SEQ ID NO 234	1	-----
At2g36080	SEQ ID NO 235	1	-----
Bra005301	SEQ ID NO 236	1	-----
At3g11580	SEQ ID NO 237	1	-----
BraLOC103849927	SEQ ID NO 238	1	-----
BrassicarapaBra034828	SEQ ID NO 239	1	-----
At5g06250	SEQ ID NO 240	1	-----
Bra005886	SEQ ID NO 241	1	-----
GmLoc102660503	SEQ ID NO 242	1	-----MIGVEKVTICMREVNT-----EKGRR-----A
HvMLOC_38822	SEQ ID NO 243	1	-----
os01g0693400	SEQ ID NO 244	1	-----MDSS-SCLVDDTNSGGSS-----TDKL-----RALAAAAAET
HvMLOC44012	SEQ ID NO 245	1	-----
HvMLOC_7940	SEQ ID NO 246	1	-----MGVEIL-SSTGEHSS-----QYSSGA-STAT-ESGVGGRPTAP
HvMLOC_75135	SEQ ID NO 247	1	-----MGMEIL-SSTVEHCS-----QYSSA-STAT-ESGAAGRSTTAL
TRAECDM81004	SEQ ID NO 248	1	-----MGVEIL-SSMVEDSS-----QYSSGA-STAT-ESGTTGRALTAL
HvMLOC_56567	SEQ ID NO 249	1	-----MGVEIL-SSMVEHSF-----QYSSGA-SSATA-ESGAVGTPPRHL
TRAES3BF098300010CFD_t1	SEQ ID NO 250	1	-----MGVEIL-SSMVEHSF-----QYSSGV-STAT-ESGTAGTTPPRPL
HvMLOC_63261	SEQ ID NO 251	1	MASSKPTNP--EVD-ND----M-----ECSS-----P-----ESGAEDAV-ESS
TRAES3BF062700040CFD_t1	SEQ ID NO 252	1	MASGKPTNHGMEDD-ND----M-----EYSS-----A-----ESGAEDAA-EPS
TRAES3BF062600010CFD_t1	SEQ ID NO 253	1	MASGKPTNHGMEDD-ND----M-----EYSS-----A-----ESGAEDAA-EPS
Bra038346	SEQ ID NO 254	1	-----MVF-SCIDESSST--SESFSPAT-ATAT-----ATATKFSAPPLPP
GmLoc732601	SEQ ID NO 255	1	-----MDGG-CVTDETT-TSSDS-----LSV-----PP-
GmLoc100789009	SEQ ID NO 256	1	-----MDGG-SVTDETT-TTSNS-----LSVPANLSPPP-
GmLoc100776987	SEQ ID NO 257	1	-----MDAI-SCLDESTTTESLSIS-----QAKPSSTIMSSEKASPPPPP
GmLoc100801107	SEQ ID NO 258	1	-----MDAI-SCMDESTTTESLSISLSPTS--SSEKAKPSSMITSSSEKVSLSPPPS

GRMZM2G053008	17	TPPSPWTITDGAISGTLPAAEAFVHYPGYPSSP---ARAARTLGGLPGLAKVRSSDPGA
HvMLOC_57250	1	-----
Os12g0157000	1	-----
GmLoc100778733	1	-----
Bra004501	1	-----
Bra000434	1	-----
Bra040478	1	-----
Bra014415	1	-----
Bra003482	1	-----
Bra007646	1	-----
GlycinemaxLoc100781489	1	-----
GRMZM2G024948_T01	1	-----
os02g0683500	1	-----
HvMLOC_66387	1	-----
os04g0581400	1	-----
GRMZM2G102059_T01	1	-----
Os10g0537100	1	-----
GRMZM2G142999_T01	1	-----
GRMZM2G125095_T01	1	-----
os03g0120900	1	-----
GRMZM2G098443_T01	1	-----
GRMZM2G082227_T01	1	-----
Os11g0156000	1	-----
GRMZM2G328742_T01	1	-----
GmLoc100802734	1	-----
GmLoc100795470	1	-----
GmLoc100818164	1	-----
Bra017262	1	-----
At2g36080	1	-----
Bra005301	1	-----
At3g11580	1	-----
BraLOC103849927	1	-----
BrassicarapaBra034828	1	-----
At5g06250	1	-----
Bra005886	1	-----
GmLoc102660503	24	LMDC-WQIS-G-----VHESSDC-SEIK----FAFDVVKRAR---HEENN
HvMLOC_38822	1	-----
os01g0693400	32	APLE-RMGS-GA-----SAVVDAEPGAEADSGSGGRVCGGGGGGAG---GAGGK
HvMLOC44012	1	-----
HvMLOC_7940	39	SLP-----VSIAD--ES-----ATSR---SASAQ
HvMLOC_75135	38	SLP-----VAITD--ES-----VTSR---SASAQ
TRAECDM81004	38	SLP-----VAIAD--ES-----VTS-----AQ
HvMLOC_56567	38	SLP-----VAIAD--ES-----LTS-----R
TRAES3BF098300010CFD_t1	38	SLP-----VAIAD--ES-----VTS-----R
HvMLOC_63261	32	-----SPVA
TRAES3BF062700040CFD_t1	34	SSP-----VL---APPra
TRAES3BF062600010CFD_t1	34	SSP-----VL---APPra
Bra038346	38	LRLN-RMRS-GG-----SNVVLDKNG-----VD---IDSRK
GmLoc732601	22	--PS-RVGS-VA-----SAVVDPDGCCVS-----GE---AESRK
GmLoc100789009	28	--LS-LVGS-GA-----TAVVYPDGCCVS-----GE---AESRK
GmLoc100776987	41	NRLC-RVGS-GA-----SAVVDSGGGGG-----GSTE---VESRK
GmLoc100801107	49	NRLC-RVGS-GA-----SAVVDPDGGGSG-----AE---VESRK

```
GRMZM2G053008      74 RLELRFRPEDPYCHPAFGQSRAS---GLLLRLSKRKGAAPCAHVVA-----
HvMLOC_57250        1 -----
Osl2g0157000        1 -----
GmLoc100778733      1 -----MELMQ-EVKG-YSDGREEEEEEEAEEII-----
Bra004501           1 -----
Bra000434           1 -----
Bra040478           1 -----
Bra014415           1 -----
Bra003482           1 -----
Bra007646           1 -----
GlycinemaxLoc100781489 1 -----MELMQ-QVKGNYSDSREEEE-----
GRMZM2G024948_T01  1 -----MDQFA--ASGRFSREEEADE-----
os02g0683500        1 -----MEFTT---SSRFSKE--EED-----
HvMLOC_66387        1 -----MEFTA--TSSRFSKGEEVE-----
os04g0581400        1 -----MEFAT--TSSRFSKEEEEEEGEQEMEQQ-----
GRMZM2G102059_T01  1 -----MEFAS--SSSRFSREEDEEEE-----Q-----
Osl0g0537100        1 -----
GRMZM2G142999_T01  1 -----
GRMZM2G125095_T01  1 -----
os03g0120900        1 -----
GRMZM2G098443_T01  1 -----
GRMZM2G082227_T01  1 -----
Osl1g0156000        1 -----
GRMZM2G328742_T01  1 -----
GmLoc100802734      1 -----
GmLoc100795470      1 -----
GmLoc100818164      1 -----
Bra017262           1 -----
At2g36080           1 -----
Bra005301           1 -----
At3g11580           1 -----
BraLOC103849927     1 -----
BrassicarapaBra034828 1 -----
At5g06250           1 -----
Bra005886           1 -----
GmLoc102660503      60 AAAQKFKGVVSQQNGNWGAQIYAHQQRIWLGTfKsEREAMAYDSASIKLRSGECHRNfP
HvMLOC_38822        1 -----
os01g0693400      77 LPSSKFKGVVPQPNGRWGAQIYERHQRVWLGTfAGEDDAARAYDVAAQRfRGRDAVTNfR
HvMLOC44012         1 -----
HvMLOC_7940         58 STSSRFKGVVPQPNGRWGAQIYERHARVWLGTfPDEDsAARAYDVAAALRYRGREAATNfP
HvMLOC_75135        57 PASSRFKGVVPQPNGRWGSQIYERHARVWLGTfPDQDsAARAYDVASLRYRGRDAATNfP
TRAECDM81004        53 SAPSRFKGVVPQPNGRWGSQIYERHARVWLGTfPDQDLAARAYDVAAALRYRGRDAATNfP
HvMLOC_56567        52 SASSRFKGVVPQPNGRWGAQIYERHARVWLGTfPDQDsAARAYDVASLRYRGDDAAfNfP
TRAES3BF098300010CFD_t1 52 SASSRFKGVVPQPNGRWGAQIYERHARVWLGTfPDQDsAARAYDVASLRYRGRDVAfNfP
HvMLOC_63261        36 APSSRFKGVVPQPNGRWGAQIYEKHSRVWLGTfGDEEEAAACAYDVAAALRfRGRDAVTNHQ
TRAES3BF062700040CFD_t1 44 APSSRFKGVVPQPNGRWGAQIYEKHSRVWLGTfPDEDAAVRAYDVAAALRfRGPDAVINHQ
TRAES3BF062600010CFD_t1 44 APSSRFKGVVPQPNGRWGAQIYEKHSRVWLGTfPDEDAARAYDVAAALRfRGPDAVINHQ
Bra038346           65 LSSSKYKGVVPQPNGRWGAQIYVKKHQRVWLGTfCDEEEAAHSYDIAARKfRGRDAVVNfK
GmLoc732601         49 LPSSKYKGVVPQPNGRWGAQIYEKHQRVWLGTfNEEDEAARAYDIAALRfRGPDAVTNfK
GmLoc100789009      55 LPSSKYKGVVPQPNGRWGAQIYEKHQRVWLGTfNEEDEAARAYDIAAHRfRGRDAVTNfK
GmLoc100776987      72 LPSSKYKGVVPQPNGRWGSQIYEKHQRVWLGTfNEEDEAARAYDVAVQRfRGRKDAVTNfK
GmLoc100801107      78 LPSSKYKGVVPQPNGRWGAQIYEKHQRVWLGTfNEEDEAARAYDIAAQRfRGRKDAVTNfK
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GRMZM2G053008	119	-----RRTAYFEGMADFQ---HVVPVHA
HvMLOC_57250	1	-----
Os12g0157000	1	-----
GmLoc100778733	29	-----TREESSRLHQQHQAAGSNFI>NNNNHHHQHHH-----
Bra004501	1	-----
Bra000434	1	-----
Bra040478	1	-----
Bra014415	1	-----
Bra003482	1	-----
Bra007646	1	-----
GlycinemaxLoc100781489	20	-----EE-----EAA-----
GRMZM2G024948_T01	19	-----EQE-----DAS-----
os02g0683500	16	-----EE-----QDE-----
HvMLOC_66387	19	-----EE-----QEE-----
os04g0581400	29	-----DEEEE-----EAE-----
GRMZM2G102059_T01	21	-----EEEE-----EEE-----
Os10g0537100	1	-----
GRMZM2G142999_T01	1	-----
GRMZM2G125095_T01	1	-----
os03g0120900	1	-----
GRMZM2G098443_T01	1	-----
GRMZM2G082227_T01	1	-----
Os11g0156000	1	-----AMNHPL-FSQEQP---QSWPWGV
GRMZM2G328742_T01	1	-----ATNHLSQGQHQP---QAWPWGV
GmLoc100802734	1	-----MSSINHY---SPETT---LYWTNDQ
GmLoc100795470	1	-----SINHYSMDLPEPT---LWWPHPH
GmLoc100818164	1	-----STNHYSMDLPEPT---LWWPHPH
Bra017262	1	-----SINQYSSSEFYH---SLMWQQQQ
At2g36080	1	-----SINQYSSDFHYH---SLMWQQQQ
Bra005301	1	-----SINQYSSDFNYH---SLMWQQQQ
At3g11580	1	-----SVNHYNH---TLS---LH---
BraLOC103849927	1	-----SGNHYSRDIHHTPS---V---
BrassicarapaBra034828	1	-----SVNHYSNTL---S---SH---
At5g06250	1	-----SVNHYSTDHHHTL---LWQQQQ
Bra005886	1	-----SVNHYSTDHHQVHHHTLFLQ---
GmLoc102660503	120	WNDQ-----TVQEPQFQSHYSAETVLN-MRDGTWPSKFAT-----
HvMLOC_38822	1	-----MRKHITPDELAQ-----
os01g0693400	137	PLAEADP-----DAAELRFLATRSKAEVVD-MRKHTYFDELAQ-----
HvMLOC44012	1	-----MRKHITPDELAQ-----
HvMLOC_7940	118	CAA-----AEAELAFLAHSAEIVD-MRKHTYFDELRLQ-----
HvMLOC_75135	117	CAA-----AEAELAFLAHSAEIVD-MRKHTYFDELRLQ-----
TRAECM81004	113	CAA-----AEAELAFLAHSAEIVD-MRKHTYFDELRLQ-----
HvMLOC_56567	112	CVV-----VEAELAFLAHSAEIVD-MRKQTYADELRQ-----
TRAES3BF098300010CFD_t1	112	CAA-----VEGELAFLAHSAEIVD-MRKQTYADELRQ-----
HvMLOC_63261	96	RLPAAEGAGWSS-----TSELAFADHSAEIVD-MRKHTYDDELRLQ-----
TRAES3BF062700040CFD_t1	104	RPTAAEEAGSSSRSELDPELGFLADHSAEIVD-MRKHTYDDELRLQ-----
TRAES3BF062600010CFD_t1	104	RPTAAEEAGSSSRSELDPELGFLADHSAEIVD-MRKHTYDDELRLQ-----
Bra038346	125	TFLASED-----DNGELCFLEAHSAEIVD-MRKHTYADELAQ-----
GmLoc732601	109	PPAASD-----DAESEFLNSHSEIVD-MRKHTYDDELRLQ-----
GmLoc100789009	115	PLAGAD-----DAEAEFLSTHSEIVD-MRKHTYDDELRLQ-----
GmLoc100776987	132	PLSGTDD-----DDGESEFLNSHSEIVD-MRKHTYDDELEQ-----
GmLoc100801107	138	PLAGADD-----DDGESEFLNSHSEIVD-MRKHTYDDELEQ-----

GRMZM2G053008
HvMLOC_57250
Osl2g0157000
GmLoc100778733
Bra004501
Bra000434
Bra040478
Bra014415
Bra003482
Bra007646
GlycinemaxLoc100781489
GRMZM2G024948_T01
os02g0683500
HvMLOC_66387
os04g0581400
GRMZM2G102059_T01
Osl0g0537100
GRMZM2G142999_T01
GRMZM2G125095_T01
os03g0120900
GRMZM2G098443_T01
GRMZM2G082227_T01
Osl1g0156000
GRMZM2G328742_T01
GmLoc100802734
GmLoc100795470
GmLoc100818164
Bra017262
At2g36080
Bra005301
At3g11580
BraLOC103849927
BrassicarapaBra034828
At5g06250
Bra005886
GmLoc102660503
HvMLOC_38822
os01g0693400
HvMLOC44012
HvMLOC_7940
HvMLOC_75135
TRAECDM81004
HvMLOC_56567
TRAES3BF098300010CFD_t1
HvMLOC_63261
TRAES3BF062700040CFD_t1
TRAES3BF062600010CFD_t1
Bra038346
GmLoc732601
GmLoc100789009
GmLoc100776987
GmLoc100801107

141 AQT~~Q~~KR-----KHSDSQNDNENF
1 -----MYC-SR-----GRIDPAE-----E
1 -----
63 HTT~~Q~~LD~~F~~MDLSLGSSKDEGNLQ-----GSSSSVYAHH-----HHAASA--SS--SAN
1 -----MMTNLS-----L--SRE
1 -----MMTNLS-----L--ARE
1 -----MMTNLS-----L--ARE
1 -----MERKSNL-----ERSE--NI--DSQ
1 -----
1 -----
25 AIT~~E~~SESSRLHQ-----DTASNFGKKLDLMDLSLGS-SKEE---EEE
25 NSM~~E~~ISFMPPAAASSSSAAASASA-----SASTSASACASGSSSAPFRSASASG---DAA
21 AG~~E~~IPFMTATAEAAAPTSSSSPAHHAASASASASAS-GSSTPFRSD-----DGA
24 AS~~E~~IPFMT~~P~~AAATCAA-----APPSASASASTPASAS-GSSPPFRSG-----DDA
37 AS~~E~~IPFMTSAAAAATASSSSPTSV-SPSATAASAAA-STASGSPFRSS-----DGA
29 AS~~E~~IPFMTAAATADTGAAASSSS--PSA-----A-ASSGPAAAPRSS-----DGA
1 -----MEFTPIS-----PP-TRVAGGEE--D--
1 -----MEFTPAH-----AH-ARVVE-----D--
1 -----MEFRPA-----H-ARVFE-----D--
1 -----MEFITPIVR-----PASAAAGGGEV---QE-
1 -----MEFTTPP-----PATRSGGEE---RA-
1 -----MEFTAPP-----PATRSGGEE---RA-
21 AM-----
22 AM-----
20 QQQAAMWLSNSH-----TPRFNLNDEEEEDDV
22 HQQQLTLMDPD-----PLRLNLNSDDG-NGNDN
22 Q--QQLTLIDPD-----PLPLNLNDDNDNGDDN
22 Q--~~H~~-----
22 QQ--Q-----
22 HRH~~H~~-----
14 HHH-----
18 -HH-----
14 NHH-----
21 HRHTT-----
23 NLHTT-----
155 -FL~~T~~RQTQKGV-----AK-----
14 -H~~K~~AFFFAAAS-----SPTSSS--S-----
176 -SK~~T~~FAASTPS-----AATTTA--SL-----
14 -SK~~A~~FAASAAL-----SAPPTS-----
152 -GL~~R~~RGRGMGAR-----
151 -GL~~R~~RGRGMGAR-----
147 -GL~~R~~RGRGMGAR-----
146 -GL~~R~~RGRGMGVR-----
146 -GL~~R~~RGRGMGAR-----
138 -GL~~R~~RGHG---R-----
151 -GL~~R~~RGRG---R-----
151 -GL~~R~~RGRG---R-----
163 -SN~~R~~SGANTN-----
145 -ST~~G~~GGR--RR-----
151 -ST~~G~~GGR--RR-----
170 -SK~~S~~RGFVRRR-----GSAA-----
176 -SK~~S~~RGVRRR-----GSAA-----

GRMZM2G053008	159	GSDKTGHDEADGDVMMVLVPLFSVKDRPTKIALVPSSNAISKTMHGRGVVQ	EWEN
HvMLOC_57250	14	G---QVMGGLG-----	VRDASWAT
Os12g0157000	1	-----	MAMHAGHAWWGVA
GmLoc100778733	107	G--NNNNSSSSN-----	LQQQQQQPA
Bra004501	12	G--EEEEEEQE-----	EAKKPMEEV
Bra000434	11	G--EEEEEE--A-----	GAKKPTEEV
Bra040478	11	G--EA-----	QVKKPIEEV
Bra014415	18	N--KKMNLLEER-----	PVQE-ASSM
Bra003482	1	----MNQEEEN-----	PVEK-ASSM
Bra007646	1	----MNQENKK-----	PLEEASTSM
GlycinemaxLoc100781489	65	G--NLQQ-GGGG-----	VVHHAHQV
GRMZM2G024948_T01	78	G--ASGSGGP-A-----	DADAEAEAV
os02g0683500	73	G--ASGSGGGG-----	G--GGEAEVV
HvMLOC_66387	70	G--ASGSGAGDG-----	SRSNVAAV
os04g0581400	88	G--ASGSGGGG-----	G--EDVEVI
GRMZM2G102059_T01	73	G--ASGSGG-GG-----	S--DDVQVI
Os10g0537100	19	----S---E-----	RGAAAWAV
GRMZM2G142999_T01	16	----S---E-----	RPRGGVAW
GRMZM2G125095_T01	14	----S---E-----	RPRGGVAW
os03g0120900	23	-----	SGGRSLAAV
GRMZM2G098443_T01	20	A--AEHN---Q-----	HHQQQHATV
GRMZM2G082227_T01	20	A--AEHH---Q-----	QQQQATV
Os11g0156000	23	-----YANFH-----	YH---HHY
GRMZM2G328742_T01	24	-----YTNLH-----	YHHQQHHHY
GmLoc100802734	49	-----IVSDK-----	ATNNLTQEE
GmLoc100795470	50	DNDENQ---TTT-----	TGGEQEILD
GmLoc100818164	49	DNDENQTVTTTT-----	TGGEETIINN
Bra017262	25	-----	HQNEVVE
At2g36080	26	-----	HQNDVVE
Bra005301	27	-----	HQNDVAE
At3g11580	17	-----	QNDVAIAQ
BraLOC103849927	20	-----	HQNYAVV
BrassicarapaBra034828	17	-----	NEH
At5g06250	26	----DTSE-TT-----	TTATWLHDDI
Bra005886	28	----DTSEPTT-----	TAATSLREDO
GmLoc102660503	168	-----HIG-----	LKGDDEEQFCCTQ
HvMLOC_38822	32	-----PLA-----	SPAPSAAAAP
os01g0693400	195	----SNGHLSS-----	PRSPFAPAAA
HvMLOC44012	31	-----GDAGG-----	SASPPSPAAY
HvMLOC_7940	163	-----	AQPTPSWA
HvMLOC_75135	162	-----	AQPTPSWA
TRAECM81004	158	-----	AQPTPSWA
HvMLOC_56567	157	-----	AQMPPSWA
TRAES3BF098300010CFD_t1	157	-----	AQPTPSWA
HvMLOC_63261	146	-----	AQPTPAWA
TRAES3BF062700040CFD_t1	159	-----	AQPTPAWA
TRAES3BF062600010CFD_t1	159	-----	AQPTPAWA
Bra038346	173	-----TN-----	TTQSHTVSRT
GmLoc732601	153	-----LDADT-----	ASSGVFDAKA
GmLoc100789009	159	-----RDAET-----	ASSGAFDAKA
GmLoc100776987	185	-----GAGNGN-----	SISGACVMKA
GmLoc100801107	191	-----AGTAN-----	SISGACFTKA

GRMZM2G053008	215	-----VGEELALP-FNTGVPEKINWEDHIRKNSVEWGQMA--VCKLFDERPVWPROSL
HvMLOC_57250	30	--FVVVEQSDVQVQGNRLTKEAVWGGPIPKLFELEELR-----GDGLNAENL
Os12g0157000	20	IHHYRHKTSDVGK-NR--K-----HA--YGGGD-----SGK
GmLoc100778733	132	---KVVTPSDVGK-LNRLVIPKQH-----AEKYFPLDS--S-----ANEKGL
Bra004501	37	---KVVTPSDVGK-LNRLVIPKQY-----AE--YFPLDS--S-----TNEKGL
Bra000434	34	---KVVTPSDVGK-LNRLVIPKQH-----AE--YFPLDS--S-----TNEKGL
Bra040478	29	---KVVTPSDVGK-LNRLVIPKQH-----AE--YFPLDS--S-----SNEKGL
Bra014415	42	---KVVTPSDVGK-LNRLVIPKQH-----AE--YFPLDNNSS-----DNNKGL
Bra003482	22	---KVVTPSDVGK-LNRLVIPKQH-----AE--YFPLDNN-S-----DSSKGL
Bra007646	23	---KVVTPSDVGK-LNRLVIPKQH-----AE--YFPLDNS-S-----TNNKGL
GlycinemaxLoc100781489	89	---EKVATPSDVGK-LNRLVIPKQH-----AEKYFPLDSST-----NEKGL
GRMZM2G024948_T01	102	---KVVTPSDVGK-LNRLVIPKQY-----AEKYFPLDAAA-----NEKGL
os02g0683500	97	---KVVTPSDVGK-LNRLVIPKQY-----AEKYFPLDAAA-----NEKGL
HvMLOC_66387	95	---KVVTPSDVGK-LNRLVIPKQY-----AEKYFPLDSAA-----NEKGL
os04g0581400	111	---KVVTPSDVGK-LNRLVIPKQH-----AEKYFPLDSAA-----NEKGL
GRMZM2G102059_T01	95	---KVVTPSDVGK-LNRLVIPKQH-----AEKYFPLDAAA-----NEKGL
Os10g0537100	36	---EKVTPSDVGK-LNRLVIPKQH-----AE--YFPLDAAAGA--GGGGGGGGGGGGKGL
GRMZM2G142999_T01	33	---EKVTPSDVGK-LNRLVIPKQH-----AE--YFPLDASSA--AA-AAAAAAGGGKGL
GRMZM2G125095_T01	31	---EKVTPSDVGK-LNRLVIPKQH-----AE--YFPLDASAA--AA-SASASAGGGKGL
os03g0120900	38	---KVVTPSDVGK-LNRLVIPKQH-----AEKYFPLDAAS-----NEKGL
GRMZM2G098443_T01	41	---KVVTPSDVGK-LNRLVIPKQH-----AEKYFPLDAAA-----NEKGL
GRMZM2G082227_T01	39	---KVVTPSDVGK-LNRLVIPKQH-----AE--YFPLDAAA-----NDKGL
Os11g0156000	39	---EKF--TPSDVGK-LNRLVIPKQH-----AE--YFPLGAGD-----AADKGL
GRMZM2G328742_T01	44	---EKF--TPSDVGK-LNRLVIPKQH-----AE--YFPLSSSG-----AGDKGL
GmLoc100802734	69	---EKF--TPSDVGK-LNRLVIPKQH-----AEKHFFPLDS-----SAAKGL
GmLoc100795470	74	---EKF--TPSDVGK-LNRLVIPKQH-----AEKYFPLSG-DS---G-----GSECKGL
GmLoc100818164	76	---EKF--TPSDVGK-LNRLVIPKQH-----AEKYFPLSGGDS---G-----SSECKGL
Bra017262	38	---EKF--TPSDVGK-LNRLVIPKQH-----AE--YFPLAAAA-----VDAVEKGL
At2g36080	39	---EKF--TPSDVGK-LNRLVIPKQH-----AE--YFPLAAAA-----ADAVEKGL
Bra005301	40	---EKF--TPSDVGK-LNRLVIPKQH-----AE--YFPLAAAA-----ADAMEKGL
At3g11580	30	---EKS--TPSDVGK-LNRLVIPKQH-----AEKYFPLNNNNN---NGSGDDVATTEKGL
BraLOC103849927	33	---EKS--TPSDVGK-LNRLVIPKQH-----AEKHFFPLNNAGD---D---VAAAEETTEKGL
BrassicarapaBra034828	25	---EKS--TPSDVGK-LNRLVIPKQH-----AE--YFPLNNCGG---GG---DVTAESTEKEKGL
At5g06250	47	---EKS--TPSDVGK-LNRLVIPKQH-----AEKYFPLNAVLV---SSA-AADTSSEKEKGL
Bra005886	50	---EKS--TPSDVGK-LNRLVIPKQH-----AEKYFPLNTIIS---N-----NAEEKGL
GmLoc102660503	186	---CKE--TPSDVGK-LNRLVIPKQH-----AVSYFFHYVGGSD-----ESGSVDKGL
HvMLOC_38822	50	---KTVTPSDVGK-LNRLVIPKQH-----AEKHFFPLQLPSAS-----AAVPGECKGL
os01g0693400	217	---KTVTPSDVGK-LNRLVIPKQH-----AEKHFFPLQLPSA-----GGESKGL
HvMLOC44012	51	---KTVTPSDVGK-LNRLVIPKQN-----AEKHFFPLQLPAG-----GGESKGL
HvMLOC_7940	176	---EKAVTPSDVGK-LNRLVIPKQH-----AEKHFFPLKRTPE-----TTTTTGKGL
HvMLOC_75135	175	---EKAVTPSDVGK-LNRLVIPKQH-----AEKHFFPLKCTAE-----TTTTTGNKGL
TRAECDM81004	171	---EKAVTPSDVGK-LNRLVIPKQH-----AEKHFFPLKRTPE-----RTTTTGNKGL
HvMLOC_56567	170	---EKAVTPSDVGK-LNRLVIPKQH-----AEKHFFPLKRSPE-----TTTTTGNKGL
TRAES3BF098300010CFD_t1	170	---EKAVTPSDVGK-LNRLVIPKQH-----AEKHFFPLKRTPE-----TPTTTGKGL
HvMLOC_63261	159	---EKAVTPSDVGK-LNRLVIPKQH-----AEKHFFPLPTTA-A-----AAGSDGKGL
TRAES3BF062700040CFD_t1	172	---EKAVTPSDVGK-LNRLVIPKQH-----AEKHFFPLPTTA-A-----ATGSNGKGL
TRAES3BF062600010CFD_t1	172	---EKAVTPSDVGK-LNRLVIPKQH-----AEKHFFPLPTTA-A-----ATGSNGKGL
Bra038346	190	---EKVTPSDVGK-LNRLVIPKQH-----AEKYFPLPS-----LSVTKGL
GmLoc732601	173	---EKTVTPSDVGK-LNRLVIPKQH-----AEKHFFPLSGSGDESSPCV---AGASAACKGL
GmLoc100789009	179	---EKTVTPSDVGK-LNRLVIPKQH-----AEKHFFPLSGSGGGALPCM---AAAAGACKGL
GmLoc100776987	206	---EKAVTPSDVGK-LNRLVIPKQH-----AEKHFFPLQSAAN-----GVSATATAACKGL
GmLoc100801107	211	---EKAVTPSDVGK-LNRLVIPKQH-----AEKHFFPLQSSNGVSATTIAAVTATPTAAKGL

GRMZM2G053008	267	VERLLDNVHVSQNFRLLLFRAGYFSSGP---	GKFWIERGYPKDS-----
HvMLOC_57250	79	KILDA-----DCEGDANRLNSSKAYRW	GPQWSRLVKETCKGLRDLYAAT
Os12g0157000	49	-----S-DSGKWR-RYSYWTSS--	YTKGWSRFVK--RDAGLVHRVGG
GmLoc100778733	169	ILNFEDR-----NGHLWRFRYSYWNSSQSY	VTKGWSRFVKEKLDAGLVSEQRGV
Bra004501	74	ILNFEDL-----AGHSWRFRYSYWNSSQSY	VTKGWSRFVKKLDAGLVSEQRGV
Bra000434	71	ILNFEDL-----TGHSWRFRYSYWNSSQSY	VTKGWSRFVKKLDAGLVSELRGV
Bra040478	66	ILNFEDL-----TGHSWRFRYSYWNSSQSY	VTKGWSRFVKKLDAGLVSEQRGV
Bra014415	81	ILNFEDR-----IGILWSFRYSYWNSSQSY	VTKGWSRFVKKLDAGLVSEHRRGS
Bra003482	60	ILNFEDR-----TGNWRFRYSYWNSSQSY	VTKGWSRFVKKLDAGLVSEQRDP
Bra007646	61	ILNFEDR-----TGNWRFRYSYWNSSQSY	VTKGWSRFVKKLDAGLVSEQRDP
GlycinemaxLoc100781489	126	ILNFEDR-----NGHWWRFRYSYWNSSQSY	VTKGWSRFVKEKLDAGLVSEQRGV
GRMZM2G024948_T01	139	ILSFEDS-----AGHWRFRYSYWNSSQSY	VTKGWSRFVKEKLDAGLVSESRGA
os02g0683500	134	ILNFEDR-----AGHPWRFRYSYWNSSQSY	VTKGWSRFVKEKLDAGLVSESRGI
HvMLOC_66387	132	ILNFEDS-----AGHPWRFRYSYWNSSQSY	VTKGWSRFVKEKLDAGLVSESRGA
os04g0581400	148	ILSFEDR-----TGHLWRFRYSYWNSSQSY	VTKGWSRFVKEKLDAGLVSECRGA
GRMZM2G102059_T01	132	ILSFEDR-----AGHLWRFRYSYWNSSQSY	VTKGWSRFVKEKLDAGLVSECRGA
Os10g0537100	86	ILSFEDR-----TGHWWRFRYSYWNSSQSY	VTKGWSRFVKEKLDAGLVSEGRGL
GRMZM2G142999_T01	82	ILSFEDR-----AGHAWRFRYSYWNSSQSY	VTKGWSRFVKEKLDAGLVSEHARGA
GRMZM2G125095_T01	80	ILSFEDR-----AGHAWRFRYSYWNSSQSY	VTKGWSRFVKEKLDAGLVSEHARGA
os03g0120900	75	ILSFEDR-----TGHPWRFRYSYWNSSQSY	VTKGWSRFVKEKLDAGLVSEGRGV
GRMZM2G098443_T01	78	ILSFEDR-----TGHPWRFRYSYWNSSQSY	VTKGWSRFVKEKLDAGLVSEGRGI
GRMZM2G082227_T01	76	ILSFEDR-----AGHPWRFRYSYWNSSQSY	VTKGWSRFVKEKLDAGLVSEGRGV
Os11g0156000	77	ILSFEDS-----AGAPWRFRYSYWTSSQSY	VLTGWSRFVKEKLDAGLVSEHRRV
GRMZM2G328742_T01	82	ILCFEDDDDDDEAAAANPEWRFRYSYWTSSQSY	VLTGWSRFVKEKLDAGLVSEHRRV
GmLoc100802734	105	ILSFEDS-----SGHCWRFRYSYWNSSQSY	VLTGWSRFVKEKLDAGLVSEHRRV
GmLoc100795470	114	ILSFEDS-----SGHCWRFRYSYWNSSQSY	VLTGWSRFVKEKLDAGLVSEHRRV
GmLoc100818164	117	ILSFEDS-----SGHCWRFRYSYWNSSQSY	VLTGWSRFVKEKLDAGLVSEHRRV
Bra017262	78	ILCFEDS-----EGHPWRFRYSYWNSSQSY	VLTGWSRFVKEKLDAGLVSEHRRV
At2g36080	79	ILCFEDS-----EGHPWRFRYSYWNSSQSY	VLTGWSRFVKEKLDAGLVSEHRRV
Bra005301	80	ILCFEDS-----EGHPWRFRYSYWNSSQSY	VLTGWSRFVKEKLDAGLVSEHRRV
At3g11580	78	ILSFEDS-----SGHCWRFRYSYWNSSQSY	VLTGWSRFVKEKLDAGLVSEHRRV
BraLOC103849927	78	ILTFEDS-----SGHCWRFRYSYWNSSQSY	VLTGWSRFVKEKLDAGLVSEHRRV
BrassicarapaBra034828	71	ILSFEDS-----SGHSWRFRYSYWNSSQSY	VLTGWSRFVKEKLDAGLVSEHRRV
At5g06250	94	ILSFEDS-----SGHSWRFRYSYWNSSQSY	VLTGWSRFVKEKLDAGLVSEHRRV
Bra005886	91	ILSFEDS-----SGHCWRFRYSYWNSSQSY	VLTGWSRFVKEKLDAGLVSEHRRV
GmLoc102660503	227	SAVETDK-----LNLWWRFRYSYWNSSQSY	VLTGWSRFVKEKLDAGLVSEHRRV
HvMLOC_38822	94	ILNFEDA-----TGHWWRFRYSYWNSSQSY	VLTGWSRFVKEKLDAGLVSEHRRV
os01g0693400	257	ILNFEDA-----AGHWWRFRYSYWNSSQSY	VLTGWSRFVKEKLDAGLVSEHRRV
HvMLOC44012	91	ILNFEDD-----AGHWWRFRYSYWNSSQSY	VLTGWSRFVKEKLDAGLVSEHRRV
HvMLOC_7940	218	ILNFEDG-----EGHWWRFRYSYWNSSQSY	VLTGWSRFVKEKLDAGLVSEHRRV
HvMLOC_75135	217	ILNFEDG-----EGHWWRFRYSYWNSSQSY	VLTGWSRFVKEKLDAGLVSEHRRV
TRAECMD81004	213	ILNFEDG-----EGHWWRFRYSYWNSSQSY	VLTGWSRFVKEKLDAGLVSEHRRV
HvMLOC_56567	212	ILNFEDG-----QGHVWRFRYSYWNSSQSY	VLTGWSRFVKEKLDAGLVSEHRRV
TRAES3BF098300010CFD_t1	212	ILNFEDG-----EGHWWRFRYSYWNSSQSY	VLTGWSRFVKEKLDAGLVSEHRRV
HvMLOC_63261	200	ILNFEDG-----QGHVWRFRYSYWNSSQSY	VLTGWSRFVKEKLDAGLVSEHRRV
TRAES3BF062700040CFD_t1	213	ILNFEDG-----EGHWWRFRYSYWNSSQSY	VLTGWSRFVKEKLDAGLVSEHRRV
TRAES3BF062600010CFD_t1	213	ILNFEDG-----EGHWWRFRYSYWNSSQSY	VLTGWSRFVKEKLDAGLVSEHRRV
Bra038346	227	ILNFEDV-----TGHWWRFRYSYWNSSQSY	VLTGWSRFVKEKLDAGLVSEHRRV
GmLoc732601	221	ILNFEDV-----GKHVWRFRYSYWNSSQSY	VLTGWSRFVKEKLDAGLVSEHRRV
GmLoc100789009	227	ILNFEDV-----GKHVWRFRYSYWNSSQSY	VLTGWSRFVKEKLDAGLVSEHRRV
GmLoc100776987	251	ILNFEDV-----GKHVWRFRYSYWNSSQSY	VLTGWSRFVKEKLDAGLVSEHRRV
GmLoc100801107	263	ILNFEDV-----GKHVWRFRYSYWNSSQSY	VLTGWSRFVKEKLDAGLVSEHRRV

GRMZM2G053008	314	-----ESQYQFIDMPPE	RYLLRLKN-----
HvMLOC_57250	132	ATA--A-----SSCSGARA	VAP-AIPPGAI-----VKAAGF--
Os12g0157000	90	AA-----DRGC	RRGSAAVRVTA-----
GmLoc100778733	221	GE-LY-RHRL	FIDWRRRPDHHHHHGGPDHSTTLFTPFLLIPNQPHHLSIRWGAT-G-
Bra004501	126	GD-SGRDSRL	FIDWRRRPKPDHPTSLAHFA-----AGSMF-P-
Bra000434	123	GD-TGRDSRL	FIDWRRRPKPDYTTSTSHFP-----AGAMF-P-
Bra040478	118	GD-----SRL	FIDWRRRPKPDYPTSTAHFA-----AGAMF-P-
Bra014415	133	---CNKDRL	FIDWRRRPKPDH-----QV-----VGAMF-P-
Bra003482	112	---GNKDRL	FIDWRRRPKPDHH-----HQF-----AGAMF-P-
Bra007646	113	---CNKDRL	FIDWRRRPKPDH-----HQF-----AGAMF-P-
GlycinemaxLoc100781489	178	GD-LYRHRL	FIDWRRRPDAHAHP-----PHHHDPFLPLPSI--
GRMZM2G024948_T01	191	AE--DARHRL	FIDWRRRPVDRGPLRFS-----GLALPMLP-S-
os02g0683500	186	GD-EAARHRL	FIDWRRRADRDPLRLPR-----GLPLPMLT--
HvMLOC_66387	184	GE-AARHRL	FIDWRRRADRDPLRLPR-----LPLPMLT--
os04g0581400	200	AE--ATRDRRL	FIDWRRRADRDPHRFQR-----LPLPM--T-
GRMZM2G102059_T01	184	GD-TARDRL	FIDWRRRADRDPHRMPR-----LPLPMAFV-A-
Os10g0537100	138	GD-AARGRL	FIDWRRRQAGSFMFPPTAA----PPSHSHHHQRRHHPPLP--S-
GRMZM2G142999_T01	134	GG--ARGRF	FIDWRRRLAF-LQPTLA-----SAQRLPLP--S-
GRMZM2G125095_T01	132	GA--TRGRF	FIDWRRRLAF-LQPPLA-----SAQRLPLP--S-
os03g0120900	127	GE-AARGRL	FIDWRRRPDAALQ-----PPTHFAHHLPS--S-
GRMZM2G098443_T01	130	GE--AARDRL	FIDWRRRPDPVVH-----QYH-HRLPLPSA--V-
GRMZM2G082227_T01	128	GE--AARGRL	FIDWRRRPDPVVH-----QYHHHRLPLPSA--V-
Os11g0156000	129	GS-FGVGDRL	FIGWRRRGDAAAQTPAPPPAV-----RV--AP--
GRMZM2G328742_T01	141	G--FGMPDRL	FIGWRRRGSTATAATTVPAAAA-----VRVVAPAQSAGA
GmLoc100802734	157	S----LPQRF	FIGWRRRQF--FVPAHVSTR-----
GmLoc100795470	166	F----DAQRL	FIGWRRRQDAALPPAHVSSRK-----SGGGDGN
GmLoc100818164	169	A----DAQRL	FIGWRRRQDALPPAHVSSRK-----SGGD-GN
Bra017262	130	A----DGRF	FIGWRRRGDSSSSDSYRNQ-----
At2g36080	131	S----DGRF	FIGWRRRGDSSSSDSYRHVQ-----
Bra005301	132	F----DGRF	FIGWRRRGNSSSSDSYRHQ-----
At3g11580	130	F----DLHRL	FIGWRRRGESSSPAVSVVSQEA-----L-----
BraLOC103849927	130	F----DLHRL	FIGWRRRGESSSPTAVSVVSQEA-----R-----
BrassicarapaBra034828	123	F----DIHRL	FIGWRRRGESSSSAVSAVTQDP-----R-----
At5g06250	146	S----DSRRL	FIGWRRRGSSSSVAATNSAVN-----T-----
Bra005886	143	S----DSRRL	FIGWRRRGSSSAANTTSY-----S-----
GmLoc102660503	279	K--SGGEGEAFALIDVIYNNN	EEDS---KGDTKQVL-----GNQLQLA----
HvMLOC_38822	146	SG---NNQL	FIDCLRSKTTTTT---SV-----NSEA-----
os01g0693400	309	A-SAGDDGRL	FIDCLVRS--GA-A-----L-----ASP-----
HvMLOC44012	143	AGRTGEDSRL	FIDCLRPNTAA-----E-----ADPV-----
HvMLOC_7940	270	YGQ---EKQF	FIDCNKTTSCPADDRGAATA-----SPPVS-----
HvMLOC_75135	269	YGQ---EKQL	FIDCNTTNG-----GKTAL-----PLPVV-----
TRAECDM81004	265	YGQ---EKQL	FIDCNTTNS-----GKSAS-----PLPVV-----
HvMLOC_56567	264	YGQ---EKQF	FIDCNTTNG-----GKSAS-----PLQVM-----
TRAES3BF098300010CFD_t1	264	YEQ---EKQF	FIDCNTSNG-----GKSAS-----PLPVG-----
HvMLOC_63261	252	VMNDTDEQL	FIDQSSKDEAAD-----VAT-----
TRAES3BF062700040CFD_t1	265	YG-NDTEDQL	FIDMKNKDDAAD-----AAI-----
TRAES3BF062600010CFD_t1	265	YG-NDTEDQL	FIDMKNKDDAAD-----AAI-----
Bra038346	279	---GSDRQL	IDWIRSGSKN-----
GmLoc732601	273	---GPDRL	IDCARSGVNNNA---G--GL-----FVPI-----
GmLoc100789009	279	---GLDRQL	IDCARSGVNNNA---A--GL-----FIPV-----
GmLoc100776987	303	---GPDRL	IDWIRNVVNE---V--AL-----F-----
GmLoc100801107	315	---GPDRL	IDWIRNVVNE---V--AL-----FGPV-----

GRMZM2G053008	338	-----SESRKWADMCKLETMPQSFIYLQLYELKDDFIQAEIRKPSYQSVCSRSTGWFS
HvMLOC_57250		-----
Os12g0157000	109	-----NGGWSMCMYSTSG---SSYDT--S-----
GmLoc100778733	275	RLYSLPSPPTPPRHHEH-----LNYNNA-----MYH-----
Bra004501	162	RFYSFPTAT---SYNL-----YNYQQP-----
Bra000434	159	RFYSFQTATTSTSYNP-----YNHQQP-----
Bra040478	150	RFYSFPTATTSTCYDL-----YNHQQP-----
Bra014415	160	RFYSYPYPQIQASYER-----
Bra003482	141	RFYSFSHPQN-----
Bra007646	141	RFYSFPHPMPTSFES-----
GlycinemaxLoc100781489	213	RLYSLPPTMPPRYHHDHFFH---HHLNYYNLF-----T-----
GRMZM2G024948_T01	226	SHYGGPHHYSPPWFGGGGG-----GGGGFFM
os02g0683500	222	-----SHYAPWGI GGG-----GGFFV
HvMLOC_66387	218	-----SHYSPWGLGAG-----ARGFFM
os04g0581400	232	-----SPYGPWGGGA-GA-----SSCRPRR
GRMZM2G102059_T01	219	-----SPYGPWGGGG-GG-----GAGGFFM
Os10g0537100	185	-----VPLCPWRDYTTAYG---GGYGY-----
GRMZM2G142999_T01	170	-----VPICPWQDYG-----
GRMZM2G125095_T01	168	-----VPICPWQGYG-----
os03g0120900	163	-----IPFAPWAHHH---G---H-----G-----AAAAA-AAAAGARFLL
GRMZM2G098443_T01	165	-----VPYAPWAA---HAHH---HHYPADGHT-----EPVTPCLCATLVATEM
GRMZM2G082227_T01	164	-----VPYAPWAAAAHAHH---HHYPAAGVG-----AARTTTTTTTTTVLHHL
Os11g0156000	164	---AAQNAGEQQPWSPPMCYSTS---GGGSY-----
GRMZM2G328742_T01	186	---DHQQQQQPSPWSPPMCYSTS---GSYSY-----
GmLoc100802734	183	-----SSASFYSAH---PP--Y-----
GmLoc100795470	202	SNK-----NEGWTGRGFYSAH---HP--Y-----
GmLoc100818164	204	SSKNEGDVGVGWTRGFYPAH---HP--Y-----
Bra017262	157	-----
At2g36080	158	-----
Bra005301	159	-----
At3g11580	160	----VNTTAYWSGLT-----TP--Y-----
BraLOC103849927	160	----VNTTAYWSGLT-----TP--Y-----
BrassicarapaBra034828	153	----ANTTAYWNGLT-----TP--Y-----
At5g06250	176	----SS-----MGA-----LS--Y-----
Bra005886	170	----SS-----MT-----
GmLoc102660503	318	-----
HvMLOC_38822	173	-----
os01g0693400	336	-----
HvMLOC44012	172	-----
HvMLOC_7940	305	-----
HvMLOC_75135	298	-----
TRAECDM81004	294	-----
HvMLOC_56567	293	-----
TRAES3BF098300010CFD_t1	293	-----
HvMLOC_63261	280	-----
TRAES3BF062700040CFD_t1	292	-----
TRAES3BF062600010CFD_t1	292	-----
Bra038346	298	-----
GmLoc732601	301	-----
GmLoc100789009	307	-----
GmLoc100776987	325	-----
GmLoc100801107	341	-----

GRMZM2G053008	392	KPMIKTL-RLQVSIR-----	LLSLLHNE-----
HvMLOC_57250		-----	-----
Os12g0157000	127	-----	ANSYAYHRSV-----
GmLoc100778733	301	-----P-FHHHGAGSGINATTHHYNHYHEMSSTTT--	SGSAGSVFYHRS-TPPISMPLA
Bra004501	181	-----RHHHHS	YNYPQIPRE-----FGYGYLV-----
Bra000434	181	-----RH-HHSG-----	YCYPQIPRE-----FGYGYVVRV-----
Bra040478	172	-----RH-HHIG-----	YGYEQIPRE-----FGYGYFVRV-----
Bra014415	176	-----H-----	NLYHRYQRD-----IGIGYVVRV-----
Bra003482	151	-----	LYHRYQQD-----LGIGYVVRV-----
Bra007646	157	-----SHN-----	LYHHRFQRD-----LGIGYV-----
GlycinemaxLoc100781489	243	-----FQQHQYQQIGAATTHHNNYGY-----	QNSGSGSLYLRSSMSMGG-----
GRMZM2G024948_T01	252	PPSPPATLYEH-RLR--Q----	GLDFRSMTTTTYPAPTQVGRQLLFSGSARMPPHHAPPP
os02g0683500	238	QPSPPATLYEH-RLR--Q----	GLDFRAFNPAA--AMGRQVLLFGSAR-IPQAP--
HvMLOC_66387	235	PPSPPATLYEH-RLR--Q----	GDFRGMNPSYP--TMGRQVILFGSARMPPHGPAP
os04g0581400	251	PPRSTSI----TAF--R----	AST--SATS-----TPLCRGSSS-----SSAPQ
GRMZM2G102059_T01	238	PPAPPATLYEHHRFR--Q----	ALDFRNINAAA--APARQLLFSGSAGMPPRASMPQ
Os10g0537100	204	-----	GYG--GGSTPASSRHVLF-----
GRMZM2G142999_T01	180	-----	ASAPAPNRHVLF-----
GRMZM2G125095_T01	178	-----	ASAPAPSRHVLF-----
os03g0120900	191	PPSST-PIYDHHRRH-----	AHAVGYDAYA----AATSRQVLFY-----
GRMZM2G098443_T01	202	RASSS-QLSLTRSNTLS--RPPQPRIARVDGAQPRPSSSPRQPQSLWC-----	
GRMZM2G082227_T01	203	PPSPS-PLYLDTRRR-----	HVGYDAY-----GAGTRQLLFY-----
Os11g0156000	188	-----PT-----	SPANSY-----AY-----
GRMZM2G328742_T01	210	-----PT-----	SSPANSQH-----AYH-----
GmLoc100802734	195	-----PA-----	HH-----F-----
GmLoc100795470	220	-----PT-----	HH-----LHH-----
GmLoc100818164	227	-----PT-----	HH-----
Bra017262	157	-----	
At2g36080	158	-----	
Bra005301	159	-----	
At3g11580	174	-----RQ-----	VH-----AST-----
BraLOC103849927	174	-----RQ-----	VH-----AST-----
BrassicarapaBra034828	167	-----RQ-----	VH-----AST-----
At5g06250	184	-----HQ-----	IH-----ATS-----
Bra005886	174	-----	AP-----PYS-----
GmLoc102660503	318	-----	
HvMLOC_38822	173	-----	
os01g0693400	336	-----	
HvMLOC44012	172	-----	
HvMLOC_7940	305	-----	
HvMLOC_75135	298	-----	
TRAECMD81004	294	-----	
HvMLOC_56567	293	-----	
TRAES3BF098300010CFD_t1	293	-----	
HvMLOC_63261	280	-----	
TRAES3BF062700040CFD_t1	292	-----	
TRAES3BF062600010CFD_t1	292	-----	
Bra038346	298	-----	
GmLoc732601	301	-----	
GmLoc100789009	307	-----	
GmLoc100776987	325	-----	
GmLoc100801107	341	-----	

GRMZM2G053008	414	-----EAKNLLRNAHELIER--SKKQEALSRSELSIEYNDA--DQVSAHTGT-----
HvMLOC_57250		-----
Os12g0157000	137	-----DDHSDHAGSRA-----
GmLoc100778733	351	DHQTlnTRQ-----QQQQQQQEGAGNVSLSPMIIDSVPVAAHHLHHQQHGGKSSG---
Bra004501	204	-----DQRAVVADPLVIESVPVMMHGG-A-----
Bra000434	206	-----DQRAVVADPLVIESVPVMMHGG-A-----
Bra040478	197	-----DQRAVVADPLVIESVPVMMRGG-A-----
Bra014415	196	-----ERYD--PTAVIESVPVIMQRR-A-----
Bra003482	169	-----ERND--PTAVIESVPLIMQRRAA-----
Bra007646	175	-----PTAVIESVPVIMQRRAA-----
GlycinemaxLoc100781489	284	-----GDQNLQGRGSNIVPMIIDSVPVNVAHHNNNRHNGG-----
GRMZM2G024948_T01	303	QP-----RPFSLPLHHYTVQP-SAAGVTAASRPVLLDSVPVIESP-----
os02g0683500	283	-L---LARAPSLHHHTLQP-SGDGVRAAGSPVVLDSVPVIESP-----
HvMLOC_66387	284	LL--VPRPPPLHFTVQQQSGSDAGGSVTAGSPVVLDSVPVIESP-----
os04g0581400	285	GRGFISTRPCHRRRRLRLT--NSTLRCTTRAP-----
GRMZM2G102059_T01	287	QQ--QPPPPPHPLHSIMLVQ-PSPAPPTASVPMMLDSVPLVNSP-----
Os10g0537100	221	-----RPQV-----PAAVVLKSVPVHVAATSAVQ-----
GRMZM2G142999_T01	193	-----RPQV-----PAAVVLKSVPVHVAASAV-----
GRMZM2G125095_T01	191	-----RPQV-----PAAVVLTSVPVRVAASAV-----
os03g0120900	225	-----RPLPPQQ-----QHHPAVVLESVPVRMTAGH--A--EP-----
GRMZM2G098443_T01	246	-----RSC-----QPQPRRTA--DV-----
GRMZM2G082227_T01	234	-----RPH--Q-----QPSTTVMLDSVPVRLPPTPGQHA--EP-----
Os11g0156000	198	-----RRAADHDH-----GDMHHADES---PRD
GRMZM2G328742_T01	223	-----RHSADHDH-----SNNMQHAGES---QSD
GmLoc100802734	200	-----FFPYQ-----PHSLHAPGGGSQGGNE
GmLoc100795470	227	-----HQFSPYQ-----QQHDC LHAGRGSGGQGNQ
GmLoc100818164	231	-----HHFSPYHH-----QQDDSLHAVRGSGGQGNQ
Bra017262	157	-----SNSSL-----QY--YPHAG--AQA---
At2g36080	158	-----SNASL-----QY--YPHAG--AQA---
Bra005301	159	-----SNASL-----QY--YPHAG--VQA---
At3g11580	181	-----TYPN-IH-----QE--YSHYG--AVVDHA
BraLOC103849927	181	-----SSYPN-IH-----QE--YSHYG--AVA---
BrassicarapaBra034828	174	-----SSYPNNIH-----QE--YSHYG--PVA---
At5g06250	191	-----NYSNPPSH-----SE--YSHYG--AAVATA
Bra005886	179	-----NYSNRPAH-----SE--YSHYG--AAVATA
GmLoc102660503	318	-----GSEE-----GED-----
HvMLOC_38822	173	-----
os01g0693400	336	-----
HvMLOC44012	172	-----
HvMLOC_7940	305	-----
HvMLOC_75135	298	-----
TRAECDM81004	294	-----
HvMLOC_56567	293	-----
TRAES3BF098300010CFD_t1	293	-----
HvMLOC_63261	280	-----
TRAES3BF062700040CFD_t1	292	-----
TRAES3BF062600010CFD_t1	292	-----
Bra038346	298	-----
GmLoc732601	301	-----
GmLoc100789009	307	-----
GmLoc100776987	325	-----
GmLoc100801107	341	-----

GRMZM2G053008	458	-----EDQVGPNNSDSDVDDEE-----EEEELEGY----
HvMLOC_57250		
Os12g0157000	148	-----DAKSSSAASSRRCVNCGADAT-----AMYGVMHHS
GmLoc100778733	402	----PSSTSTSPSTAGRLFGVNCASSTSEDPKCF----SLLSS-----
Bra004501	227	----QVSQAVVGTAGRLFGVD-----EESSSSGGSLPR-
Bra000434	229	----RVNQAAVGTAGRLFGVD-CGSSGG----T----NSTEESSSSGGSLPR-
Bra040478	220	----RVSQEVVGTAGRLFGVD-----EESSSSGGSLPRA
Bra014415	216	----HVATMASSRGERLFGVNCVGGRGGGGSV----NSTEEESSTSGGSISRG
Bra003482	190	----HVAaipssRGERLFGVD-CG--GGG-GSV----NSTEESSSSGG--GG
Bra007646	192	----QVANMASSRGERLFGVD-CG--GGGGGSV----NSTEESSSSGGSMSRG
GlycinemaxLoc100781489	320	-----ITSGGTNCSERLFGVNCASAEDESKELS---SGSAAHVTTAASSSSLH
GRMZM2G024948_T01	342	-----TTAANVRLFGVNNNDGG-----GEASHQGDALSLQM--P
os02g0683500	323	-----TTAANVRLFGVNNPAGGGGAAA---GESSNHGNALSLQ--TP
HvMLOC_66387	326	-----TTATKVVRLFGVNNPHPGDGG-----GESSNYGSALPLQMPAS
os04g0581400		
GRMZM2G102059_T01	329	-----TAASVVRLFGVNNPFGTS-----AESSQDANAL--SLRTP
Os10g0537100	245	-----EAATTTTRPVRLFGVNCPLAMDDDDIA---GA-----
GRMZM2G142999_T01	215	-----E-ATMSVVRLFGVNCPLDAEDSATV---P-----
GRMZM2G125095_T01	213	-----EEATRSVVRLFGVNCPLDAEDGATA---T-----
os03g0120900	254	-----PSAPSVVRLFGVNCPLSEQDHAGVV---GK-----
GRMZM2G098443_T01	259	-----P-----
GRMZM2G082227_T01	263	-----PPPAVASSASVVRLFGVNCPLAAGSEENV---GG-----
Os11g0156000	218	TDSPSFS---AGSAPSVRLFGVNCGEPEADT-----TAA---ATMYGYMHQQ
GRMZM2G328742_T01	244	RDNRSCSAASAPPPSVRLFGVNCGEPEPET-----P---TAMYGVMHQS
GmLoc100802734	221	T-TPGGN---SSSSSGSVRLFGVNCQDNHNSQNS---TPEC---SYTHLYHH--
GmLoc100795470	251	RMRPVGNNSSSSSSSVRLFGVD-CQEH-DDSGPS---TPQC---SYNSNNMLPS
GmLoc100818164	256	RTRPVGNNSSSSSSSVRLFGVNCQEH-DDSGPS---TPQC---SYNTNNILPS
Bra017262	172	-----VENQRGNSRLFGVNCQDS-DWSEPS---TPDG---FTTCPT---
At2g36080	173	-----VESQRGNSRLFGVNCQDS-DWSEPS---TPDG---SNTYTT---
Bra005301	174	-----VESQRGNSRLFGVNCQDS-DLPDPS---TPDG---STICPT---
At3g11580	200	-QSI---PPVAGSSVVRLFGVNCCHSDA-VE-----PPP---
BraLOC103849927	198	--EI---PTVVTGSSVVRLFGVNCCHSDV-VE-----TPP---
BrassicarapaBra034828	192	--ET---PTVAAGSSVVRLFGVNCCHSDV-VE-----PPP---
At5g06250	212	-AETHSTPSSSVVGSSVVRLFGVNCQDE-NDGDDS---VAVA---TTVES----
Bra005886	200	-TETHFIPSSSAVGSSVVRLFGVNCQDE-DEGDDS---VATA---AAACEP----
GmLoc102660503	325	----EDANIGKDFNAQRLFGVCT-----
HvMLOC_38822	173	-----APSPAPVTVVRLFGVDLIAAARHAHEHEDYGMAKTNKRT-----MEAS
os01g0693400	336	-----DQPAPSPVVRLFGVDLTAAPVEQM-----AGCKRA-----RDL
HvMLOC44012	172	-----DQSSAPVQVVRLFGVDLAAEQ-GMP-----GGCKRA-----RDL
HvMLOC_7940	305	-----EPTKGEQVVRLFGVDAGEGRRAAPV-----
HvMLOC_75135	298	-----ETAKGEQDVRLFGVDAGVGRVRAA--T-----G-----
TRAECDM81004	294	-----ETAKGEQVVRLFGVDAGVGRRAA--T-----A-----
HvMLOC_56567	293	-----EIAKAEQVVRLFGVDAGVGRRAA--T-----A-----
TRAES3BF098300010CFD_t1	293	-----VTTKGEQVVRLFGVDAGVGRRAATAT-----A-----
HvMLOC_63261	280	-----ADENEAGHVRLFGVDGWAAMAGSSGG-----
TRAES3BF062700040CFD_t1	292	-----SDENETGHVRLFGVDAGGAMAGSSGG-----
TRAES3BF062600010CFD_t1	292	-----SDENETGHVRLFGVDAGGAMAGSSGG-----
Bra038346	298	-----PVQVVRLFGVDENVSAKP-----SNVVDACGGK-----
GmLoc732601	301	-----GPVVEPVQ-MVRLFGVNLKLPVPGS-----DGVG-----
GmLoc100789009	307	-----GPVVEPVQ-MVRLFGVNLKLPVPGS-----DGIGVGCD-G-----
GmLoc100776987	325	-----GPVVEPIQVVRLFGVNLKLGSDSIA-NN-NNASGCCN-G-----
GmLoc100801107	341	-----GPVVEPIQVVRLFGVNLKLGSDTIVGNN-NNASGCCN-G-----

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Bra000434
Bra040478
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GRMZM2G125095_T01
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Bra005886
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HvMLOC_75135
TRAECM81004
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GmLoc732601
GmLoc100789009
GmLoc100776987
GmLoc100801107

484 -----DSP-----PMADDIHEFTLG---DSYAFGEGFSNG

181 YAA--VSTVNVSV-----
442 --S-SMANSNSQPPLQLLREDTLSSSS---ARFGD---QRGVGEPSMLFD--DPSLQ
261 --G-DASPSSS--QLRLGSSSEDDH-----FSKKGKSSLPFD--DQ---
274 --G-GASPSSS--QLRLGNSSSEDDH-----LFKKGKSSLPFN--DQ---
255 GGG-GASSSSS--QLRLGSSCEDDH-----FSKKGKSSLPFD--DQ---
267 GV--SMAGVGSPLQLRLVSSDGDQSLVARGAARVDEDHHLFT-KKGKSSLSFD--DK---
234 GV--SMASVGS--LQLRLVSSD--DESLVAMEAASVDEDHHLFT-KKGKSSLSFD--DRK---
240 GV--SMAGVGS--LQLRLVSSD--DESLVAMEGATVDEDHHLFTTKKGKSSLSFD--DI---
370 HQR-LRV-----PVPVPLEDPLSSSA--AAAARFG---DHKGASTGTSLLF--DPSLQ
378 GWQ-QRTPTLR--LELPRHG--GESSA--ASSPSSS--SSSKREARSALDLD-----
365 AWM-RRDPTLR--LELPPHHHGAESSA--ASSPSSS--SSSKRDAHSALDLD-----
368 AWR-PRDHTLR--LEFP SHGA-----E--ASSPSSS--SSSKREAHSGLDLD-----

366 GWQ-RPGP-LR--ESPQR--GAESSA--ASSPSSS--SSSKREAHSSLDLD-----
278 -AS-RTAA-SS--LQLPSP-----SSSTS--SSTAGKMKCSLDLG-----
243 -RG-RAAS-TT--LQLPSP-----SSSTS--SSTAGKDVCCLDLG-----
242 -R----TP-ST--LQLPSP-----SSSTS--SSTGGKDVRSLDLG-----
284 ----TAPP-----PLPSP-----P-SSS--SSSGKARC SINLD-----

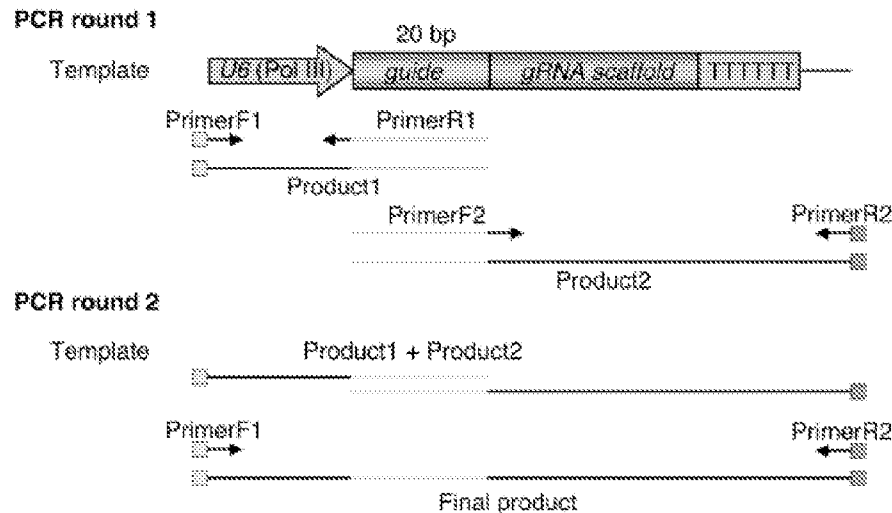
298 -WR-TSAP-----PTQQA-----S-SSS--SYSSGKARC SINLD-----
264 SSYAAMS--A-----V-----PSYWGN-----S-----
291 PYA-----YNNWGS-----PYQHDEEI-----
269 -----QTSS-----YSSSN-----PHHMH--PQQP-
303 TQGTDH-SHHNF--QQ-QP-----SNSNPS-----PHHMH--HHQPY
308 TQGTDIHSHLNF--QQQQT-----SNSKPP-----PHHMH--RHQPY
212 --NHD--QFP--P--EH-----FPP-----PYMYD--SFTGD
213 --NHD--QFHF--PQQQH-----YPP-----PYMYD--SFTGD
214 --SHD--QFH--PQ-QH-----YPP-----PYMYD--SFTGD
232 --RPDVYNDQH-----Y-----YST-----PHPMN--SFAGE
229 --CPDGYNGQH-----Y-----YST-----PDPMN--SFAGE
223 --CPDAYNGQH-----Y-----YST-----PHPMN--SFAGE
258 --PDGYYGQN-----Y-----YYS-----HPHNM--ILTLL
247 --RQDSYYDQN-----N-----YYT-----PHSSAS-----

219 V-A-APTPAHA--KKRCV-----DFALTYP--ATTPQ
373 A-A-TTPPQAAA--KKQCI-----ELALV-----
208 V-K-PP-PPKVA--KKQCI-----ELALA-----
334 -----EQE--KRQCV-----AHS--QH--SPA
327 -----ELGPPE--KRQSV-----AHGCGRM--NYI
323 -----EQGPPE--LKRQCV-----PLPHGQR--SPA
322 -----EQGPQGW--KRQCM-----AHGQH--SPA
324 -----EQGLQE--KRQCV-----APGQH--SPA

329 ----RSRDVD--ALRCS-----KKH-----
330 ----KRKEME--AFEC--K-----KKL-----
341 ----KRKEME--AFEC--K-----KKL-----
363 ----KRREME--SLECS-----KKP-----
380 ----KRREME--SLECS-----KKP-----

GRMZM2G053008	511	YLEEVLRSLPLQEDGQKKL-CDAPINADASD
HvMLOC_57250		-----
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GmLoc100778733	489	YRQ-----
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Bra000434		-----
Bra040478		-----
Bra014415		-----
Bra003482		-----
Bra007646		-----
GlycinemaxLoc100781489	417	YHRH-----
GRMZM2G024948_T01		-----
os02g0683500		-----
HvMLOC_66387		-----
os04g0581400		-----
GRMZM2G102059_T01		-----
Os10g0537100		-----
GRMZM2G142999_T01		-----
GRMZM2G125095_T01		-----
os03g0120900		-----
GRMZM2G098443_T01		-----
GRMZM2G082227_T01		-----
Os11g0156000		-----
GRMZM2G328742_T01		-----
GmLoc100802734		-----
GmLoc100795470	336	YY-----
GmLoc100818164	343	YY-----
Bra017262	237	VHQTSS-Q----QG-----
At2g36080	240	MNRTS-----
Bra005301	240	VHQTRS-P----QG-----
At3g11580	258	ALEQVGDG----RG-----
BraLOC103849927	255	AMEQVGDG----RR-----
BrassicarapaBra034828	249	AMEQVGDG----RG-----
At5g06250	283	-----
Bra005886		-----
GmLoc102660503		-----
HvMLOC_38822	248	CPRSRDQL----EGVQAAGSTFAL-----
os01g0693400		-----
HvMLOC44012		-----
HvMLOC_7940	352	-----L-GAFVL-----
HvMLOC_75135	350	CYSIG-----TI-GPLMLN-----
TRAECDM81004	346	-----L-GAFVL-----
HvMLOC_56567	343	-----L-GDFAL-----
TRAES3BF098300010CFD_t1	345	-----L-GAFAL-----
HvMLOC_63261		-----
TRAES3BF062700040CFD_t1		-----
TRAES3BF062600010CFD_t1		-----
Bra038346	345	---A-----II-NAL-----
GmLoc732601	346	---K-----VI-GAL-----
GmLoc100789009	357	---K-----VI-GAL-----
GmLoc100776987	379	---K-----II-GAL-----
GmLoc100801107	396	---K-----II-GAL-----

Figure 14



gRNA sequence (SEQ ID NO: 146)

gacggccagtccaagcttCTCGGATCCACTAGTAACGGCCGCCAGTGTGCTGGAATTGCCCTTGGATCA
 TGAACCAACGGCCTGGCTGTATTGGTGGTTGTGTAGGGAGATGGGGAGAAGAAAAGCCCGATT
 CTCCTCGCTGTGATGGGCTGGATGCATGCCGGGGAGCGGGAGGCCCCAAGTACGTGCACGGTGAG
 CGGCCCCACAGGGCGAGTGTGAGCGCGAGAGGCGGGAGGAACAGTTTAGTACCACATTGCCCAG
 CTAACCTCGAACGCGACCAACTTATAAACCCGCGCGCTGTGCTTGTGTGGGAAGGAAGAGAC
 AGATTGGTTTTAGAGCTAGAAATAGCAAGTTAAAATAAGGCTAGTCCGTTATCAACTTGAAAA
 AGTGGCACCCGAGTCGGTGTCTTTTTTGTCCCTTCGAAGGGCAATTCTGCAGATATCCATCACACT
GGCGGCCGCTCGAGGTCGaaagcttgcattgcctgcagg

os11g01560000

A-R1

GGACTGGGGTTGCTCCTGGGACACAAGCGACAGCGCGCGGG (SEQ ID NO: 147)

A-F2

CCCAGGAGCAACCCCAGTCCGTTTTAGAGCTAGAAATAGCA (SEQ ID NO: 148)

B-R1

TGCTATTTCTAGCTCTAAAACACACAAGCGACAGCGCGCGGG (SEQ ID NO: 149)

B-F2

GCCCCTGACGCCAGTGACGGTTTTAGAGCTAGAAATAGCA (SEQ ID NO: 150)

Os12g0157000

C-R1

GGGGGTGCCCTGGGCGAGAACACAAGCGACAGCGCGCGGG (SEQ ID NO: 152)

C-F2

TCTCGCCCAGGGGCACCCCCGTTTTAGAGCTAGAAATAGCA (SEQ ID NO: 153)

D-R1

CTCGTAGTGGTGGTGGTAGTACACAAGCGACAGCGCGCGGG (SEQ ID NO: 154)

D-F2

ACTACCACCACCACTACGAGGTTTTAGAGCTAGAAATAGCA (SEQ ID NO: 155)

INTERNATIONAL SEARCH REPORT

International application No

PCT/GB2016/050245

A. CLASSIFICATION OF SUBJECT MATTER

INV. A01H5/00 A01H5/10 C07K14/415 C12N15/82
ADD.

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

A01H C07K C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

EPO-Internal , BIOSIS, Sequence Search , EMBASE, MEDLINE, WPI Data

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	wo 2004/031349 A2 (MENDEL BIOTECHNOLOGY INC [US] ; JIANG CAI -ZHONG [US] ; HEARD JACQUELINE) 15 April 2004 (2004-04-15) Claims 1, 3-9 ; pages 32, 34, 41, 62, 110, 115-117, 247 ; Tables 4-6; Seq. ID Nos 395 and 396 -----	1-6, 10, 11, 14-18, 23, 24, 29-33, 35-38
X	US 2005/193443 A1 (DALE ROCK CHRISTOPHER [US] ET AL) 1 September 2005 (2005-09-01)	36-38
A	claims 1, 10, 11, 13, 22, 25, 29, 30, 36, 37 ; paragraphs [0020] , [0122] and [0140] ----- -/--	1-35



Further documents are listed in the continuation of Box C.



See patent family annex.

* Special categories of cited documents :

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier application or patent but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

"&" document member of the same patent family

Date of the actual completion of the international search

8 April 2016

Date of mailing of the international search report

26/04/2016

Name and mailing address of the ISA/

European Patent Office, P.B. 5818 Patentlaan 2
NL - 2280 HV Rijswijk
Tel. (+31-70) 340-2040,
Fax: (+31-70) 340-3016

Authorized officer

Kurz , Bi rg i t

INTERNATIONAL SEARCH REPORT

International application No.

PCT/GB20 16/050245

Box No. I Nucleotide and/or amino acid sequence(s) (Continuation of item 1.c of the first sheet)

1. With regard to any nucleotide and/or amino acid sequence disclosed in the international application, the international search was carried out on the basis of a sequence listing:
- a. ☒ forming part of the international application as filed:
- ☒ in the form of an Annex C/ST.25 text file.
- ☐ on paper or in the form of an image file.
- b. ☐ furnished together with the international application under PCT Rule 13fer1 (a) for the purposes of international search only in the form of an Annex C/ST.25 text file.
- c. ☐ furnished subsequent to the international filing date for the purposes of international search only:
- ☐ in the form of an Annex C/ST.25 text file (Rule 13fer1 (a)).
- ☐ on paper or in the form of an image file (Rule 13fer1 (b) and Administrative Instructions, Section 7 13).
2. ☐ In addition, in the case that more than one version or copy of a sequence listing has been filed or furnished, the required statements that the information in the subsequent or additional copies is identical to that forming part of the application as filed or does not go beyond the application as filed, as appropriate, were furnished.
3. Additional comments:

INTERNATIONAL SEARCH REPORT

International application No

PCT/GB2016/050245

C(Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	<p>SWAMI NATHAN K ET AL: "The plant B3 superfami ly" , TRENDS IN PLANT SCIENCE, ELSEVIER SCIENCE, OXFORD, GB, vol . 13, no. 12 , 1 December 2008 (2008-12-01) , pages 647-655 , XP025694257 , ISSN: 1360-1385 , DOI : 10.1016/J.TPLANTS.2008.09.006 [retri eved on 2008-11-26] cited in the appl icati on abstract; pages 647 , 649 , 650; Fi gures 1 and 2</p> <p>-----</p>	1-38
A	<p>M. IKEDA ET AL: "A Novel Group of Transcri pti onal Repressors in Arabi dopsi s", PLANT AND CELL PHYSIOLOGY, vol . 50, no. 5, 26 March 2009 (2009-03-26) , pages 970-975 , XP055263546, UK ISSN: 0032-0781 , DOI : 10.1093/pcp/pcp048 cited in the appl icati on abstract; pages 971-974; Tabl e 1</p> <p>-----</p>	1-38
A	<p>LI YUNHAI ET AL: "Control of final seed and organ size by the DAI gene fami ly in Arabi dopsi s thal iana" , GENES AND DEVELOPMENT, COLD SPRING HARBOR LABORATORY PRESS, PLAINVI EW, NY, US, vol . 22, no. 10, 1 May 2008 (2008-05-01) , pages 1331-1336, XP002512707 , ISSN: 0890-9369 , DOI : 10.1101/GAD.463608 cited in the appl icati on abstract; page 1332</p> <p>-----</p>	1-38
X, P	<p>YUEYING ZHANG ET AL: "Transcri pti on Factors S0D7/NGAL2 and DPA4/NGAL3 Act Redundantly to Regul ate Seed Si ze by Di rectly Repressi ng KLU Expressi on in Arabi dopsi s thal iana" , THE PLANT CELL, vol . 27, no. 3, 1 March 2015 (2015-03-01) , pages 620-632 , XP055263586, US ISSN: 1040-4651 , DOI : 10.1105/tpc.114.135368 abstract; pages 621-624, 628, 629 ; Fi gures 2, 3 and 5</p> <p>-----</p>	1-38

INTERNATIONAL SEARCH REPORT

Information on patent family members

International application No

PCT/GB2016/050245

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO 2004031349 A2	15-04-2004	AT 539158 T AU 2003285856 A1 BR 0314389 A DK 1546336 T3 EP 1546336 A2 EP 2270166 A2 EP 2270167 A2 EP 2272962 A2 ES 2380017 T3 PT 1546336 E US 2007033671 A1 US 2007101454 A1 WO 2004031349 A2	15-01-2012 23-04-2004 12-07-2005 10-04-2012 29-06-2005 05-01-2011 05-01-2011 12-01-2011 07-05-2012 09-04-2012 08-02-2007 03-05-2007 15-04-2004
US 2005193443 A1	01-09-2005	US 2005193443 A1 US 2009083877 A1	01-09-2005 26-03-2009